

Research article

Open Access

Gene networks driving bovine milk fat synthesis during the lactation cycle

Massimo Bionaz and Juan J Loor*

Address: Mammalian NutriPhysioGenomics, Department of Animal Sciences and Division of Nutritional Sciences, University of Illinois, Urbana, 61801 Illinois, USA

Email: Massimo Bionaz - bionaz@illinois.edu; Juan J Loor* - jlloor@illinois.edu

* Corresponding author

Published: 31 July 2008

Received: 17 January 2008

BMC Genomics 2008, 9:366 doi:10.1186/1471-2164-9-366

Accepted: 31 July 2008

This article is available from: <http://www.biomedcentral.com/1471-2164/9/366>

© 2008 Bionaz and Loor; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: The molecular events associated with regulation of milk fat synthesis in the bovine mammary gland remain largely unknown. Our objective was to study mammary tissue mRNA expression via quantitative PCR of 45 genes associated with lipid synthesis (triacylglycerol and phospholipids) and secretion from the late pre-partum/non-lactating period through the end of subsequent lactation. mRNA expression was coupled with milk fatty acid (FA) composition and calculated indexes of FA desaturation and *de novo* synthesis by the mammary gland.

Results: Marked up-regulation and/or % relative mRNA abundance during lactation were observed for genes associated with mammary FA uptake from blood (*LPL*, *CD36*), intracellular FA trafficking (*FABP3*), long-chain (*ACSL1*) and short-chain (*ACSS2*) intracellular FA activation, *de novo* FA synthesis (*ACACA*, *FASN*), desaturation (*SCD*, *FADS1*), triacylglycerol synthesis (*AGPAT6*, *GPAM*, *LPIN1*), lipid droplet formation (*BTN1A1*, *XDH*), ketone body utilization (*BDHI*), and transcription regulation (*INSIG1*, *PPARG*, *PPARGC1A*). Change in *SREBF1* mRNA expression during lactation, thought to be central for milk fat synthesis regulation, was ≤ 2 -fold in magnitude, while expression of *INSIG1*, which negatively regulates SREBP activation, was > 12 -fold and had a parallel pattern of expression to *PPARGC1A*. Genes involved in phospholipid synthesis had moderate up-regulation in expression and % relative mRNA abundance. The mRNA abundance and up-regulation in expression of *ABCG2* during lactation was markedly high, suggesting a biological role of this gene in milk synthesis/secretion. Weak correlations were observed between both milk FA composition and desaturase indexes (i.e., apparent SCD activity) with mRNA expression pattern of genes measured.

Conclusion: A network of genes participates in coordinating milk fat synthesis and secretion. Results challenge the proposal that *SREBF1* is central for milk fat synthesis regulation and highlight a pivotal role for a concerted action among *PPARG*, *PPARGC1A*, and *INSIG1*. Expression of *SCD*, the most abundant gene measured, appears to be key during milk fat synthesis. The lack of correlation between gene expression and calculated desaturase indexes does not support their use to infer mRNA expression or enzyme activity (e.g., *SCD*). Longitudinal mRNA expression allowed development of transcriptional regulation networks and an updated model of milk fat synthesis regulation.

Background

Progress in lactation biology of the bovine mammary gland advanced substantially during the 20th century (review by [1]). Early studies with ruminants (1960 through 1980s) defined and quantified major metabolic aspects of mammary lipid metabolism, including *de novo* synthesis and fatty acid (FA) uptake from blood [2]. Milk lipid synthesis as well as droplet formation and secretion [3] received particular interest due to their influence on the manufacturing properties and organoleptic quality of milk and dairy products. Recent work has been more focused on qualitative aspects of lipid feeding to manipulate milk FA composition. Milk FA profiles and fat production are affected by stage of lactation and nutrition [4-6]. The latter, however, is by far the predominant environmental factor affecting milk fat production and it represents a practical tool to alter the yield and composition of FA regarded as functional food components (e.g., conjugated linoleic acid and omega-3 fatty acids; [1]). Clearly, deep understanding of mammary physiology and molecular adaptations to diet and/or physiological state are required for efficient manipulation of milk component synthesis and development of dairy products with specific characteristics (e.g., more unsaturated FA, more CLA).

Functional genomics studies highlighted the complexity and coordinated set of molecular events that encompass murine (reviewed in [7]), bovine [8], caprine [9], and porcine [10] mammary adaptations to lactation, revealing new insights about the underlying transcriptomic regulation [11]. Until recently bovine functional genomics studies were not feasible. However, up-to-date bovine genome sequencing and annotation efforts combined with quantitative PCR (qPCR) have become powerful tools for high-precision gene expression analysis. Genetic engineering studies in plants have revealed that an increase in metabolic flux requires manipulation of most of the enzymes in a biosynthetic pathway, challenging the idea of a "limiting enzyme" [12]. Therefore, measurement of mRNA for multiple genes and their networks in a pathway/s is essential to enable conclusions about a metabolic process and its outputs. Previous work in functional genomics also has reinforced the view that transcriptional regulation of gene expression is crucial because it is one of the major long-term regulatory mechanisms of cellular metabolism.

We recognize, however, that mRNA expression is one of multiple factors to be considered when studying the complex molecular networks working simultaneously in tissues. In fact, the ratio between mRNA abundance and abundance of the functional protein coded by the mRNA is hardly 1:1. This has been demonstrated in yeast, especially for the low abundant proteins [13]. There are numerous post-transcriptional and post-translational regulatory steps that preclude from inferring precisely pro-

tein abundance from mRNA. Numerous types of molecular and chemical relationships also exist which directly or indirectly (e.g., protein-protein interaction, phosphorylations) could affect protein activity. The fact remains that post-transcriptional regulation pertains more to short- than long-term regulation [14].

One of the long-term goals in our laboratory is to define gene networks involved in regulating mammary lipid synthesis in dairy cows. As an initial step to characterize these networks and their behavior, we have studied mammary tissue mRNA expression across changes in physiological state. Selected genes included those associated with FA uptake from blood, intracellular FA activation/channeling, *de novo* synthesis, desaturation, regulation of transcription, utilization of ketone bodies, phospholipid and triacylglycerol (TAG) synthesis, lipid globule membrane formation, as well as novel "lipogenic" genes (see Table 1 for details and gene description). Most of the selected genes were chosen based on previous studies with mammary tissue [2,6,15,16]. Others have only recently been discovered and their initial functional characterization conducted in mammary (e.g. *ABCG2* [17]) or other tissues (e.g. *LPIN*, [18]). Specific isoforms for several families of genes involved in TAG synthesis were chosen based on previous published data from our laboratory [19]. The biological effect of changes in gene expression was evaluated via milk fatty acid secretion.

Methods

Animals, sampling, and diet

Holstein dairy cows of high genetic merit were used (Additional file 1, Table S1). Details of the experimental design were reported previously [20]. Briefly, percutaneous biopsies from each of 6 cows were obtained from the right or left rear quarter of the mammary gland at -15 (-13 \pm 3), 1, 15, 30, 60, 120, and 240 d relative to parturition.

RNA extraction, PCR, and design and evaluation of primers

Specific details of these procedures are presented in the Additional file 1 (Supplementary Materials and Methods and Table S2, Table S3, and Table S4).

Data processing and statistical analysis

PCR-normalized data are presented as *n*-fold change relative to -15 d. To estimate standard errors at -15 d, and prevent biases in statistical analysis, normalized data were transformed to obtain a perfect average of 1.0 at -15 d, leaving the proportional difference between the biological replicates. The same proportional change was calculated at all other time points to obtain a fold change relative to -15 d. This final dataset was analyzed using a MIXED model with repeated measures in SAS (SAS Inst. Inc. Cary, NC, release 8.0) to evaluate the effect of time relative to

Table 1: Gene symbol, description, and overall % mRNA abundance among genes investigated

FA import into cells		% RNA ¹	Triacylglycerol synthesis		% RNA
<i>LPL</i>	Lipoprotein lipase	9.56	<i>GPAM</i>	Glycerol-3-phosphate acyltransferase, mitochondrial	2.31
<i>CD36</i>	CD36 molecule (thrombospondin receptor)	4.66	<i>AGPAT6</i>	1-acylglycerol-3-phosphate O-acyltransferase 6	1.28
<i>VLDLR</i>	Very-Low Density Lipoprotein Receptor	0.09	<i>DGAT1</i>	Diacylglycerol acyltransferase 1	0.14
Xenobiotic and Cholesterol transport					
<i>ABCA1</i>	ATP-binding cassette, sub-family A (ABC1), member 1	0.07	<i>DGAT2</i>	Diacylglycerol acyltransferase 2	<0.01
<i>ABCG2</i>	ATP-binding cassette, sub-family G (WHITE), member 2	8.54	<i>LPIN1</i>	Lipin 1	0.13
Acetate and FA activation and intra-cellular transport					
<i>ACSS1</i>	acyl-CoA synthetase short-chain family member 1	0.33	<i>INSIG1</i>	Insulin induced gene 1	0.35
<i>ACSS2</i>	acyl-CoA synthetase short-chain family member 2	0.59	<i>INSIG2</i>	Insulin induced gene 2	0.09
<i>ACSL1</i>	Acyl-CoA synthetase long-chain family member 1	0.89	<i>SCAP</i>	SREBP cleavage activating protein	0.13
<i>ACBP</i>	Acyl-CoA binding protein (diazepam binding inhibitor)	0.17	<i>SREBF1</i>	Sterol regulatory element-binding transcription factor 1	0.15
<i>FABP3</i>	Fatty acid-binding protein, heart	15.49	<i>SREBF2</i>	Sterol regulatory element-binding transcription factor 2	0.10
Fatty acid synthesis and desaturation					
<i>ACACA</i>	Acetyl-coenzyme A carboxylase alpha	0.91	<i>THRSP</i>	Thyroid hormone responsive SPOT14	0.01
<i>FADS1</i>	Fatty acid desaturase 1 (delta-5 desaturase)	0.20	<i>PPARG</i>	Peroxisome proliferator-activated receptor gamma	0.01
<i>FADS2</i>	Fatty acid desaturase 2 (delta-6 desaturase)	<0.01	<i>PPARGC1A</i>	PPAR gamma, coactivator 1 alpha	0.04
<i>FASN</i>	Fatty acid synthase	7.05	<i>PPARGC1B</i>	PPAR gamma, coactivator 1 beta	0.01
<i>SCD</i>	Stearoyl-CoA desaturase (delta-9-desaturase)	23.14	Sphingolipid synthesis		
Lipid droplet formation					
<i>ADFP</i>	Adipose differentiation related protein (adipophilin)	9.56	<i>SPTLC1</i>	Serine palmitoyltransferase, long chain base subunit 1	0.15
<i>BTN1A1</i>	Butyrophilin, subfamily 1, member A1	4.78	<i>SPTLC2</i>	Serine palmitoyltransferase, long chain base subunit 2	0.15
<i>XDH</i>	Xanthine dehydrogenase	7.39	<i>LASS2</i>	LAG1 homolog, ceramide synthase 2	0.61
<i>PLIN</i>	Perilipin	0.01	<i>SPHK2</i>	Sphingosine kinase 2	0.09
Ketone body utilization					
<i>BDHI</i>	3-hydroxybutyrate dehydrogenase, type 1	0.02	<i>ASAHL</i>	N-acylsphingosine amidohydrolase-like	0.05
<i>OXCT1</i>	3-oxoacid CoA transferase 1	0.07	<i>SGPL1</i>	Sphingosine-1-phosphate lyase	0.06
			<i>UGCG</i>	Ceramide glucosyltransferase	0.18
			<i>OSBP</i>	Oxysterol-binding protein 1	0.12
			<i>OSBPL2</i>	Oxysterol binding protein-like 2	0.17
			<i>OSBPL10</i>	Oxysterol binding protein-like 10	0.06

¹ The % mRNA abundance is calculated by $[(1/E^{\Delta Ct}) \text{ specific gene} / \sum (1/E^{\Delta Ct}) \text{ all genes}] \times 100$. See Materials and Methods for details

parturition on gene expression. Compound symmetry was the most appropriate covariate structure used for repeated measures analysis. The model included the fixed effect of time (-15, 1, 15, 30, 60, 120, and 240 d) and the random effect of cow.

Relative mRNA abundance among transcripts

Efficiency of PCR amplification for each gene was calculated using the standard curve method ($E = 10^{-1/\text{log curve slope}}$) (Additional file 1, Table S5). Relative mRNA abundance among measured genes was calculated as previously reported [19], using the inverse of PCR efficiency raised to ΔCt (gene abundance = $1/E^{\Delta Ct}$, where $\Delta Ct = Ct \text{ sample} - \text{geometric mean } Ct \text{ of 3 internal control genes}$). Overall mRNA abundance for each gene among all samples measured was calculated using the median ΔCt . Use of this technique for estimating relative mRNA abundance among genes was necessary because relative mRNA quan-

tification was performed using a standard curve (made from a mixture of RNA from several bovine tissues [20]), which precluded a direct comparison among genes. Together, use of Ct values corrected for the efficiency of amplification plus internal control genes as baseline overcome this limitation. Description of genes measured and overall % relative mRNA abundance are reported in Table 1.

Milk yield, composition and fatty acid analysis

Specific details regarding measurement of milk yield, composition, and fatty acid analysis are presented in the Additional file 1. Daily yield of fatty acids (mole/d) synthesized *de novo* was calculated by the sum of FA 4-14-carbon FA, and yield of FA taken up from blood by the sum of 18-24-carbon FA. The index of acetyl-CoA incorporated during FA elongation (ACE or FA synthesis from acetyl-CoA) was calculated as suggested previously [5] with

modifications (see caption in Additional file 1, Table S6 for details).

Gene network analysis

Gene networks were evaluated using Ingenuity Pathway Analysis® (IPA; <http://www.ingenuity.com>, Redwood City, CA). This is a web-based application that enables the discovery, visualization, and exploration of interaction networks. The software relies on currently known relationships (i.e., published manuscripts) among human, mouse, and rat genes/proteins.

