

RESEARCH ARTICLE

Open Access

DNA methylation patterns provide insight into epigenetic regulation in the Pacific oyster (*Crassostrea gigas*)

Mackenzie R Gavery, Steven B Roberts*

Abstract

Background: DNA methylation is an epigenetic mechanism with important regulatory functions in animals. While the mechanism itself is evolutionarily ancient, the distribution and function of DNA methylation is diverse both within and among phylogenetic groups. Although DNA methylation has been well studied in mammals, there are limited data on invertebrates, particularly molluscs. Here we characterize the distribution and investigate potential functions of DNA methylation in the Pacific oyster (*Crassostrea gigas*).

Results: Methylation sensitive PCR and bisulfite sequencing PCR approaches were used to identify CpG methylation in *C. gigas* genes and demonstrated that this species possesses intragenic methylation. *In silico* analysis of CpGo/e ratios in publicly available sequence data suggests that DNA methylation is a common feature of the *C. gigas* genome, and that specific functional categories of genes have significantly different levels of methylation.

Conclusions: The Pacific oyster genome displays intragenic DNA methylation and contains genes necessary for DNA methylation in animals. Results of this investigation suggest that DNA methylation has regulatory functions in *Crassostrea gigas*, particularly in gene families that have inducible expression, including those involved in stress and environmental responses.

Background

Epigenetic mechanisms induce changes in gene activity without alteration to the underlying DNA sequence [1]. Common epigenetic mechanisms include DNA methylation, histone modifications and non-coding RNA activity. The most well-studied of these is DNA methylation, which refers to the addition of a methyl group to position 5 of cytosines. In animals, this reaction is catalyzed by a family of enzymes called DNA (cytosine-5) methyltransferases (DNMTs) and occurs almost exclusively in CpG dinucleotides. DNA methylation is typically associated with transcriptional repression, and is primarily achieved by methylation in gene promoters [2-4]. The functional significance of DNA methylation in vertebrates includes providing genomic stability [5], regulation of imprinted genes [6] and X-chromosome inactivation [7]. In mammals, DNA methylation is essential for development and cell differentiation [8] and defects or unintended changes

in DNA methylation can have deleterious consequences such as embryonic lethality [9] and tumorigenesis [10]. DNA methylation, like many epigenetic marks, may be heritable, therefore unintended changes as a result of environmental exposures or other processes can be passed on for multiple generations [11].

The extent of cytosine methylation varies considerably among eukaryotes. In vertebrates, approximately 70-80% of cytosines in CpG dinucleotides are methylated [12], a pattern referred to as global methylation. Invertebrates display a wide range of DNA methylation, from very limited methylation in *Drosophila melanogaster* [13] and *Caenorhabditis elegans* [14] to a mosaic pattern of methylation in the sea urchin (*Strongylocentrotus purpuratus*) [15] and *Ciona intestinalis* [16,17]. Bird and Taggart [12] concluded that there were three general types of methylation patterns: the 'insect-type' which shows little to no methylation, the 'echinoderm-type', the genomes of which contain both methylated and non-methylated fractions, and the heavily methylated 'vertebrate-type'. Recent studies in the honey bee (*Apis*

* Correspondence: sr320@u.washington.edu
School of Aquatic and Fishery Sciences, University of Washington, 1122 NE Boat Street, Seattle, Washington, USA

mellifera) indicate these patterns may be more complex [18,19]. In contrast to *D. melanogaster*, which lacks most of the classical DNMTs [20] and shows limited cytosine methylation [21], *A. mellifera* has a fully functional set of DNA methylation enzymes and shows substantial methylation across its genome [18].

In vertebrates, regulation of transcription by DNA methylation is accomplished by differential patterns of methylation in intergenic regions, namely gene promoters [2-4]. In contrast, there are no significant differences reported in the methylation status of gene promoters in invertebrates, where methylation appears to be targeted specifically to transcription units [17,22]. Computational analyses of the methylation status of *A. mellifera* genes have provided some of the first evidence supporting a regulatory role of intragenic DNA methylation in invertebrates [19,23]. In these studies, genes associated with general metabolic or 'housekeeping' functions were predicted to be hyper-methylated, whereas caste-specific genes were preferentially hypo-methylated. This functional clustering suggests DNA methylation serves to regulate gene transcription in *A. mellifera*, however, it is uncertain if this function is conserved across invertebrate taxa. Furthermore, it is unclear exactly how intragenic cytosine methylation directly affects transcription.