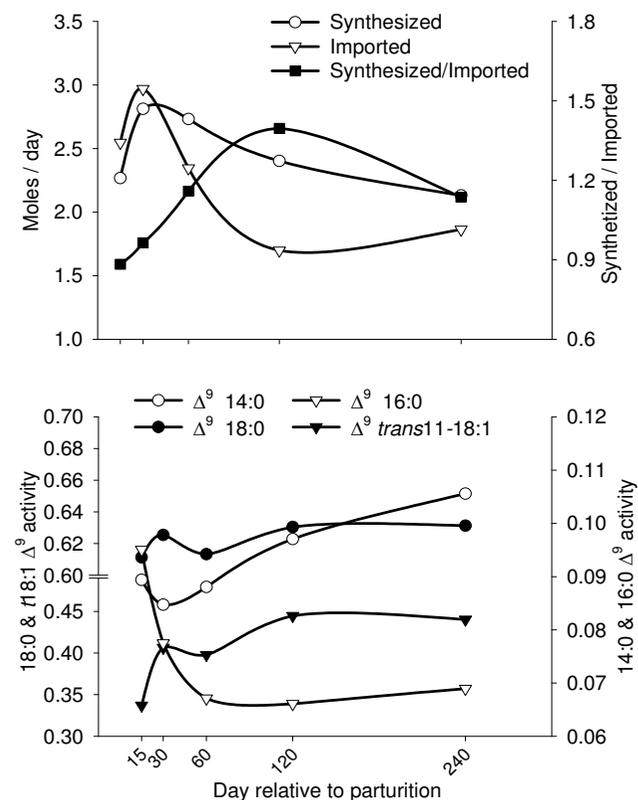


Figure 1

De novo vs. imported FA and Δ^9 desaturase indexes.

De novo FA synthesis (Synthesized) vs FA import (Imported) analysis (Top panel) and Δ^9 desaturase indexes during lactation (Bottom panel). Synthesized = FA with 4÷14 carbons except 11:1; pooled SEM = 0.34. Imported from blood = FA with carbon chain >16 plus 15:0 and 11:1; pooled SEM = 0.31. Synthesized/Imported, pooled SEM = 0.10. Pooled SEM for Δ^9 activity on 14:0, 16:0, 18:0, and trans11-18:1 was 0.008, 0.007, 0.02, and 0.06, respectively. Statistical effect of time: $P < 0.05$ for all measurements except Synthesized FA ($P = 0.24$) and Δ^9 activity on 18:0 ($P = 0.77$).

Results and Discussion

Milk fatty acid composition: a functional analysis

Lactation patterns of FA synthesized vs. FA taken up (Figure 1, top panel) suggest that uptake from blood predominated during the first mo of lactation. Calculation of synthesized FA (Figure 1, top panel) and ACE data (Table 2) suggest that synthesis of FA from acetate or butyrate began during the first 2 wk of lactation and increased rapidly thereafter reaching a peak at 30 d. Thus, we concluded based also on ratio of synthesized/imported (Figure 1, top panel, Table 2) that *de novo* FA synthesis predominated after 1st mo post-partum. The simple sum of FA yield (Σ 4- to 14-carbon FA) to estimate amount of FA originating *de novo* suffers from the assumption that butyrate in milk is completely derived from *de novo* synthesis. Reports indicated that the major part of butyrate (derived from β -hydroxybutyrate) is incorporated directly into *de novo*-synthesized FA (50–60%) [16]. However, a considerable portion also is esterified directly into the *sn*-3 of TAG [2,21]. Pattern of ACE (Table 2) corresponded with pattern of ACACA (acetyl-coenzyme A carboxylase alpha) and FASN (fatty acid synthase) expression (peaked at 60 d, see "Concerted action between *de novo* FA synthesis and desaturation in mammary TAG synthesis" section for details), both of which are key enzymes regulating *de novo* synthesis. Additional discussion on milk FA composition is available in Additional file 1 (Supplementary Results and Discussion).

Fatty acid uptake by mammary cells

LPL and VLDLR and exogenous FA utilization

Mammary cells take up LCFA from albumin-bound fatty acids (NEFA) and lipoproteins. VLDL or chylomicrons are anchored to mammary endothelium by lipoprotein lipase (LPL), which then hydrolyzes TAG in the lipoprotein core to release FA [22]. LPL has higher activity in mammary [22] compared with other tissues, probably due to its high mRNA abundance. The observed up-regulation in LPL mRNA as early as the onset of milk synthesis was remarkable (Figure 2) because mouse mammary tissue had only a 2-fold increase in LPL transcript between pregnancy and lactation. In the mouse, the increase was accompanied by 2-fold up-regulation of enzymatic activity [23,24]. In contrast with murine, bovine mammary LPL expression pattern was remarkably similar to the lactation curve (Additional file 1, Figure S1), which might be indicative of an important role of this gene in maintenance of milk synthesis.

Recent evidence points at VLDL receptor (VLDLR) as an essential component of LPL activity [25]. VLDLR expression was up-regulated throughout lactation, particularly in the first mo post-partum (Figure 2). The mRNA abundance of LPL and VLDLR accounted for ~10% and ~0.1% of total genes measured (Table 1). Despite these differ-

Table 2: % FA from blood, synthesized de novo, and calculated incorporation of acetyl-CoA into de novo synthesized FA (ACE)¹

Item	Day relative to parturition					SEM	P-Value ²
	15	30	60	120	240		
Synthesized FA % ³	46.4 ^a	48.9 ^{ab}	53.5 ^{bc}	57.8 ^c	52.8 ^{abc}	2.08	< 0.01
FA from blood % ⁴	53.6 ^c	51.1 ^{bc}	46.5 ^{ab}	42.2 ^a	47.2 ^{abc}	2.08	< 0.01
ACE mol/d ⁵	5.2 ^a	14.0 ^b	13.8 ^b	12.9 ^b	12.0 ^b	1.46	< 0.01
ACE mol/d ⁶	4.9 ^a	13.7 ^b	13.5 ^b	12.6 ^b	11.7 ^b	1.42	< 0.01
Δ^9 -Desaturase index ⁷	0.35 ^b	0.33 ^b	0.30 ^{ab}	0.27 ^a	0.30 ^{ab}	0.17	< 0.01

¹ The complete FA datasets (mole/day and g/100 g) are reported in Additional file 1 (Tables S6 and S7).

² Effect of day.

³ Fatty acid with carbon length 4-14 (except 11)/tot FA (except 16:0 and 16:1).

⁴ Fatty acid with carbon length >16 (included 11 and 15)/tot FA (except 16:0 and 16:1).

⁵ ACE corrected (Additional file 1, Table S6).

⁶ ACE corrected without considering odd chain FA (11:0, 15:0) (Additional file 1, Table S6).

⁷ Overall Δ^9 desaturase index, calculated from (14:1 c9 + 16:1 c9 + 18:1 c9 + 18:2 c9, t11)/(14:0 + 14:1 c9 + 16:0 + 16:1 c9 + 18:0 + 18:1 c9 + 18:1 t11 + 18:1 c9, t11)

a,b,c denote P < 0.05

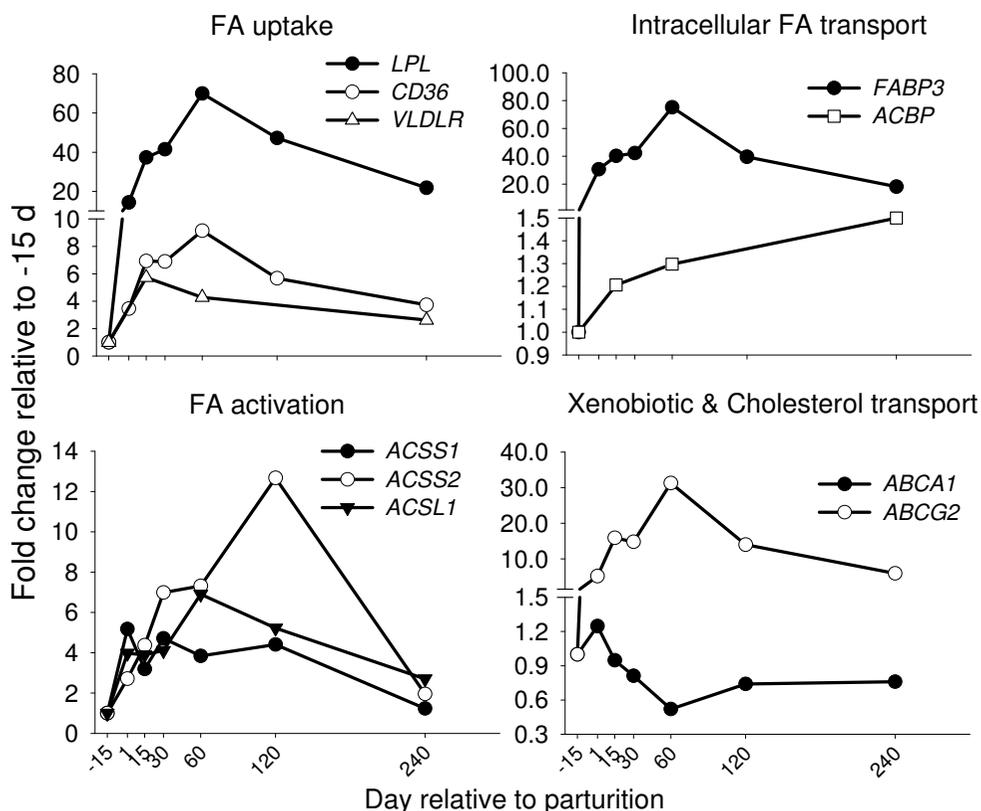


Figure 2

Genes involved in FA uptake, activation, intracellular trafficking, and xenobiotic and cholesterol transport.

Temporal expression patterns in bovine mammary of genes involved in FA uptake (*LPL*, SEM = 8.0; *CD36*, SE = 0.97; *VLDLR*; SEM = 0.72), FA and acyl-CoA transport (*FABP3*, SEM = 6.18; *ACBP*, SE = 0.11), short- and long-chain FA activation (*ACSS1*, SEM = 0.95; *ACSS2*, SEM = 1.66; *ACSL1*, SEM = 0.61), and xenobiotic and cholesterol transport (*ABCA1*, SEM = 0.22; *ABCG2*, SEM = 2.69). Statistical effect of time: P < 0.05 for all genes except *ABCA1* (P = 0.06).

ences, data suggest an important role for VLDLR in concert with LPL [25] in milk fat synthesis during lactation. Mammary VLDLR could act on chylomicrons or intestinal VLDL, which contain apo-B48 [26]. In general, our data are in agreement with previous work reporting higher efficiency of mammary TAG uptake from lipoproteins at the beginning of lactation [26]. The pattern of mammary tissue expression of *LPL* during lactation was in accordance with the typical increase in blood LDL in dairy cows post-partum, which is an indirect index of VLDL utilization [27].

FAT/CD36 and FA internalization

Passive diffusion of FA across membranes plays a minor role compared with protein-mediated FA uptake and the flip-flop mechanism [28]. The main proteins involved in FA uptake in non-ruminant cells include fatty acid translocator FAT/CD36 (*CD36*) and fatty acid transport proteins (FATP or *SLC27A*) [28]. *CD36* mRNA in our study accounted for ~5% of total genes measured (Table 1) and had a large increase in expression (>8-fold) during lactation (Figure 2). This protein is believed to participate in the process of milk fat secretion [3] because of its presence in the milk fat globule membranes (MGFM) [29]. Our data support an important role for CD36 in milk fat synthesis. Although a role for this gene in milk fat secretion cannot be excluded we believe that its involvement in FA import in bovine mammary cells is more important.

We previously showed that bovine mammary tissue expresses most of the known *SLC27A* isoforms, but only expression of *SLC27A6* was up-regulated during the first mo of lactation suggesting a role in NEFA uptake [19]. Up-regulation in expression of *SLC27A6* and *CD36*, and the fact that their proteins co-localize in murine heart subcellular fractions, support the concept of cooperation between both proteins during FA uptake. *CD36* also co-localizes with acyl-CoA synthetases (*ACSL*) and fatty acid binding proteins (*FABP*) [30]. Clearly, FA uptake by bovine mammary cells is a complex and coordinated mechanism requiring evaluation of multiple genes/proteins.

Activation and intracellular channelling of FA

ACSL1 and ACSS2 and FA activation for milk TAG

Long-chain FA (LCFA) are esterified with CoA in the inner face of the plasma membrane prior to participating in metabolic pathways. FA activation occurs primarily via acyl-CoA synthetase long-chain family member isoforms (*ACSL*) [31]. *ACSL1* mRNA is predominant among *ACSL* isoforms in bovine mammary tissue [19], and it increased >4-fold at the onset of lactation suggesting this isoform is important for copious milk fat synthesis (Figure 2). Among enzymes involved in activation of short chain FA (SCFA), acyl-CoA synthetase short-chain family member

2 (*ACSS2*) had greater mRNA abundance and up-regulation in expression than *ACSS1* (a.k.a. *ACAS2L*). mRNA abundance of *ACSS1*, *ACSS2*, and *ACSL1* in each case was <1% of genes investigated (Table 1). Bovine *ACSS* isoforms have been isolated and characterized in tissues other than mammary [32]. In the mouse, *ACSS* isoforms only have 43.8% amino acid similarity and are located in different cell compartments. *ACSS2* (originally named *AceCS1*) is exclusively present in cytosol, while *ACSS1* (originally named *AceCS2*) is primarily found in mitochondria [32]. Both enzymes have high affinity for acetate, with *ACSS2* showing greater affinity than *ACSS1*. The latter also has modest affinity for propionate [32].

Bovine *ACSS1* activated >4-fold more ¹⁴C-acetate into CO₂ than lipid, suggesting it targets acetate towards oxidation [32]. Human *ACSS2* was shown to channel acetate towards FA synthesis [33]. In our study, both *ACSS1* and *ACSS2* mRNA increased substantially during lactation (Figure 2). *ACSS2* transcript pattern corresponded with bovine mammary acetyl-CoA production throughout lactation [34]. Thus, its large increase at the onset of lactation along with the pattern of *ACE* during the first 60 d post-partum, suggest the protein encoded by this gene provides activated acetate for *de novo* FA synthesis. In addition to its use in FA synthesis, acetate is the chief carbon source for energy generation in mammary accounting for ~33% of total CO₂ produced by the tissue [35]. Lower mRNA abundance and pattern of expression of *ACSS1* throughout lactation is in agreement with acetate use for oxidation [2]. Overall, *ACSS* isoforms expression reflected the need for activation of acetate in mammary tissue.

FABP3 and FA trafficking towards milk TAG

Free diffusion of LCFA into cells is too slow to account for the rapid transport and selective targeting towards specific organelles [36], thus, LCFA require specific transporters. Fatty acid binding protein (*FABP*) and acyl-CoA binding protein (*ACBP* or *DBI*) are the main intracellular FA transporters in non-ruminant cells [36]. The former has high affinity for LCFA but also can bind acyl-CoA [37,38]. *ACBP* is the major intracellular transporter of acyl-CoA in several mammalian tissues [39]. We previously observed the presence of mRNA of all *FABP* isoforms, except *FABP2* mRNA, in bovine mammary tissue with greater abundance and up-regulation of *FABP3* mRNA during lactation. Transcript of *FABP4* and *FABP5* also were up-regulated during lactation but were less abundant compared with *FABP3* [19]. In the present study, *FABP3* was the second most abundant transcript (~16%) among all measured, in accord with the large cytosolic content of its protein in mammary epithelium [38]. The large mRNA abundance of this gene also was a consequence of the 80-fold up-regulation during lactation, whereas *ACBP* mRNA abundance was <0.2% among all genes and had a small

increase (1.5-fold) during lactation (Table 1 and Figure 2). Low *ACBP* mRNA abundance agrees with protein abundance data in bovine mammary [40]. Our results suggest a minor role of *ACBP* in bovine mammary lipid synthesis, also supported by murine data [23].

In addition to a trafficking role, FABP3 through binding of activated acyl-CoAs could buffer cells from negative effects of activated FA and prevent inhibition of *ACACA* and *SCD* (stearoyl-CoA desaturase), roles usually attributed to *ACBP* [39]. A positive relationship between FABP and *SCD* has been demonstrated in chickens [41], indicating a coordinated function between both proteins in mammary tissue as also suggested previously from an evaluation of published data [6]. Based on our longitudinal mRNA expression and fatty acid data we propose that an important function of FABP3 in bovine mammary is to provide FA for *SCD*. Large affinity of FABP4 for oleic acid and up-regulation of its mRNA during lactation in bovine mammary tissue [19], led us to propose that FABP3 provides stearoyl-CoA (or other substrates such as 16:0 and *trans*11-18:1) [38] to *SCD* which then releases oleic acid to FABP4. The FA are then available to other enzymes involved in TAG synthesis.

Membrane-associated ATP transporters

ABCG2 and its potential role in milk synthesis

We observed a 30-fold increase in *ABCG2* [ATP-binding cassette, sub-family G (WHITE), member 2] transcript during lactation (Figure 2). mRNA abundance of this gene accounted for ~9% of total genes measured (Table 1). *ABCG2* is a member of the large ATP binding cassette family of membrane-spanning efflux pumps that actively extrude a wide range of xenobiotics [42]. It is present in apical membrane of murine mammary alveolar epithelia and plays a role in active secretion of toxins into milk [42]. It also is present in the milk fat globule membrane (MFGM) [29], probably in the external bilayer originating from plasma membrane, due to its apical membrane localization. The large *ABCG2* mRNA abundance and up-regulation, both in lactating bovine and murine mammary tissue [17,23], is biologically puzzling because the primary role of this transporter in other tissues is detoxification. Recently, it was reported that one amino acid substitution at position 581 (Tyr to Ser; Y581S) of *ABCG2* resulted in decreased milk production but increased milk fat and protein concentration and yield [43]. Those data clearly do not support a role of *ABCG2* in synthesis or secretion of milk fat. What seems apparent based on current bovine data is that *ABCG2* plays an essential role in secretion of "some" important milk constituent [42]. Cholesterol transport was suggested [43] but it is not supported by the low amount [3] and pattern of cholesterol in bovine milk throughout lactation [44]. The only demonstrated role of *ABCG2* in secretion of a milk component

is for riboflavin, an essential, but quantitatively marginal, nutrient for the neonate [17]. Therefore, its large up-regulation is suggestive of other functions in milk synthesis besides riboflavin secretion.

The amount and pattern of cholesterol secretion into bovine milk agrees with the pattern of *ABCA1* (ATP-binding cassette, sub-family A, member 1) transcript we observed (Figure 2). *ABCA1* mRNA accounted for <1% of all genes measured (Table 1). This gene also has low expression in a number of other bovine tissues, and bovine mammary in particular [45]. *ABCA1* is crucial for efflux of cholesterol from cells [46]. Taken together, data suggest a minor role of *ABCA1* in bovine mammary cholesterol flux.

Concerted action between de novo FA synthesis and desaturation in mammary TAG synthesis

ACACA, FASN, and de novo FA synthesis

Production of SCFA and palmitate from acetate is under control of *ACACA*, considered the rate-limiting step in *de novo* FA synthesis [2]. In subsequent steps, both, acetyl-CoA and butyryl-CoA (mostly from plasma β -hydroxybutyrate) are primers for the cytosolic multifunctional protein fatty acid synthase (*FASN*) [16]. The major product of *FASN* is palmitate but in ruminants the enzyme also produces SCFA [16]. In the present study, *ACACA* mRNA abundance accounted for <1% whereas *FASN* accounted for 7% of total genes measured (Table 1). However, *ACACA* mRNA had greater up-regulation during lactation compared with *FASN* (Figure 3). These data are consistent with activity values of the two enzymes from pregnancy through lactation in dairy cows [34]. Despite differences in magnitude, expression patterns among both genes were similar ($r = 0.90$; $P < 0.01$; see Excel File in Additional file 2), confirming previous findings summarized in a recent review article [6]. In fact, this was the case for several genes involved in TAG synthesis and transport. Clearly, bovine mammary lipid synthesis requires coordinate expression of several genes for TAG synthesis and secretion. This point was stressed in studies of rabbit mammary lipid metabolism several decades ago [47]. However, the scope of enzymes studied previously was relatively small compared with our study.

Stearoyl-CoA desaturase and milk TAG synthesis

Only a fraction of FA taken up by the mammary gland is unsaturated owing to extensive ruminal biohydrogenation. The primary enzyme involved in monounsaturated FA synthesis is stearoyl-CoA desaturase (*SCD*), which introduces a double bond in the Δ^9 position of myristoyl-, palmitoyl-, and stearoyl-CoA, primarily [48]. To date, two *SCD* isoforms have been identified and characterized in bovine: *SCD1* and *SCD5*. *SCD1* was first characterized in bovine adipose [49] and until the discovery of *SCD5*, it

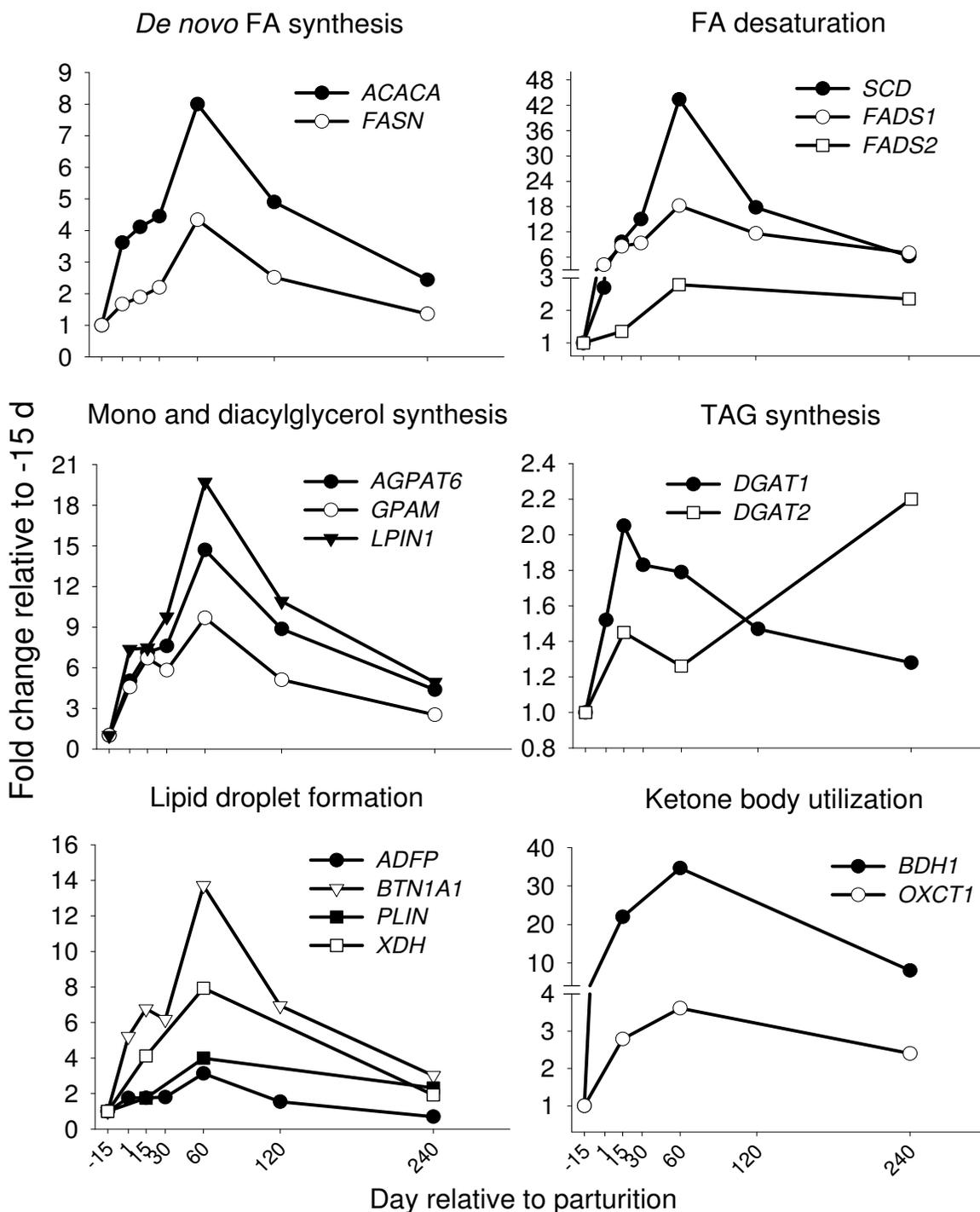


Figure 3
Genes involved in de novo FA synthesis, LCFA desaturation, TAG synthesis, lipid droplet formation, and BHBA utilization. Temporal expression patterns in bovine mammary of genes involved in de novo FA synthesis (*ACACA*, SEM = 0.62; *FASN*, SEM = 0.33), long-chain FA desaturation (*SCD*, SEM = 6.20; *FADS1*, SEM = 1.74; *FADS2*, SEM = 0.36), TAG synthesis (*GPAM*, SEM = 0.96; *AGPAT6*, SEM = 1.24; *LPIN1*, SEM = 3.9; *DGAT1*, SEM = 0.16; *DGAT2*, SEM = 0.86), lipid droplet formation (*ADFP*, SE = 0.23; *BTN1A1*, SEM = 1.10; *PLIN*, SEM = 1.29; *XDH*, SEM = 0.90), and ketone body utilization (*BDH1*, SEM = 8.60; *OXCT1*, SEM = 0.54). Statistical effect of time: $P < 0.05$ for all genes except *PLIN* ($P = 0.17$) and *DGAT2* ($P = 0.54$).

was believed to be the only *SCD* present in this species. Thus, the bovine gene is simply referred to as *SCD*. Bovine *SCD5* has been identified and characterized only recently [50] and it is expressed almost exclusively in brain. The nucleotide sequence for a bovine *SCD6* homolog A (*S. cerevisiae*) has been deposited at NCBI [51], but no characterization of this isoform has been performed to date. *SCD1* has 33.5% and 31.3% global nucleotide alignment identity [52] with *SCD5* and *SCD6*, respectively. The primer pair used in the present study (Additional file 1, Table S2) is specific only for *SCD1*. The *SCD* mRNA abundance was the highest (23%; Table 1) among all genes measured. The large *SCD* mRNA abundance, relative to other classical lipogenic genes (e.g., *ACACA*, *FASN*), and the >40-fold up-regulation during lactation (Figure 3) agrees with the suggestion by Kinsella, based on lactating mammary *SCD* activity [53], that it plays a crucial role in TAG synthesis.

All indexes of "apparent" desaturase activity increased during lactation except for stearic acid and palmitic acid desaturation. The latter index had a peak at the beginning of lactation followed by a dramatic decrease (Figure 1, bottom panel). The pattern of *SCD* mRNA was not significantly correlated with any of the Δ^9 desaturase indexes (Additional file 2), and was nearly opposite to the overall Δ^9 desaturase index (Table 2). A lack of correlation between desaturase indexes and *SCD* desaturase activity was found previously in bovine intramuscular fat [54], which along with our findings suggests that use of indexes is inappropriate for inferring *SCD* gene expression/activity at least when considering the lactation cycle [4]. Previous single-time point studies observed a positive correlation between *SCD* mRNA in goat mammary tissue and milk fat oleic acid [55] or desaturase indexes [56]. In our study, relative % mRNA abundance of *SCD* (23%; Table 1) was related to the total amount of Δ^9 FA (14:1c9 + 16:1c9 + 18:1c9 + *cis*9, *trans*11-18:1), which accounted for 19% of total milk FA (molar proportion; Additional file 1, Table S6).

Overall, our results support a central function of *SCD* in milk fat synthesis as previously demonstrated by intravenous infusions of sterculic acid, an inhibitor of *SCD* activity [57]. This point is supported, to some extent, by the recent discovery that both *SCD* and *DGAT1* polymorphisms affect saturation level of milk fat and largely explain genetic variance on the desaturase indexes. The effect, however, was greater for *SCD* and primarily on medium/long-chain FA (from 10- to 16-carbon FA). Polymorphism in *DGAT1* explained less variability but had a greater effect on long-chain FA (18-carbon) [58].

The lack of correlation between temporal desaturase indexes (i.e., apparent *SCD* activity) and *SCD* mRNA

expression is not surprising due to the many factors that likely play a role in determining milk FA output. An important factor to consider is the selective uptake of stearic acid from blood VLDL by the mammary gland [59]. Mammary tissue relies heavily on utilization of VLDL-TAG during lactation [27]. Another key aspect is the high concentration of oleic acid, both in plasma at mid-lactation [60] and the NEFA pool at early lactation [61]. Oleic acid is the predominant FA in ER membranes [62] and Δ^9 desaturase is essential for their functional maintenance [48]. *FABP3* expression and activity also could play a role in preferential channelling of palmitic and stearic acid for desaturation. Bovine *FABP3* has a high affinity for both stearic and palmitic acid [38,63].

Very long-chain fatty acid desaturases

Synthesis of very-long-chain FA is carried out by fatty acid desaturase 1 (*FADS1*) and 2 (*FADS2*), which add double bonds at the Δ^5 and Δ^6 position of PUFA. Arachidonic acid (20:4n-6), eicosapentaenoic acid (20:5n-3), and docosahexaenoic acid (22:6n-3) are synthesized via *FADS1* and *FADS2* [64,65]. *FADS1* and *FADS2* mRNA abundance was <1% of total genes measured (Table 1). The greater relative mRNA abundance of *FADS1* compared with *FADS2* is similar to the one reported in rat mammary [65]. Relative mRNA abundance in bovine mammary tissue was in concordance with the amount of product of both enzymes, which accounted only for 0.12% and 0.03% of total milk FA (Additional file 1, Table S6). *FADS1* mRNA expression increased 18-fold by d 60 postpartum, whereas *FADS2* mRNA increased only 3-fold (Figure 3). Expression pattern of *FADS1* mRNA did not agree with the observed Δ^5 desaturase index, whereas *FADS2* mRNA had a similar pattern compared with its index of activity (Additional file 1, Figure S2). In practical terms, it appears that *FADS1* mRNA abundance/activity might be more amenable to dietary manipulation in order to increase omega-3/omega-6 ratio in milk fat.