Studies in *A. mellifera* and others illustrate the diversity of DNA methylation patterns in invertebrate taxa and highlight gaps in our understanding of the evolutionary and functional significance of DNA methylation. One taxonomic group that has been notably absent from these investigations is the phylum Mollusca. Molluscs were first categorized as having 'echinoderm-type' DNA methylation patterns based on experimental evidence using the common mussel (*Mytilus edulis*) [12]. Since then, there has been little investigation of DNA methylation in molluscs with the exception of evidence suggesting the presence CpG methylation in the clam, *Donax trunculus* [24]. In addition to increasing our understanding of the evolution of DNA methylation in invertebrate taxa, this study provides an opportunity to evaluate the Pacific oyster (*Crassostrea gigas*) as a model organism for analyzing DNA methylation in an aquatic species. Bivalve molluscs are important bioindicators [25] and elucidating the functional significance of DNA methylation in these organisms may prove valuable for understanding the effects of environmental stress in aquatic organisms. Here, we report the first investigation into DNA methylation profiles in the genome of the Pacific oyster. We confirm the presence of intragenic CpG methylation in *C. gigas*. We also demonstrate a relationship between predicted methylation status and gene function, suggesting that DNA methylation performs important regulatory functions in *C. gigas*.

Implications of these findings are discussed in both an evolutionary and ecological context.

Results

Methylation Sensitive PCR

A Methylation Sensitive PCR (MSP) approach was used to identify specific methylated sites. Five genes associated with immune function were analyzed and methylation status determined (Table 1). Methylation status can be concluded based on the presence or absence of a PCR product in the methylation sensitive HpaII digest. Of the five genes analyzed, CpG methylation was confirmed for *heat shock protein 70* (*hsp70*), whereas no methylation was detectable at restriction site(s) for the other sequences examined. The CpG observed to expected ratios (see Methods for calculation) are included in Table 1 for each gene. It should be noted that *hsp70* has the lowest ratio of all the genes analyzed (0.57). This low ratio is predictive of a hyper-methylated status, which is confirmed here by MSP.

Bisulfite Sequencing PCR

In order to describe methylated cytosines outside of CCGG sites, Bisulfite Sequencing PCR (BSP) was used. Five genes predicted to be hyper-methylated, and five predicted to be hypo-methylated (based on CpG observed to expected ratio) were randomly selected for analysis. Valid PCR products were produced for two of the genes. This is a typical result as the conversion of unmethylated cytosines results in challenges for primer specificity. Four individual clones were sequenced for each of the two products. There was a 100% conversion rate for non-CpG cytosines for each of the clones sequenced. In the first fragment, a 136 bp fragment with homology to the amino terminal fragment of the human neuromedin-u receptor [Swiss-Prot: Q9GZQ4"], one of seven CpGs sites displayed methylation in 25% of the clones sequenced (Figure 1(a)). In a second fragment, one of two CpGs sites was determined to be methylated in 50% of the clones sequenced in a 93 bp region (Figure 1(b)). The latter sequence has significant homology to human bromodomain adjacent to zinc finger domain, 1A [Swiss-Prot: Q9NRL2].

In Silico Analysis of *C. gigas* Transcriptome

The ratio of observed to expected CpG dinucleotides (CpGo/e) was used to predict methylation status in the *C. gigas* transcriptome. This approach is based on the known hyper-mutability of methylated cytosines, which readily deaminate to thymine residues [26]. This CpG mutation is not easily corrected by DNA repair machinery, and as a result consistently methylated regions of DNA are depleted of CpG dinucleotides over evolutionary time [27]. Consequently, regions of DNA with a low

Table 1 Results of Methylation Specific PCR analysis for five *C. gigas* genes

Accession #	Best blast hit [Organism]	Undigested	HpaII	MspI	Number of restriction sites	CpG o/e
EW778441	heat shock protein 70 [<i>Crassostrea gigas</i>]	+	+	-	2	0.57
EW777519	heat shock protein 25 [<i>Danio rerio</i>]	+	-	-	3	0.81
EW778166	cytochrome P450 [<i>Haliotis diversicolor</i>]	+	-	-	1	0.85
EW778608	macrophage expressed protein 1-like protein [<i>Crassostrea gigas</i>]	+	-	-	6	1.08
EW778905	14-3-3 protein gamma (Protein kinase C inhibitor protein 1 [<i>Bos taurus</i>])	+	-	-	2	0.92

Results of methylation status of five genes associated with immune response by MSP. PCR was carried out on undigested, HpaII digested, and MspI digested DNA. Presence (+) or absence (-) of PCR product is indicated. Number of CCGG restriction sites in the indicated sequence and CpGo/e ratios are also provided.