Formation of TAG and milk lipid droplets

Role for *AGPAT6* and *LPIN1* in TAG synthesis

Discrete steps in the pathway of TAG synthesis [66] have been discerned in classical lipogenic tissues (e.g., liver, adipose; [67]) and, despite lack of functional studies of mammary lipin, the same steps likely apply to the mammary gland. Expression of *GPAM* (glycerol-3-phosphate acyltransferase, mitochondrial), *AGPAT6* (1-acylglycerol-3-phosphate O-acyltransferase 6), *DGAT1* (diacylglycerol acyltransferase 1), and *LPIN1* (lipin 1) mRNA accounted for >2%, >1%, ~0.1%, and ~0.1%, respectively, of total transcripts measured (Table 1). *DGAT2* mRNA expression was nearly undetectable. Despite these differences, we observed that *LPIN1* mRNA was up-regulated during lactation by 20-fold (Figure 3). The more abundant *GPAM* and *AGPAT6* mRNA increased by 10- and 15-fold by d 60

post-partum (Figure 3). *GPAM* expression agrees with the greater enzyme activity in mammary gland during lactation in non-ruminants (e.g., [47]), and confirms its crucial role in TAG synthesis [16]. *AGPAT6* and *LPIN1* are the major isoforms within each gene family in bovine mammary tissue [19]. When the former was knocked out in lactating mice, they failed to synthesize milk fat [68].

Recent characterization of *AGPAT6* showed that this gene is in fact the ortholog of human *GPAT4* (microsomal glycerol-3-phosphate acyltransferase 4) [69]. The product of *GPAT4* did not have *AGPAT* activity but instead a clear glycerol-3-phosphate acyltransferase activity [69]. However, authors failed to demonstrate an increase in TAG synthesis after overexpression of *GPAT4*. Despite these results in the mouse, our data support an important role of *AGPAT6* in bovine mammary and very likely in TAG synthesis, as previously discussed [19]. Our data seem to suggest that *AGPAT6* has *AGPAT* activity. This is inferred by the lower mRNA abundance and temporal increase in the transcript of other *AGPAT* in bovine mammary tissue [19], as well as the large up-regulation of *GPAM* expression. It seems unlikely that bovine mammary tissue would require a larger number of enzymes with *GPAT* activity relative to other downstream FA acylating enzymes.

A potential function of *LPIN1* in regulation of transcription of other genes involved in milk fat synthesis cannot be excluded. Recently, it was demonstrated that *LPIN1* is essential for *PPAR α* [70] activation but it also interacts with *PPAR γ* [71]. In addition, *LPIN1* is a target of insulin-stimulated phosphorylation through *mTOR* [72], which in turn seems to promote microsomal vs. cytosolic localization of the protein. It could be possible that in bovine mammary tissue insulin signalling through *INSR* (insulin receptor) and *IRS1* (insulin receptor substrate-1) as well as *mTOR* (*FRAP1*), all of which had a significant increase in mRNA expression (1.5-4-fold) through lactation [8], induces *LPIN1* phosphorylation and localization to ER for DAG synthesis and TAG formation.

Relative mRNA abundance of *DGAT1* was 17-fold greater compared with *DGAT2* (Table 1) and had modest up-regulation particularly in early lactation (Figure 3). The temporal pattern in expression of this gene was similar to butyrate yield (Additional file 1, Table S7). *DGAT1* acylates the *sn*-3 position of DAG and most butyrate in milk TAG is found here [44]. This protein has high affinity for butyryl-CoA and even higher for palmitoyl-CoA [73]. However, *DGAT1* might favour use of butyrate for the *sn*-3 position of DAG as indicated by the larger affinity of *AGPAT* for LCFA in mammary [74] along with the preferential incorporation of palmitic acid in the *sn*-1 and *sn*-2 position [3].

The pattern of *DGAT1* expression was unexpected because it is considered a QTL [75] for milk production traits, and is essential for murine mammary gland development and milk synthesis [76]. Data from the present study (i.e., fold-change and mRNA abundance) suggest that *DGAT1*, compared with other genes involved in TAG synthesis, is of minor importance in the overall process of milk fat synthesis. The temporal decrease or lack of increase in expression of *Dgat1* in mammary tissue of FVB mouse [23] provides additional support. We do not, however, believe our findings contradict previous functional studies [77], demonstrating a pivotal role for *DGAT1* in increasing milk TAG. The fact remains that *DGAT1* is one of many proteins composing the TAG synthesis pathway [67]. A lack in functionality of any gene in this pathway can likely reduce the efficiency of TAG synthesis. Protein expression and functional studies during the entire lactation should be conducted to clarify the importance of *DGAT*, and others, in mammary lipid synthesis.

Milk lipid droplet formation

Milk fat globules are formed in the ER membrane via incorporation of newly-formed TAG, transported to the apical membrane, and eventually released [3]. Well-defined proteins involved in these processes in mammary include butyrophilin (*BTN1A1*), xanthine dehydrogenase (*XDH*), and adipophilin (*ADFP*) [3,15,29]. Relative mRNA abundance among these genes confirms the large amount of the respective protein product found in MFGM [3]. *ADFP* mRNA abundance (~10%) relative to other genes measured was almost twice that of *BTN1A1* (~5%) (Table 1). Expression of *BTN1A1*, *XDH*, and *ADFP* increased during lactation and averaged 14-, 8-, and 3-fold by d 60 postpartum, respectively (Figure 3). The larger increase in expression of *BTN1A1* seems to support a more crucial role for this gene, as recently suggested [78], in milk fat secretion compared with *XDH* and *ADFP*. However, despite the greater increase in mRNA expression for *BTN1A1* than *ADFP* at 60 d post-partum relative to pre-partum, the larger overall abundance of *ADFP* transcript resulted in similar mRNA abundance for both genes at 60 d (data not shown). The similar proportion in relative mRNA of *BTN1A1* and *ADFP* is in accordance with their protein abundance in mammary tissue [3]. Our results highlight the limitations of reporting gene expression data exclusively as *n*-fold change. Relative mRNA abundance also needs to be considered. The precise mechanism of milk fat secretion is still debated (e.g. [15,78]) but our data confirmed a role for *BTN1A1*, *XDH*, and *ADFP*. The similar pattern of expression and large correlation observed ($r \geq 0.92$, $P < 0.01$; Supplemental Excel file) are indicative of a concerted action among the 3 genes and, thus, provides support for the tripartite model of murine milk lipid secretion [15].

Perilipins are a family of proteins localized in the periphery of intracellular lipid droplets and are essential for droplet formation as well as lipolysis in adipose tissue [79]. *ADFP* and the perilipin gene (*PLIN*) are both part of the perilipin family. Relative *PLIN* mRNA abundance was ~0.01% of total genes measured (Table 1) and its expression was only numerically up-regulated (~3-fold by d 60) during lactation. Our data do not support a significant role for *PLIN* in mammary lipid droplet formation. Functional studies could clarify the involvement, if any, of *PLIN* in this process.

BDH1 and *OXCT1* and utilization of ketone bodies by mammary β -hydroxybutyrate (BHBA) is the major ketone body produced in bovine species under most circumstances. Bovine mammary gland takes up large amounts of BHBA from blood [80]. Previous studies, focused mostly on *de novo* synthesis and oxidation of FA, concluded that use of BHBA (as 4-carbon units) by mammary cells is primarily for *de novo* FA synthesis. A minor portion is used as energy source through the TCA cycle, leaving unaccounted the fate of a substantial portion of the BHBA taken up [80]. *BDH1* (3-hydroxybutyrate dehydrogenase, type 1) and *OXCT1* (3-oxoacid CoA transferase 1) catalyze the initial and committed steps of BHBA utilization in mitochondria [81]. mRNA abundance for *BDH1* and *OXCT1* accounted for <0.1% of genes measured (Table 1). The large increase in expression and relative mRNA abundance during lactation (Figure 3; Table 1) correspond with their enzymatic activity level in lactating rat mammary tissue [82]. Based on these data, and a previous microarray study [8], we propose that the major fate of BHBA in bovine mammary is the synthesis of citrate (Additional file 1, Figure S3 for model and details).

Transcription factors and nuclear receptors during lactation: potential roles in mammary lipid metabolism *SREBF-related networks and TAG synthesis*

A large body of evidence supports the suggestion that SREBP1 (sterol regulatory element-binding protein 1) is pivotal in the regulation of milk fat synthesis in mouse [7] and cow [83]. SREBP1 and 2 reside as inactive precursors in the ER membrane and are transported to the Golgi for proteolytic cleavage (i.e. activation) prior to entering the nucleus and activation of sterol responsive element-containing genes (e.g., *ACACA*, *FASN*). The transport step to the Golgi is blocked by sterols via the sterol-sensing protein SCAP (SREBP cleavage activating protein). SCAP is essential for the movement of SREBP isoforms from the ER to the Golgi, essentially acting as gate keeper for movement of inactive SREBP1 and 2 [84]. Insulin induced gene (*INSIG*) 1 and 2 are proteins that interact with SCAP in an oxysterol-dependent and independent fashion and regulate the responsiveness of SREBP1 and 2 processing via SCAP, thus altering rates of lipogenesis [84].

Expression of SREBP genes (*SREBF1* and *SREBF2*), and thyroid hormone responsive *SPOT14* (*THRSP*) averaged ~2-fold by day 60 postpartum (Figure 4) but relative mRNA abundance was ~0.13% for *SREBF1* and *SREBF2*, and only 0.01% of total genes measured for *THRSP* (Table 1). There are two isoforms of SREBP1 (a and c) that can be expressed at different levels in tissues. The two isoforms differ by only 84 nucleotides at the first exon [85] and each appears to be specific in the control of transcription of genes involved in cholesterol (isoform a) or TAG synthesis (isoform c) [86]. The primer pair used in our study is unable to differentiate between these because it amplifies mRNA at the 14th exon.

INSIG1, *INSIG2*, and *SCAP* mRNA abundance accounted for ~0.4%, ~0.1%, and ~0.1%, respectively, of total genes studied (Table 1). These genes had increased mRNA expression during lactation, ranging from ~1.5-fold for *SCAP* to ~12-fold for *INSIG1* at or close to peak lactation (Figure 4). It is apparent from our comprehensive analysis of *SREBF1*, in concert with its co-factors (*SCAP*, *INSIG1* and 2), that the complete activation program of SREBP is up-regulated in the bovine mammary gland during lactation. More *INSIG1* mRNA was expressed in mammary tissue than *SCAP* or *SREBF1* and 2 (Table 1), and its level of up-regulation reached 10-12-fold between peak through mid-lactation (Figure 4). If this pattern of *INSIG1* mRNA expression extended to the protein level, more SREBP might be retained in the ER via SCAP-*INSIG1* binding (i.e., rendered inactive) [84].

INSIG1 binds oxysterols (not cholesterol) specifically and this specificity is directly correlated with the ability of these compounds to inhibit SREBP cleavage [84]. Data on oxysterol concentration/synthesis in bovine mammary tissue are not readily available but considering that mammary tissue uptake and synthesis of cholesterol is small we can infer low presence of these compounds. In this regard, it is interesting that genes coding for oxysterol binding proteins were up-regulated during lactation (discussed below). Cleavage inhibition of SREBP also can occur in the absence of sterols, particularly when the ratio of *INSIG1*/*SCAP* increases as in the present study [84]. Therefore, it appears that an increase of *INSIG1* alone can be enough to block SREBP cleavage. In fact, decreased SREBP activity as a consequence of increased *INSIG1* transcript has been observed in liver when *INSIG1* is overexpressed [87] or during high fat diet-induced *INSIG1* up-regulation [88]. Based on previous results and our data we postulate that mammary lipid synthesis cannot rely solely on transcriptional regulation via SREBP1 which is probably inhibited or controlled by *INSIG1*.

The observed up-regulation of *INSIG1* during lactation apparently does not make teleological sense because

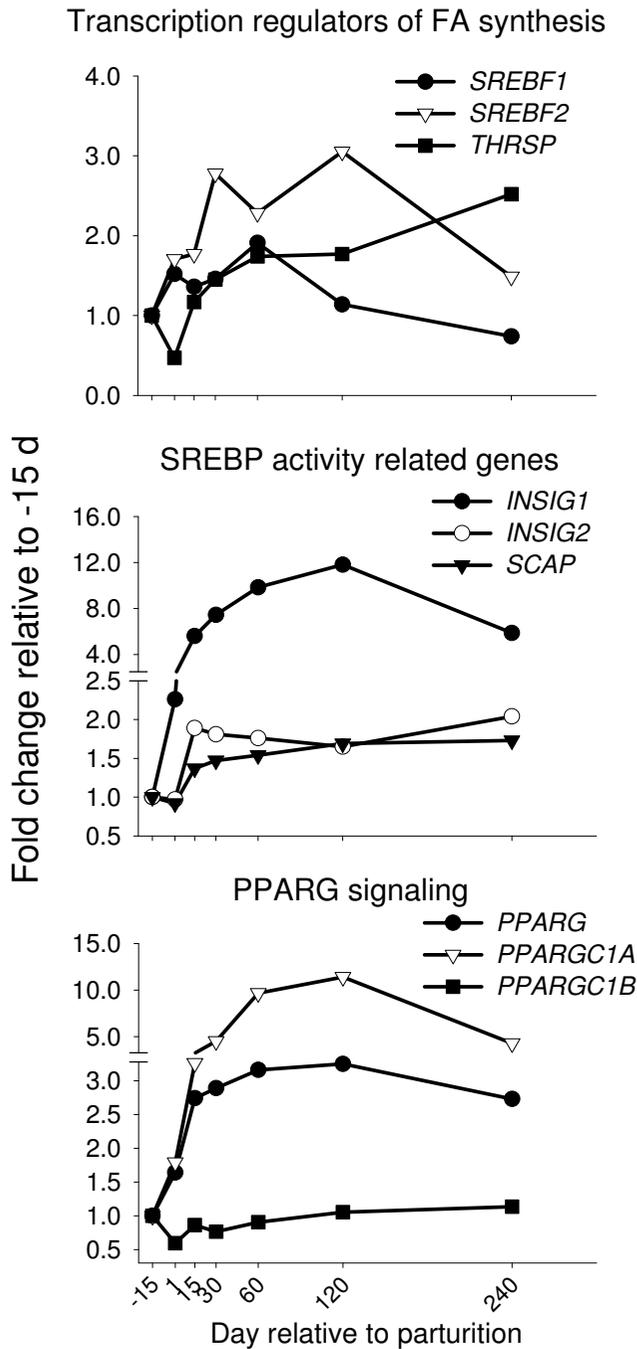


Figure 4
Regulation of transcription in bovine mammary. Temporal expression patterns in bovine mammary of genes involved in regulation of transcription (*SREBF1*, SEM = 0.18; *SREBF2*, SEM = 0.36; *THRSP*, SEM = 0.49; *INSIG1*, SEM = 1.30; *INSIG2*, SEM = 0.18; *SCAP*, SEM = 0.08; *PPARG*, SEM = 0.43; *PPARGC1A*, SEM = 6.6; *PPARGC1B*, SEM = 0.09). Statistical effect of time: $P < 0.05$ for all genes

mammary tissue requires substantial up-regulation of genes involved in *de novo* lipid synthesis (Figure 1; Table 2). It is striking, however, that *INSIG1* mRNA was at its peak (d 120) when the cows likely were in positive energy balance, i.e. high blood insulin/glucagon, and the ratio of *de novo*-synthesized FA to imported FA was maximal (Figure 1). *INSIG1* expression was positively correlated with the ratio of synthesized/imported FA ($r = 0.38$, $P < 0.05$; Additional file 2). Paradoxically, these data suggest some involvement of *INSIG1* in inducing FA synthesis. This suggestion is supported by recent data [83], where a decrease of *INSIG1* during experimentally-induced milk fat depression in lactating cows was observed.

Several factors could drive the marked increase in *INSIG1* mRNA during lactation. For example, up-regulation of *INSIG1* expression might be a consequence of *SREBF* isoform mRNA up-regulation and increased activity (i.e., induction of gene expression) of the corresponding proteins. *SREBP1a* and *SREBP2* directly regulate *INSIG1* gene expression [84]. Given the lipogenic capacity of mammary tissue, it is more likely that *SREBP1c* is the more abundant isoform. Thus, *INSIG1* up-regulation in bovine mammary tissue could be under control of *SREBF2* (Figure 4). Another reason for marked *INSIG1* mRNA up-regulation might be its very short half-life [84], or as a necessary mechanism to sense low mammary cholesterol levels in order to regulate *de novo* FA synthesis.

Our data support a need of *INSIG1* in controlling the induction of gene expression by *SREBF* isoforms. Therefore, *INSIG1* could play a central role in orchestrating lipid metabolism (i.e., TAG vs. cholesterol) in bovine mammary tissue during lactation. In support of this, it previously has been suggested that high levels of *INSIG1* create a situation in which low levels of endogenous sterols can trigger *SCAP* binding to *INSIG1* without the necessity for exogenous sterols [84]. The "brake" effect of *INSIG1* on TAG accumulation has been clearly demonstrated in mouse adipose tissue [88], when overexpression of *INSIG1* in 3T3-L1 cells led to a decrease in mRNA abundance of lipogenic genes (e.g. *Srebp1c*, *Acaca*, *Chrebp*, *Pparg*). Our data, however, do not support a similar effect of *INSIG1* on expression of mammary lipogenic genes in ruminant.

SREBF2 expression was up-regulated to a greater extent than *SREBF1* expression during lactation (Figure 4). The reason for up-regulation of *SREBF2* expression, and the similar mRNA abundance compared with *SREBF1*, is not apparent because this gene is thought to be involved primarily in cholesterol biosynthesis [85] and the amount of cholesterol in milk is low [89]. Cholesterol is almost exclusively synthesized *de novo* in bovine mammary tissue [90], thus *SREBF2* expression also might be necessary to

meet cholesterol requirements for MFGM formation [3]. In support of this we observed, via microarray analysis, that expression of several genes involved in cholesterol synthesis was significantly up-regulated (1.5-2-fold compare to -30 d) during lactation [8]. An additional function of SREBP2 is to induce mRNA expression of genes involved in FA synthesis [85]. It could be possible that *SREBF2* mRNA up-regulation during lactation might compensate for the potential inhibition of *INSIG1* on both SREBP.

Nuclear receptors and the lipogenic program

Genes involved in FA transport such as *LPL*, *CD36*, and *ACSL1* are peroxisome proliferator-activated receptor gamma (PPAR γ) target genes [91]. In the lactating mouse, expression of *LPL* and *ACSL1* was up-regulated significantly despite the fact that expression of PPAR γ was down-regulated [23]. However, it has recently been demonstrated that changes in abundance of adipocytes at several stages of pregnancy in murine mammary tissue (i.e., high in early pregnancy vs. low in late pregnancy) could account for the decrease in PPAR γ gene (*PPARG*) mRNA abundance [92]. In bovine mammary, *PPARG* mRNA accounted only for 0.01% of total genes measured (Table 1) but was consistently up-regulated during lactation (Figure 4). The low mRNA abundance of "adipocyte-specific" genes (e.g., *DGAT2*, *PPARG*, *ACBP*, and *PLIN*), particularly at the end of pregnancy (i.e. -15 d), clearly indicates that biopsied cow mammary tissue contained low amounts of adipocytes. Thus, our longitudinal data on *PPARG* expression should represent that of epithelial cells. With this premise and despite the low mRNA abundance, up-regulation of *PPARG* mRNA during lactation suggests a potential role of this nuclear receptor in milk fat synthesis. A recent study with PPAR γ -knockout mice indicated that absence of *PPARG* increased utilization of FA for synthesis of inflammatory lipids due to reduced TAG synthesis [93]. A role of *PPARG* in regulating the entire bovine milk fat synthesis machinery also is supported by recent results from our laboratory where treatment of MacT cells (immortalized bovine mammary epithelial cells) with rosiglitazone, a specific PPAR γ agonist, resulted in coordinated up-regulation in expression of genes involved in FA import (e.g., *CD36*), *de novo* FA synthesis (e.g., *ACACA*, *FASN*, *SREBF1*), and TAG synthesis (e.g., *LPIN1*, *SCD*) [94].

The relative mRNA abundance for the PPAR gamma coactivators, *PPARGC1A* and *PPARGC1B*, was 0.04% and 0.01% of total genes measured (Table 1). Whereas expression of *PPARGC1A* was substantially up-regulated through d 120 post-partum (~11-fold), expression of *PPARGC1B* was consistently down-regulated during the entire lactation (Figure 4). Differences in relative mRNA abundance, large temporal up-regulation of *PPARGC1A*

mRNA, and down-regulation of *PPARGC1B* transcript, suggest an important role of the former in bovine milk fat synthesis. The importance of *PPARGC1A* in the overall process of mammary lipid synthesis likely is more related to its well-defined role in regulating mitochondrial biogenesis and energy metabolism [95]. Up-regulation of *PPARGC1A* during lactation agrees with reported increases in numbers and turn-over rate of mitochondria in lactating mammary tissue [21]. Furthermore, our combined data (Table 1, Figure 4) also point to a concerted action of *PPARGC1A* and *INSIG1*. This last observation is captivating based on previous observations in murine [96] and bovine mammary epithelial cells [94] where *INSIG1* was demonstrated to be a PPAR γ responsive gene, suggesting that PPARG in mammary tissue could serve as regulator of SREBP activity.

Ceramide-synthesis genes in bovine mammary tissue

Synthesis of Ceramide in bovine mammary

Ceramide, which is involved in cell signaling, cell cycle, and regulation of protein transport from ER to Golgi, is one of the most studied sphingolipids in nature [97,98]. Other sphingolipids with signaling roles include sphingosine (Sph) and sphingosine-1-phosphate (S1P) [98]. Sphingomyelin synthesis from ceramide is important for milk quality because this compound is considered a functional food component [16]. Although minor compared with TAG, sphingolipids are the third most important lipid component [99] in bovine milk fat. MFGM formation relies on sphingolipid and cholesterol availability [100], thus coordinated synthesis of both compounds is pivotal to milk lipid droplet formation/secretion. Sphingolipids are involved in lipid synthesis regulation through their action on SREBP [101]. Mammary tissue synthesizes sphingolipids *de novo* [99] from palmitoyl-CoA, leading to ceramide formation and incorporation into sphingomyelin [99]. Thus, palmitic acid used for ceramide synthesis in mammary appears a required step and also might represent a regulatory point for FA synthesis because ceramides can inhibit this process by blocking the activity of AKT/PKB [102]. Regulation of FA synthesis by sphingolipids in mammary tissue has never been investigated. Thus, we selected genes crucial in ceramide synthesis/degradation, as well as enzymes involved in Sph and S1P synthesis (Additional file 1, Figure S5) to explore further their role in lipid synthesis regulation.

In accordance with the minor concentration of sphingolipids in milk, genes in this pathway had low mRNA abundance ranging from 0.05% (N-acylsphingosine amidohydrolase-like or *AS AHL*) to 0.61% (*LAG1* homolog, ceramide synthase 2 or *LASS2*) of total genes examined (Table 1). *LASS2* was the most abundant among sphingolipid-related genes and the only one with >2-fold up-regulation during lactation (Figure 5). *LASS*

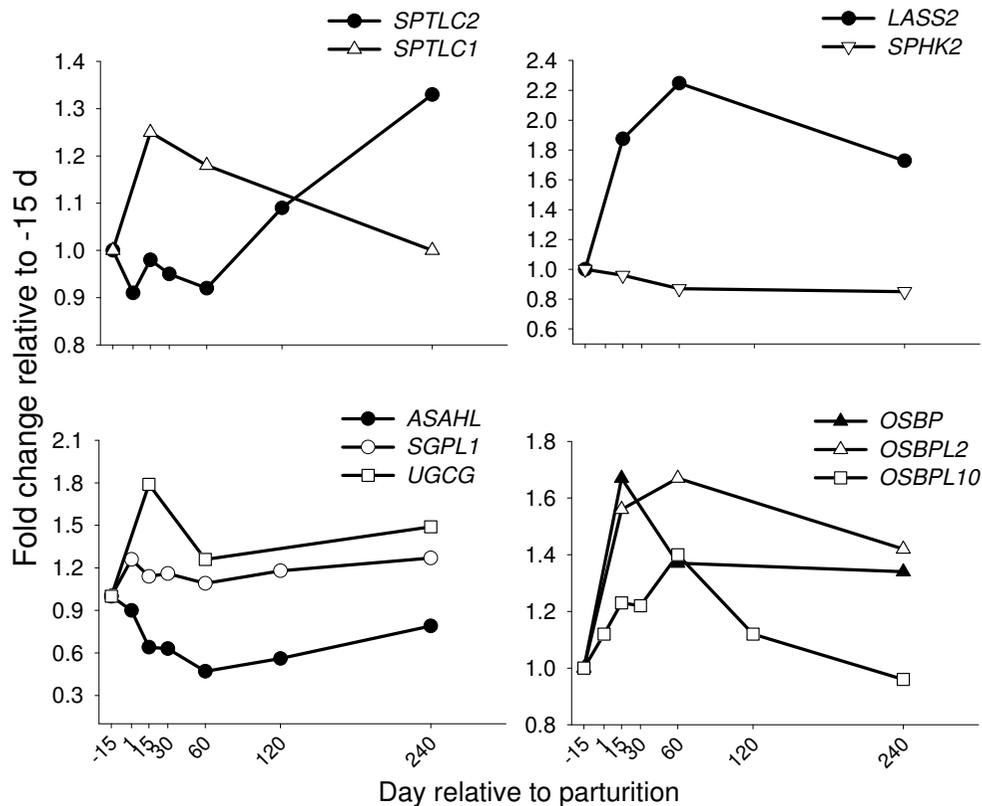


Figure 5

Genes involved in sphingolipid synthesis in bovine mammary. Temporal expression patterns in bovine mammary of genes involved in sphingolipid synthesis (*SPTLC1*, SEM = 0.10; *SPTLC2*, SEM = 0.08; *LASS2*, SEM = 0.14; *SPHK2*, SEM = 0.10; *ASAHL*, SEM = 0.10; *SGPL1*, SEM = 0.13; *UGCG*, SEM = 0.17; *OSBP*, SEM = 0.13; *OSBPL2*, SEM = 0.10; *OSBPL10*, SEM = 0.09). Statistical effect of time: $P < 0.05$ for all genes except *SGPL1* ($P = 0.63$) and *SPHK2* ($P = 0.65$).

isoforms are orthologues of the yeast Longevity-assurance gene. The enzyme is localized in the ER [97] and isoforms appear to have specific tissue distribution, suggesting they perform "specialized" functions [19,97]. *LASS2* had peak expression at 60 d postpartum, in agreement with previous data on milk sphingolipid concentration [99]. Combined data on genes associated with ceramide synthesis suggest an increase in synthesis coupled with decreased degradation throughout lactation (Figure 5 and Additional file 1, Figure S5). An increase in mammary ceramide synthesis might potentially serve as a positive signal for proteins involved in lipid synthesis through activation of SREBP1 as suggested previously [101]. Increased sphingolipid synthesis during lactation also could affect availability of cholesterol for MFGM, and might explain the inverse pattern between milk cholesterol ester and sphingolipid [99].

Sterol and ceramides and their role in mammary lipid synthesis

Transport of ceramide from ER to Golgi is achieved by oxysterol binding proteins (OSBP), which also act as

sterol sensors whose function is to integrate cellular sterol status with sphingomyelin metabolism [103]. A novel role for *OSBP* in regulation of lipid synthesis was demonstrated when overexpression of *OSBP* led to increases in TAG synthesis in mouse liver and concomitant up-regulation of *SREBF1* and *INSIG1* [104]. mRNA abundance of *OSBP* and related genes (*OSBPL2*, *OSBPL10*) in mammary was comparable with that of *SREBF1* and 2, and their expression was up-regulated ~1.5-fold during lactation (Figure 5) suggesting a functional role, likely involving the regulation of *SREBF1* and the coordination of sphingolipid and cholesterol synthesis.