CpGo/e are predicted to be methylated, whereas regions with a high CpGo/e (approaching 1.0) are predicted to be unmethylated. This approach has been used to reliably predict methylation status across many taxonomic groups [17,19,22,28].

A non-redundant *C. gigas* contig database, 'GigasDatabase' version 6 [29] was utilized for this analysis. To ensure only CpG (and not GpC) dinucleotides were being evaluated, analysis was limited to annotated sequences. The probability density function of the CpGo/e for 12,210 annotated *C. gigas* expressed sequence tag (EST) contigs is illustrated in Figure 2. We find that the data fit a bimodal mixture model (blue curve) significantly better than a unimodal distribution. The red curves represent the scaled, normal mixture components, which have means of 0.40 (\pm 0.12 SD) and 0.70 (\pm 0.21 SD) respectively (Figure 2). A majority of the contigs have a CpGo/e less than 1.0.

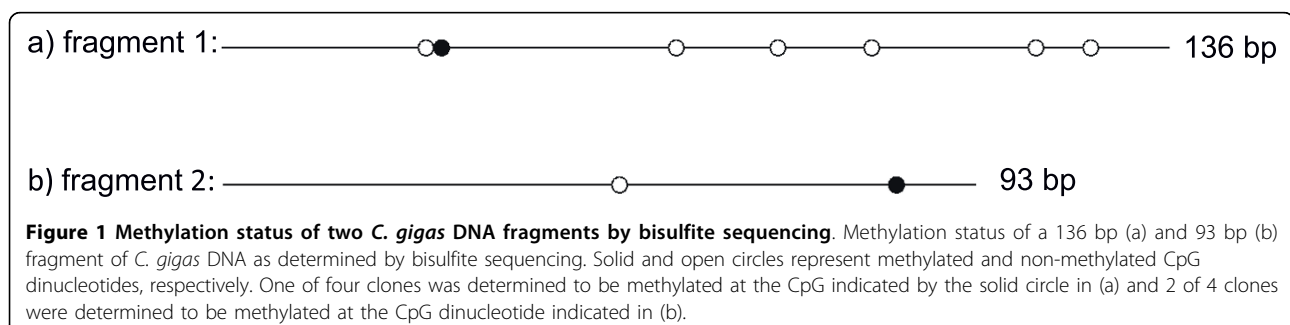
The ratio of observed to expected GpC dinucleotide frequencies (GpCo/e) was calculated in order to be assured that the bimodal distribution of CpGo/e was not biased toward G+C content of specific genes as there are no known mechanisms for preferential depletion of the GpC dinucleotide. As predicted, the ratio of observed to expected GpC's approaches 1.0 following a unimodal Gaussian distribution (Figure 2 inset). In addition, there is a significant negative correlation between CpGo/e and TpGo/e ($p = 0.00$) indicating that the