Transcription regulation networks and proposed milk fat synthesis model

We used our data set to develop gene networks using IPA (Figure 6). Details of networks are available in Additional file 1 (Supplementary Materials and Discussion) and the legend of Figure 6. The resulting networks clearly underscore a central role for *SREBF1*, *SREBF2*, and *PPARG* in controlling transcription of most of the genes assessed in

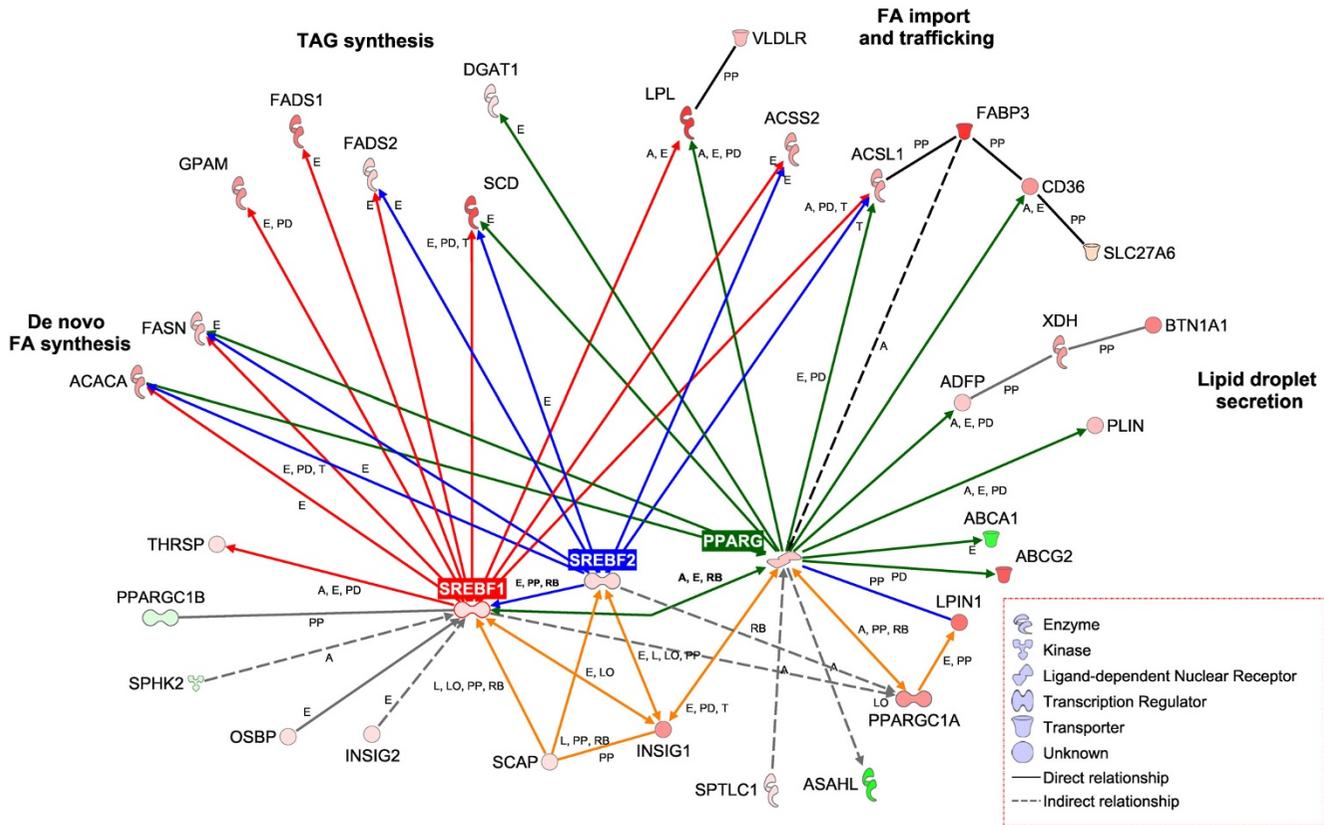


Figure 6
Networks among genes involved in milk fat synthesis. Networks were developed with Ingenuity Pathway Analysis® (Ingenuity Systems, <http://www.ingenuity.com>) and edited to incorporate results from the present and previous studies in bovine mammary tissue. Red nodes denote positive fold changes and green nodes negative fold changes in expression at 60 relative to -15 d. Red, blue, and green edges denote genes whose transcription is under the control of *SREBF1*, *SREBF2*, and *PPARG*, respectively. Highlighted in orange is the network encompassing *PPARG*, *PPARGC1A*, *LPIN1*, *INSIG1*, and *SCAP* which controls expression/function of *SREBF* proteins. Letters along the edges denote effects on activity (A), expression (E), localization (LO), proteolysis (L), RNA binding (RB), protein-DNA binding (PD), and protein-protein binding (PP). Genes are grouped based on their primary function during milk fat synthesis.

the present study which coordinately regulate milk fat synthesis. The network also highlights a putative role of *PPARG*, in coordination with *PPARGC1A* and *INSIG1*, in controlling function/expression of *SREBF1* (highlighted with orange edges). Thus, our data challenge the notion that *SREBF1* is the central player regulating lipid synthesis in bovine mammary tissue. We propose that a network of transcription regulators and nuclear receptors, including *SREBF1*, *SREBF2*, *PPARG*, *INSIG1*, and *PPARGC1A*, coordinate activation of the genes driving the lipid synthesizing machinery (Figure 6). More functional studies are clearly needed to determine whether long-term up-regula-

tion of transcription factors and nuclear receptors are determinant in inducing and maintaining milk fat synthesis. The specific roles of *INSIG1* and *PPARG* during lactation need to be determined. Besides transcriptional regulation, other regulatory steps can determine the activity of the protein. For example, short term regulation of *ACACA* activity also occurs at the post-translational level [105].

Taken together, our findings allowed for the development of an up-to-date model of milk fat synthesis regulation in bovine mammary tissue (Figure 7). The model incorpo-

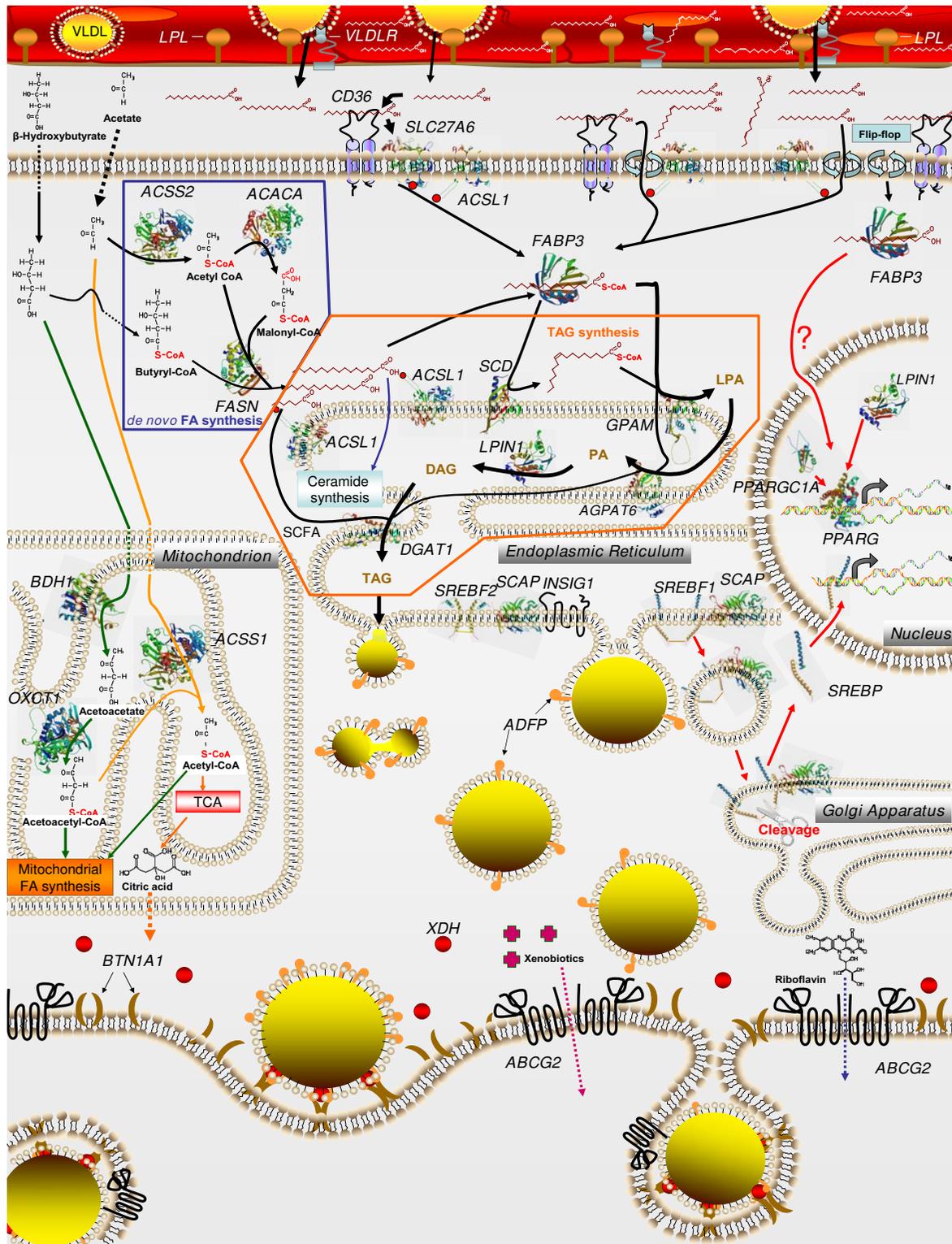


Figure 7
Interrelationships among cellular pathways regulating milk fat synthesis in bovine mammary tissue. Detailed description of the model is reported in the discussion section. Protein structures, when available, are the most updated from ModBase http://modbase.compbio.ucsf.edu/modbase.cgi/search_form.cgi.

rates the most recent information available, including our data, on enzymes involved in milk fat synthesis (i.e., sub-cellular location, fate of FA, and putative role). Following is the description of the model presented in Figure 7:

- **Endothelial long-chain FA (LCFA) transport:** the VLDL TAG core (or chylomicrons) is hydrolyzed by LPL in cooperation with VLDLR, which anchor the lipoprotein molecule and releases LCFA for transport across the endothelium into the extracellular space. LCFA are imported into mammary cells via flip-flop (passive or driven by "flippases") and protein-mediated mechanisms. CD36 appears to be the most important protein regulating LCFA import, and it might work in concert with FATP6 (SLC27A6). Once LCFA cross the membrane, most are activated into an LC-acyl-CoA (LCACoA) primarily via ACSL1 (activation shown by small red dots). Formation of LCACoA essentially traps LCFA inside the cell and activates them for subsequent utilization. LCACoA are captured primarily by FABP3, which transports them to specific intracellular organelles for utilization.
- **FA channelling and metabolic fates:** LCFA or LCACoA bound to FABP3 likely have three primary fates: 1) serve as substrate for SCD, which inserts a double bound at Δ^9 position of the LCFA (primarily 16:0 and 18:0), and the endogenous FA (primarily oleic acid) is subsequently transported by FABP4 to enzymes involved in TAG synthesis; 2) serve directly as substrate for TAG synthesis via sequential reactions carried out by GPAM (LCACoA \Rightarrow *sn*-1 of Glycerol-3-P to form lysophosphatidic acid [LPA]), GPAM in the present model is located on the ER instead in the mitochondria, where the proper location for the product of this gene is. Previous studies have demonstrated larger microsomal compared with mitochondrial GPAT activity in lactating bovine mammary tissue [106]. To date, however, only the mammalian mitochondrial GPAT have been annotated and characterized. The insertion of GPAM in the ER attempts to account for this limitation and to simplify the model. Other enzymes important for TAG synthesis include AGPAT6 (LCACoA \Rightarrow *sn*-2 position of LPA to form phosphatidic acid [PA]), LPIN1 (cleaves the P group of PA to form diacylglycerol [DAG]) and to a less extent, DGAT1 (LCACoA \Rightarrow *sn*-3 position of DAG to form TAG). The TAG pathway is denoted by black arrows and encircled by an orange line; 3) can regulate transcription via PPARG (see below).
- **Lipid droplet formation:** once TAG is formed it is inserted into the intra-leaflet of the ER membrane to form lipid droplet. In bovine mammary, ADFP (orange shape) is central for the formation of lipid droplets and for the secretory pathway involving BTN1A1 (brown half moon). The XDH (red circle) also seems to play a role in the mechanism encompassing ADFP and BTN1A1.
- **De novo FA synthesis, activation, channelling, and formation of TAG and phospholipid:** *de novo* FA synthesis (encircled in blue) is carried out by ACACA and FASN utilizing acetyl-CoA and butyryl-CoA. Formation of acetyl-CoA from acetate is carried out by ACSS2. Once FA with >10-carbons are formed they are activated by ACSL1 and bound to FABP3, which allows the FA to enter into the TAG synthesis pathway. Short-chain FA (SCFA) are inserted into TAG via DGAT1. A portion of palmitate (both from *de novo* synthesis and import) can be utilized for sphingolipid synthesis through ceramide (Additional file 1, Figure S5).
- **Transcriptional regulation:** denoted by red arrows. We propose a role for FABP3 in activation of gene expression by FA through PPAR γ . Besides up-regulation during lactation, a chief role of PPARG in mammary lipid metabolism is supported by the large up-regulation in expression of PPARGC1A and LPIN1 two important PPAR γ co-factors. Regulation of genes involved in *de novo* synthesis might partly be under control of SREBP1 (SREBF1), which is bound to SCAP in the ER and it is transported to the Golgi where it is cleaved. Activated SREBP1 then enters the nucleus and could regulate gene expression. However, the observed up-regulation of INSIG1 (binds SCAP and blocks SREBP transport to Golgi) could dampen SREBP activity in bovine mammary during lactation or at the very least elicit tight regulation of SREBP activity. Our data, together with previous findings, highlight a possible role of PPARG in regulating SREBP activity through regulation of INSIG1 expression (Figure 6). Results support an overlapping role of SREBP2 in regulation of expression of gene involved in *de novo* FA synthesis. Alternate routes of acetate and butyrate utilization are denoted by green (BHBA) and orange (acetate) arrows. Up-regulation in expression of ACSS1 during lactation allow utilization of acetate in mitochondria for energy generation primarily; whereas, mRNA up-regulation of BDH1 and OXCT1 allow BHBA entry into mitochondria. Primary routes of BHBA use are citrate synthesis (Additional file 1, Figure S3) and mitochondrial FA synthesis (Additional file 1, Figure S4).
- **Membrane-associated transporters:** the marked mRNA abundance and up-regulation of ABCG2 during lactation suggest a pivotal role of this gene in milk synthesis/secretion. Demonstrated roles of ABCG2 protein in mammary tissue include secretion of riboflavin (blue dotted arrow) and xenobiotics (violet dotted arrow).

Conclusion

Lactation was characterized by dramatic up-regulation in expression of genes associated with FA uptake from blood (e.g., *LPL*, *CD36*) and intracellular transport/channelling (e.g., *FABP3*). These adaptations were mirrored in milk FA profiles, showing that mammary uptake relative to *de novo*

synthesis predominated in early lactation. Although of lower magnitude, lactation also induced up-regulation of mRNA of genes involved in activation of FA (e.g., *ACSL1*, *ACSS2*), *de novo* synthesis (e.g., *ACACA*, *FASN*), desaturation (e.g., *SCD*, *FADS1*), synthesis of TAG (e.g., *AGPAT6*, *GPAM*), lipid droplet formation (e.g., *BTN1A1*, *XDH*), and ketone body utilization (e.g., *BDH1*, *OXCT1*). Temporal expression of genes with well-defined roles in mammary lipid metabolism peaked at 60 d post-partum and to some extent followed the lactation curve.

We could deduce a central role in endogenous oleic acid synthesis via SCD for mammary TAG synthesis. However, there was no statistical correlation between expression patterns of genes involved in desaturation and (Δ^5 , Δ^6 , Δ^9) desaturase indexes rendering their use to infer temporal enzyme expression/activity meaningless. Furthermore, expression data highlighted the importance of ketone body utilization, mitochondrial biogenesis and PPAR γ activity (*PPARGC1A*), and lipid droplet formation (*BTN1A1*, *XDH*, *ADFP*) in the global scheme of milk fat synthesis and secretion (Figure 7). Novel findings included a likely role for *PPARG*, *LASS2*, *INSIG1*, *SREBF2*, and *OSBP* in regulating lipid synthesis and mammary intracellular equilibrium between cholesterol and sphingolipids.

The complexity of mammary molecular adaptations over time was underscored by gene network analysis (Figure 6) as well as the apparent interrelationships that must coordinate the overall process of milk fat synthesis and secretion (Figure 7). This point is further exemplified by the large number of annotated transcripts, among them several transcription factors, whose expression is markedly up-regulated during lactation [8] most of which have currently unknown functions in bovine mammary tissue.

Abbreviations

ACE: acetyl-CoA incorporation during FA Elongation; BHBA: β -hydroxybutyrate; DAG: diacylglycerol; FA: fatty acid(s); LCFA: long-chain fatty acid(s); LPA: lysophosphatidic acid; MGF: milk fat globule membrane; PA: phosphatidic acid; qPCR: real-time RT-PCR; SCFA: short-chain fatty acid(s); TAG: triacylglycerol(s)

Authors' contributions

MB conducted qPCR analysis, data transformation, handling and statistical analysis of data, and drafted the manuscript. JLL collected mammary biopsies and milk samples, conceived and designed the study, participated in its coordination, and helped draft the manuscript. All authors read and approved the final manuscript.

Additional material

Additional file 1

Supplementary Materials and Methods and Results and Discussion. The file contain additional Materials and Methods (RNA extraction, PCR, and design and evaluation of primers and details about measurement of milk yield, milk composition and fatty acid analysis) accompanied by 4 tables which include cows features (Table S1), qPCR primers information (Table S2) and validation (Tables S3 and S4); the file contain also additional Results and Discussion about milk FA composition, accompanied by 3 tables (Table S5 for PCR features, Table S6 for mole/day and Table S7 for g/100 g of milk fatty acids with calculated indexes) and 6 figures which include: the entire curve of lactation and milk fat yield and % (Figure S1), Δ^5 and Δ^6 calculated indexes (Figure S2), possible model of utilization of BHBA in bovine mammary (Figure S3); possible utilization of BHBA for mitochondrial FA synthesis (Figure S4), and possible model of phospholipid metabolism in bovine mammary during lactation (Figure S5). For each figure a detailed discussion is also provided in the caption.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2164-9-366-S1.pdf>]

Additional file 2

Excel file with correlation. The file contains Pearson correlations among all measurements (genes and FA yield – mole/day). Correlations were analyzed using PROC CORR of SAS (SAS Inst. Inc. Cary, NC, release 8.0). The file contains 3 sheets with correlations and a legend.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-2164-9-366-S2.xls>]

Acknowledgements

Partial financial support for the gene expression work and milk fatty acid analysis was provided by the Cooperative State Research, Education and Extension Service, U. S. Department of Agriculture, under project No. ILLU-538-307 (to JLL). We gratefully acknowledge the help from the staff of the University of Illinois Dairy Research and Teaching Unit for animal care, Dr. Robin E. Everts (University of Illinois) for his continued support with annotation of the bovine microarray platform, Dr. Harris A. Lewin (University of Illinois) for the continued collaborations using the bovine oligonucleotide microarray, and Dr. Walter L. Hurley for helpful suggestions during development of the manuscript.

References

- Bauman DE, Mather IH, Wall RJ, Lock AL: **Major advances associated with the biosynthesis of milk.** *J Dairy Sci* 2006, **89(4)**:1235-1243.
- Bauman DE, Davis CL: **Biosynthesis of milk fat.** In *Lactation: a comprehensive treatise Volume 2*. Edited by: Larson BL, Smith VR. New York: Academic Press; 1974:31-75.
- Keenan TW, Mather IH: **Intracellular Origin of Milk Fat Globules and the Nature of the Milk Fat Globule Membrane.** In *Advanced Dairy Chemistry: Lipids Volume 2*. Edited by: Fox PF, McSweeney PLH. New York, NY: Springer; 2006:137-171.
- Kay JK, Weber WJ, Moore CE, Bauman DE, Hansen LB, Chester-Jones H, Crooker BA, Baumgard LH: **Effects of week of lactation and genetic selection for milk yield on milk fatty acid composition in Holstein cows.** *J Dairy Sci* 2005, **88(11)**:3886-3893.
- Garnsworthy PC, Masson LL, Lock AL, Mottram TT: **Variation of milk citrate with stage of lactation and de novo fatty acid synthesis in dairy cows.** *J Dairy Sci* 2006, **89(5)**:1604-1612.

6. Bernard L, Leroux C, Chilliard Y: **Expression and nutritional regulation of lipogenic genes in the ruminant lactating mammary gland.** *Adv Exp Med Biol* 2008, **606**:67-108.
7. Anderson SM, Rudolph MC, McManaman JL, Neville MC: **Key stages in mammary gland development. Secretory activation in the mammary gland: it's not just about milk protein synthesis!** *Breast Cancer Res* 2007, **9**(1):204.
8. Bionaz M, Rodriguez-Zas SL, Everts RE, Lewin HA, Loor JJ: **Mamm-Omics™: transcript profiling of the mammary gland during the lactation cycle in Holstein cows.** *ADSA-PSA-ASAS-AMPA Joint Annual Meeting: 2007: J Dairy Sci* 2007:272.
9. Ollier S, Robert-Granie C, Bernard L, Chilliard Y, Leroux C: **Mammary transcriptome analysis of food-deprived lactating goats highlights genes involved in milk secretion and programmed cell death.** *J Nutr* 2007, **137**(3):560-567.
10. Tramontana S, Hurley WL, Bionaz M, Sharma A, Graugnard D, Cutler EA, Everts RE, Ajmone-Marsan P, Rodriguez-Zas SL, Loor JJ: **Mamm-Omics™ in Sus scrofa: uncovering genomic adaptations underlying mammary development during pregnancy and lactation.** *ADSA-PSA-ASAS-AMPA Joint Annual Meeting: 2008: J Dairy Sci* 2008 in press.
11. Lemay DG, Neville MC, Rudolph MC, Pollard KS, German JB: **Gene regulatory networks in lactation: identification of global principles using bioinformatics.** *BMC Syst Biol* 2007, **1**:56.
12. Morandini P, Salamini F, Gantet P: **Engineering of Plant Metabolism for Drug and Food.** *Current Medicinal Chemistry – Immunology, Endocrine & Metabolic Agents* 2005, **5**:103-112.
13. Gygi SP, Rochon Y, Franza BR, Aebersold R: **Correlation between protein and mRNA abundance in yeast.** *Mol Cell Biol* 1999, **19**(3):1720-1730.
14. Patel MS, Korotchkina LG: **Regulation of the pyruvate dehydrogenase complex.** *Biochem Soc Trans* 2006, **34**(Pt 2):217-222.
15. McManaman JL, Russell TD, Schaack J, Orlicky DJ, Robenek H: **Molecular determinants of milk lipid secretion.** *J Mammary Gland Biol Neoplasia* 2007, **12**(4):259-268.
16. Palmquist DL: **Milk fat: Origin of fatty acids and influence of nutritional factors thereon.** In *Advanced Dairy Chemistry: Lipids Volume 2*. Edited by: Fox PF, McSweeney PLH. New York, NY: Springer; 2006:43-92.
17. van Herwaarden AE, Wagenaar E, Merino G, Jonker JW, Rosing H, Beijnen JH, Schinkel AH: **Multidrug transporter ABCG2/breast cancer resistance protein secretes riboflavin (vitamin B2) into milk.** *Mol Cell Biol* 2007, **27**(4):1247-1253.
18. Donkor J, Sariahmetoglu M, Dewald J, Brindley DN, Reue K: **Three mammalian lipins act as phosphatidate phosphatases with distinct tissue expression patterns.** *J Biol Chem* 2007, **282**(6):3450-3457.
19. Bionaz M, Loor JJ: **ACSL1, AGPAT6, FABP3, LPINI, and SLC27A6 are the most abundant isoforms in bovine mammary tissue and their expression is affected by stage of lactation.** *J Nutr* 2008, **138**(6):1019-1024.
20. Bionaz M, Loor JJ: **Identification of reference genes for quantitative real-time PCR in the bovine mammary gland during the lactation cycle.** *Physiol Genomics* 2007, **29**(3):312-319.
21. Jones DH: **The Mitochondria of the Mammary Parenchymal Cell in Relation to Pregnancy-Lactation Cycle.** In *Lactation: a comprehensive treatise Volume 4*. Edited by: Larson BL, Smith VR. New York: Academic Press; 1974:503-512.
22. Fielding BA, Frayn KN: **Lipoprotein lipase and the disposition of dietary fatty acids.** *Br J Nutr* 1998, **80**(6):495-502.
23. Rudolph MC, McManaman JL, Phang T, Russell T, Kominsky DJ, Serkova NJ, Stein T, Anderson SM, Neville MC: **Metabolic regulation in the lactating mammary gland: a lipid synthesizing machine.** *Physiological Genomics* 2007, **28**(3):323-336.
24. Jensen DR, Gavigan S, Sawicki V, Witsell DL, Eckel RH, Neville MC: **Regulation of lipoprotein lipase activity and mRNA in the mammary gland of the lactating mouse.** *Biochem J* 1994, **298**(Pt 2):321-327.
25. Tacken PJ, Hofker MH, Havekes LM, van Dijk KW: **Living up to a name: the role of the VLDL receptor in lipid metabolism.** *Curr Opin Lipidol* 2001, **12**(3):275-279.
26. Uchida T, Onda K, Bonkobara M, Thongsong B, Matsuki N, Inaba M, Ono K: **Utilization of intestinal triglyceride-rich lipoproteins in mammary gland of cows.** *J Vet Med Sci* 1999, **61**(10):1143-1146.
27. Raphael BC, Dimick PS, Puppione DL: **Lipid characterization of bovine serum lipoproteins throughout gestation and lactation.** *J Dairy Sci* 1973, **56**(8):1025-1032.
28. Doege H, Stahl A: **Protein-mediated fatty acid uptake: Novel insights from in vivo models.** *Physiology (Bethesda)* 2006, **21**:259-268.
29. Reinhardt TA, Lippolis JD: **Bovine milk fat globule membrane proteome.** *J Dairy Res* 2006, **73**(4):406-416.
30. Stahl A: **A current review of fatty acid transport proteins (SLC27).** *Pflügers Arch* 2004, **447**(5):722-727.
31. Mashek DG, Coleman RA: **Cellular fatty acid uptake: the contribution of metabolism.** *Curr Opin Lipidol* 2006, **17**(3):274-278.
32. Fujino T, Kondo J, Ishikawa M, Morikawa K, Yamamoto TT: **Acetyl-CoA synthetase 2, a mitochondrial matrix enzyme involved in the oxidation of acetate.** *Journal of Biological Chemistry* 2001, **276**(14):11420-11426.
33. Luong A, Hannah VC, Brown MS, Goldstein JL: **Molecular characterization of human acetyl-CoA synthetase, an enzyme regulated by sterol regulatory element-binding proteins.** *Journal of Biological Chemistry* 2000, **275**(34):26458-26466.
34. Mellenberger RW, Bauman DE, Nelson DR: **Metabolic adaptations during lactogenesis. Fatty acid and lactose synthesis in cow mammary tissue.** *Biochem J* 1973, **136**(3):741-748.
35. Annonson EF: **Metabolite Utilization by the Ruminant Mammary Gland.** In *Biochemistry of lactation* Edited by: Mephan TB. Amsterdam-New York: Elsevier Science Publishers; 1983:399-436.
36. McArthur MJ, Atshaves BP, Frolov A, Foxworth WD, Kier AB, Schroeder F: **Cellular uptake and intracellular trafficking of long chain fatty acids.** *J Lipid Res* 1999, **40**(8):1371-1383.
37. Frolov A, Cho TH, Murphy EJ, Schroeder F: **Isoforms of rat liver fatty acid binding protein differ in structure and affinity for fatty acids and fatty acyl CoAs.** *Biochemistry* 1997, **36**(21):6545-6555.
38. Whetstone HD, Hurley WL, Davis CL: **Identification and characterization of a fatty acid binding protein in bovine mammary gland.** *Comp Biochem Physiol B* 1986, **85**(3):687-692.
39. Faergeman NJ, Wadum M, Feddersen S, Burton M, Kragelund BB, Knudsen J: **Acyl-CoA binding proteins; structural and functional conservation over 2000 MYA.** *Mol Cell Biochem* 2007, **299**(1-2):55-65.
40. Mikkelsen J, Knudsen J: **Acyl-CoA-binding protein from cow. Binding characteristics and cellular and tissue distribution.** *Biochem J* 1987, **248**(3):709-714.
41. Awad AC, Shin HS, Romsos DR, Gray JI: **Direct desaturation of free myristic acid by hen liver microsomal Delta9-desaturase without prior activation to myristoyl-CoA derivative.** *J Agric Food Chem* 2004, **52**(10):3194-3201.
42. van Herwaarden AE, Wagenaar E, Karnekamp B, Merino G, Jonker JW, Schinkel AH: **Breast cancer resistance protein (Bcrp1/Abcg2) reduces systemic exposure of the dietary carcinogens aflatoxin B1, IQ and Trp-P-1 but also mediates their secretion into breast milk.** *Carcinogenesis* 2006, **27**(1):123-130.
43. Cohen-Zinder M, Seroussi E, Larkin DM, Loor JJ, Everts-van der Wind A, Lee JH, Drackley JK, Band MR, Hernandez AG, Shani M, Lewin HA, Weller JL, Ron M: **Identification of a missense mutation in the bovine ABCG2 gene with a major effect on the QTL on chromosome 6 affecting milk yield and composition in Holstein cattle.** *Genome Res* 2005, **15**(7):936-944.
44. Jensen RG: **The composition of bovine milk lipids: January 1995 to December 2000.** *J Dairy Sci* 2002, **85**(2):295-350.
45. Farke C, Vitorro E, Meyer HH, Albrecht C: **Identification of the bovine cholesterol efflux regulatory protein ABCA1 and its expression in various tissues.** *J Anim Sci* 2006, **84**(11):2887-2894.
46. Takahashi K, Kimura Y, Nagata K, Yamamoto A, Matsuo M, Ueda K: **ABC proteins: key molecules for lipid homeostasis.** *Med Mol Morphol* 2005, **38**(1):2-12.
47. Short VJ, Brindley DN, Dils R: **Co-ordinate changes in enzymes of fatty acid synthesis, activation and esterification in rabbit mammary gland during pregnancy and lactation.** *Biochem J* 1977, **162**(2):445-450.
48. Ntambi JM, Miyazaki M: **Recent insights into stearoyl-CoA desaturase-1.** *Curr Opin Lipidol* 2003, **14**(3):255-261.
49. Chung M, Ha S, Jeong S, Bok J, Cho K, Baik M, Choi Y: **Cloning and characterization of bovine stearoyl CoA desaturase cDNA from adipose tissues.** *Biosci Biotechnol Biochem* 2000, **64**(7):1526-1530.

50. Lengi AJ, Corl BA: **Identification and characterization of a novel bovine stearoyl-CoA desaturase isoform with homology to human SCD5.** *Lipids* 2007, **42(6)**:499-508.
51. **Cow (Bos taurus) Genome Browser Gateway** [<http://genome.ucsc.edu/>]. 19 April 2008
52. Myers EW, Miller W: **Optimal alignments in linear space.** *Comput Appl Biosci* 1988, **4(1)**:11-17.
53. Kinsella JE: **Stearyl CoA as a precursor of oleic acid and glycerolipids in mammary microsomes from lactating bovine: possible regulatory step in milk triglyceride synthesis.** *Lipids* 1972, **7(5)**:349-355.
54. Archibeque SL, Lunt DK, Gilbert CD, Tume RK, Smith SB: **Fatty acid indices of stearoyl-CoA desaturase do not reflect actual stearoyl-CoA desaturase enzyme activities in adipose tissues of beef steers finished with corn-, flaxseed-, or sorghum-based diets.** *J Anim Sci* 2005, **83(5)**:1153-1166.
55. Barber MC, Ward RJ, Richards SE, Salter AM, Buttery PJ, Vernon RG, Travers MT: **Ovine adipose tissue monounsaturated fat content is correlated to depot-specific expression of the stearoyl-CoA desaturase gene.** *J Anim Sci* 2000, **78(1)**:62-68.
56. Bernard L, Rouel J, Leroux C, Ferlay A, Faulconnier Y, Legrand P, Chilliard Y: **Mammary lipid metabolism and milk fatty acid secretion in alpine goats fed vegetable lipids.** *J Dairy Sci* 2005, **88(4)**:1478-1489.
57. Bickerstaffe R, Johnson AR: **The effect of intravenous infusions of sterculic acid on milk fat synthesis.** *Br J Nutr* 1972, **27(3)**:561-570.
58. Schennink A, Heck JML, Bovenhuis H, Visser MHPW, van Valenberg HJF, van Arendonk JAM: **Milk Fatty Acid Unsaturation: Genetic Parameters and Effects of Stearoyl-CoA Desaturase (SCD1) and Acyl CoA: Diacylglycerol Acyltransferase I (DGATI).** *J Dairy Sci* 2008, **91(5)**:2135-2143.
59. Glascock RF, Welch VA: **Contribution of the fatty acids of three low density serum lipoproteins to bovine milk fat.** *J Dairy Sci* 1974, **57(11)**:1364-1370.
60. Looor JJ, Herbein JH: **Reduced fatty acid synthesis and desaturation due to exogenous trans 10, cis 12-CLA in cows fed oleic or linoleic oil.** *J Dairy Sci* 2003, **86(4)**:1354-1369.
61. Leroy JL, Vanholder T, Mateusen B, Christophe A, Opsomer G, de Kruif A, Genicot G, Van Soom A: **Non-esterified fatty acids in follicular fluid of dairy cows and their effect on developmental capacity of bovine oocytes in vitro.** *Reproduction* 2005, **130(4)**:485-495.
62. Keenan TW, Huang CM: **Membranes of mammary gland. VI. Lipid and protein composition of Golgi apparatus and rough endoplasmic reticulum from bovine mammary gland.** *J Dairy Sci* 1972, **55(11)**:1586-1596.
63. Hanhoff T, Lucke C, Spener F: **Insights into binding of fatty acids by fatty acid binding proteins.** *Mol Cell Biochem* 2002, **239(1-2)**:45-54.
64. Nakamura MT, Nara TY: **Gene regulation of mammalian desaturases.** *Biochem Soc Trans* 2002, **30(Pt 6)**:1076-1079.
65. Rodriguez-Cruz M, Tovar AR, Palacios-Gonzalez B, Del Prado M, Torres N: **Synthesis of long-chain polyunsaturated fatty acids in lactating mammary gland: role of Delta5 and Delta6 desaturases, SREBP-I, PPARalpha, and PGC-I.** *J Lipid Res* 2006, **47(3)**:553-560.
66. Gonzalez-Baro MR, Lewin TM, Coleman RA: **Regulation of Triglyceride Metabolism. II. Function of mitochondrial GPAT I in the regulation of triacylglycerol biosynthesis and insulin action.** *Am J Physiol Gastrointest Liver Physiol* 2007, **292(5)**:G1195-1199.
67. Coleman RA, Lee DP: **Enzymes of triacylglycerol synthesis and their regulation.** *Prog Lipid Res* 2004, **43(2)**:134-176.
68. Beigneux AP, Vergnes L, Qiao X, Quatela S, Davis R, Watkins SM, Coleman RA, Walzem RL, Philips M, Reue K, Young SG: **Agpat6 - a novel lipid biosynthetic gene required for triacylglycerol production in mammary epithelium.** *J Lipid Res* 2006, **47(4)**:734-744.
69. Chen YQ, Kuo MS, Li S, Bui HH, Peake DA, Sanders PE, Thibodeaux SJ, Chu S, Qian YW, Zhao Y, Bredt DS, Moller DE, Konrad RJ, Beigneux AP, Young SG, Cao G: **AGPAT6 Is a Novel Microsomal Glycerol-3-phosphate Acyltransferase.** *J Biol Chem* 2008, **283(15)**:10048-10057.
70. Finck BN, Gropler MC, Chen Z, Leone TC, Croce MA, Harris TE, Lawrence JC Jr, Kelly DP: **Lipin 1 is an inducible amplifier of the hepatic PGC-1alpha/PPARalpha regulatory pathway.** *Cell Metab* 2006, **4(3)**:199-210.
71. Reue K, Zhang P: **The lipin protein family: dual roles in lipid biosynthesis and gene expression.** *FEBS Lett* 2008, **582(1)**:90-96.
72. Huffman TA, Mothe-Satney I, Lawrence JC Jr: **Insulin-stimulated phosphorylation of lipin mediated by the mammalian target of rapamycin.** *Proc Natl Acad Sci USA* 2002, **99(2)**:1047-1052.
73. Marshall MO, Knudsen J: **Specificity of diacylglycerol acyltransferase from bovine mammary gland, liver and adipose tissue towards acyl-CoA esters.** *Eur J Biochem* 1979, **94(1)**:93-98.
74. Marshall MO, Knudsen J: **The specificity of 1-acyl-sn-glycerol 3-phosphate acyltransferase in microsomal fractions from lactating cow mammary gland towards short, medium and long chain acyl-CoA esters.** *Biochim Biophys Acta* 1977, **489(2)**:236-241.
75. Grisart B, Coppieters W, Farnir F, Karim L, Ford C, Berzi P, Cambisano N, Mni M, Reid S, Simon P, Spelman R, Georges M, Snell R: **Positional candidate cloning of a QTL in dairy cattle: identification of a missense mutation in the bovine DGATI gene with major effect on milk yield and composition.** *Genome Res* 2002, **12(2)**:222-231.
76. Cases S, Zhou P, Shillingford JM, Wiseman BS, Fish JD, Angle CS, Hennighausen L, Werb Z, Farese RV Jr: **Development of the mammary gland requires DGATI expression in stromal and epithelial tissues.** *Development* 2004, **131(13)**:3047-3055.
77. Grisart B, Farnir F, Karim L, Cambisano N, Kim JJ, Kvasz A, Mni M, Simon P, Frere JM, Coppieters W, Georges M: **Genetic and functional confirmation of the causality of the DGATI K232A quantitative trait nucleotide in affecting milk yield and composition.** *Proc Natl Acad Sci USA* 2004, **101(8)**:2398-2403.
78. Robenek H, Hofnagel O, Buers I, Lorkowski S, Schnoor M, Robenek MJ, Heid H, Troyer D, Severs NJ: **Butyrophilin controls milk fat globule secretion.** *Proc Natl Acad Sci USA* 2006, **103(27)**:10385-10390.
79. Tansey JT, Sztalryd C, Hlavin EM, Kimmel AR, Londos C: **The central role of perilipin a in lipid metabolism and adipocyte lipolysis.** *IUBMB Life* 2004, **56(7)**:379-385.
80. Palmquist DL, Davis CL, Brown RE, Sachan DS: **Availability and Metabolism of Various Substrates in Ruminants. V. Entry Rate into the Body and Incorporation into Milk Fat of D()beta)-Hydroxybutyrate.** *J Dairy Sci* 1969, **52(5)**:633-638.
81. Robinson AM, Williamson DH: **Physiological roles of ketone bodies as substrates and signals in mammalian tissues.** *Physiol Rev* 1980, **60(1)**:143-187.
82. Page MA, Williamson DH: **Lactating mammary gland of the rat: a potential major site of ketone-body utilization.** *Biochem J* 1972, **128(2)**:459-460.
83. Harvatine KJ, Bauman DE: **SREBP1 and thyroid hormone responsive spot 14 (S14) are involved in the regulation of bovine mammary lipid synthesis during diet-induced milk fat depression and treatment with CLA.** *J Nutr* 2006, **136(10)**:2468-2474.
84. Espenshade PJ, Hughes AL: **Regulation of Sterol Synthesis in Eukaryotes.** *Annu Rev Genet* 2007.
85. Eberle D, Hegarty B, Bossard P, Ferre P, Fufelle F: **SREBP transcription factors: master regulators of lipid homeostasis.** *Biochimie* 2004, **86(11)**:839-848.
86. Horton JD, Goldstein JL, Brown MS: **SREBPs: activators of the complete program of cholesterol and fatty acid synthesis in the liver.** *J Clin Invest* 2002, **109(9)**:1125-1131.
87. Engelking LJ, Kuriyama H, Hammer RE, Horton JD, Brown MS, Goldstein JL, Liang G: **Overexpression of Insig-I in the livers of transgenic mice inhibits SREBP processing and reduces insulin-stimulated lipogenesis.** *J Clin Invest* 2004, **113(8)**:1168-1175.
88. Li J, Takaishi K, Cook W, McCorkle SK, Unger RH: **Insig-I "brakes" lipogenesis in adipocytes and inhibits differentiation of preadipocytes.** *Proc Natl Acad Sci USA* 2003, **100(16)**:9476-9481.
89. Bitman J, Wood DL, Capuco AV: **Changes in the lipid composition of the secretions of the bovine mammary gland during the dry period.** *J Dairy Sci* 1992, **75(2)**:435-442.
90. McCarthy RD, Patton S: **Cholesterol esters and the synthesis of milk fat.** *Biochim Biophys Acta* 1963, **70**:102-103.
91. Desvergne B, Michalik L, Wahli W: **Transcriptional regulation of metabolism.** *Physiol Rev* 2006, **86(2)**:465-514.

92. Wang M, Master SR, Chodosh LA: **Computational expression deconvolution in a complex mammalian organ.** *BMC Bioinformatics* 2006, **7**:328.
93. Wan Y, Saghatelian A, Chong LW, Zhang CL, Cravatt BF, Evans RM: **Maternal PPAR gamma protects nursing neonates by suppressing the production of inflammatory milk.** *Genes Dev* 2007, **21(15)**:1895-1908.
94. Kadegowda AKG, Bionaz M, Piperova LS, Erdman RA, Loor JJ: **Lipogenic gene expression in MAC-T cells is affected differently by fatty acids and enhanced by PPAR-gamma activation.** *ADSA-PSA-ASAS-AMPA Joint Annual Meeting: 2008: J Dairy Sci* 2008 in press.
95. Liang H, Ward WF: **PGC-1alpha: a key regulator of energy metabolism.** *Adv Physiol Educ* 2006, **30(4)**:145-151.
96. Kast-Woelbern HR, Dana SL, Cesario RM, Sun L, de Grandpre LY, Brooks ME, Osburn DL, Reifel-Miller A, Klausning K, Leibowitz MD: **Rosiglitazone induction of Insig-1 in white adipose tissue reveals a novel interplay of peroxisome proliferator-activated receptor gamma and sterol regulatory element-binding protein in the regulation of adipogenesis.** *J Biol Chem* 2004, **279(23)**:23908-23915.
97. Jazwinski SM, Conzelmann A: **LAG1 puts the focus on ceramide signaling.** *Int J Biochem Cell Biol* 2002, **34(11)**:1491-1495.
98. Tani M, Ito M, Igarashi Y: **Ceramide/sphingosine/sphingosine 1-phosphate metabolism on the cell surface and in the extracellular space.** *Cell Signal* 2007, **19(2)**:229-237.
99. Bitman J, Wood DL: **Changes in milk fat phospholipids during lactation.** *J Dairy Sci* 1990, **73(5)**:1208-1216.
100. Heid HW, Keenan TW: **Intracellular origin and secretion of milk fat globules.** *European Journal of Cell Biology* 2005, **84(2-3)**:245-258.
101. Worgall TS: **Sphingolipids: major regulators of lipid metabolism.** *Curr Opin Clin Nutr Metab Care* 2007, **10(2)**:149-155.
102. Zierath JR: **The path to insulin resistance: paved with ceramides?** *Cell Metab* 2007, **5(3)**:161-163.
103. Olkkonen VM, Johansson M, Suchanek M, Yan D, Hynynen R, Ehnholm C, Jauhiainen M, Thiele C, Lehto M: **The OSBP-related proteins (ORPs): global sterol sensors for co-ordination of cellular lipid metabolism, membrane trafficking and signaling processes?** *Biochem Soc Trans* 2006, **34(Pt 3)**:389-391.
104. Yan D, Lehto M, Rasilainen L, Metso J, Ehnholm C, Yla-Herttuala S, Jauhiainen M, Olkkonen VM: **Oxysterol binding protein induces upregulation of SREBP-1c and enhances hepatic lipogenesis.** *Arterioscler Thromb Vasc Biol* 2007, **27(5)**:1108-1114.
105. Munday MR: **Regulation of mammalian acetyl-CoA carboxylase.** *Biochem Soc Trans* 2002, **30(Pt 6)**:1059-1064.
106. Vernon RG, Faulkner A, Finley E, Pollock H, Taylor E: **Enzymes of glucose and fatty acid metabolism of liver, kidney, skeletal muscle, adipose tissue and mammary gland of lactating and non-lactating sheep.** *J Anim Sci* 1987, **64(5)**:1395-1411.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