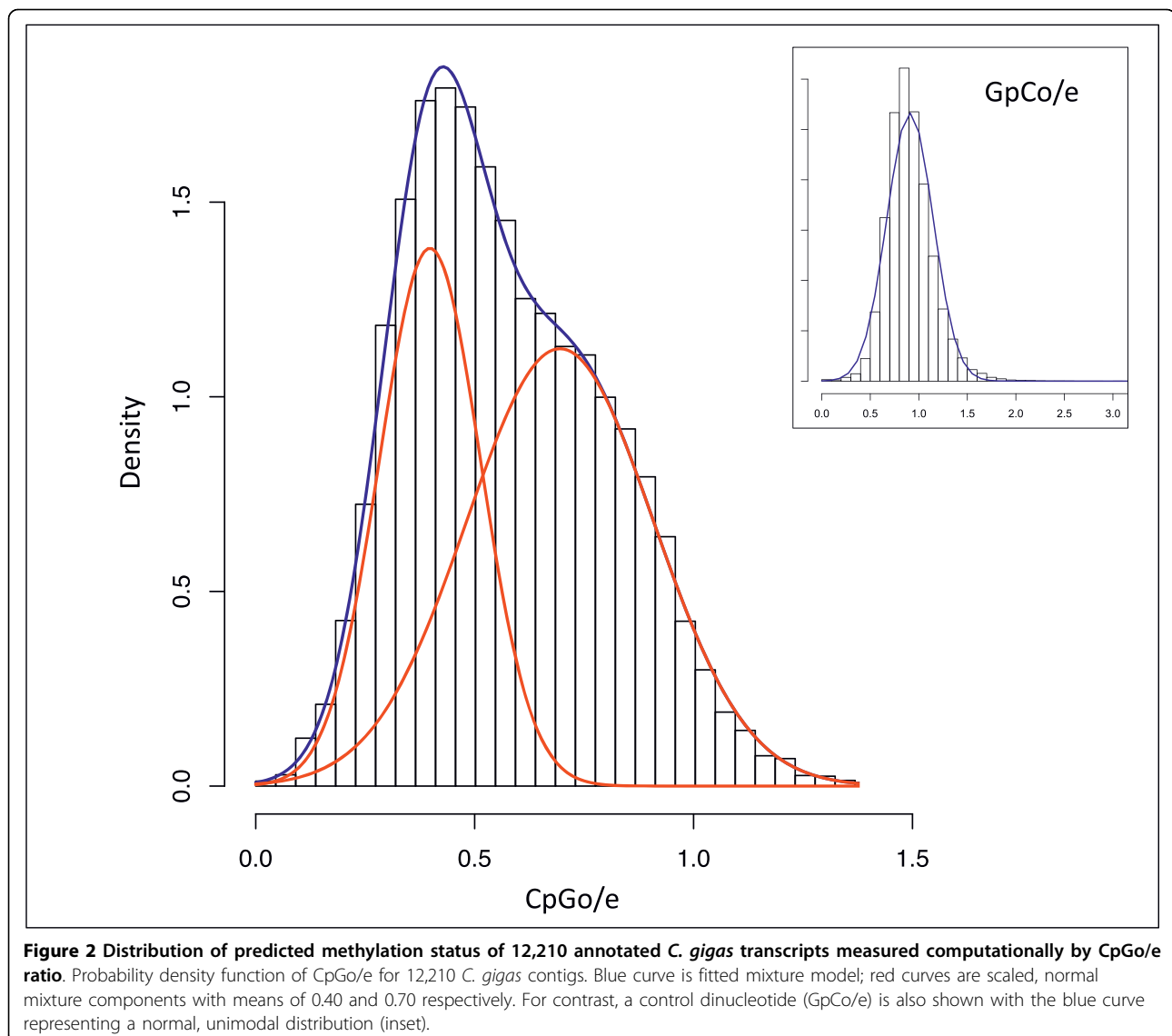
depletion of CpG dinucleotides is associated with the conversion of methylated CpG sites to TpG dinucleotides.

In order to determine any functional difference that may exist among those genes with lower than expected CpGo/e ratios, data were analyzed in the context of each gene's biological process GO Slim term (Figure 3). Several biological processes have CpGo/e ratios that are significantly different from each other (see Additional file 1: Matrix of p-values for comparisons between GO Slim categories based on CpGo/e). Specifically, genes with lower CpGo/e ratios (predicted to be hyper-methylated) were associated with DNA metabolism, RNA metabolism, and cell cycle and proliferation. Biological processes with higher CpGo/e ratios (predicted to be hypo-methylated) include cell adhesion, cell-to-cell signalling and signal transduction. This analysis confirms that the normal mixture components described previously in Figure 2 are enriched with genes from particular functional categories.

Discussion

Results of methylation specific PCR and bisulfite sequencing PCR indicate that the Pacific oyster (*Crassostrea gigas*) genome is methylated. Further evidence supporting the presence and importance of methylation in *C. gigas* is the identification of genes that encode DNA methyltransferases (DNMT), the family of proteins responsible for the

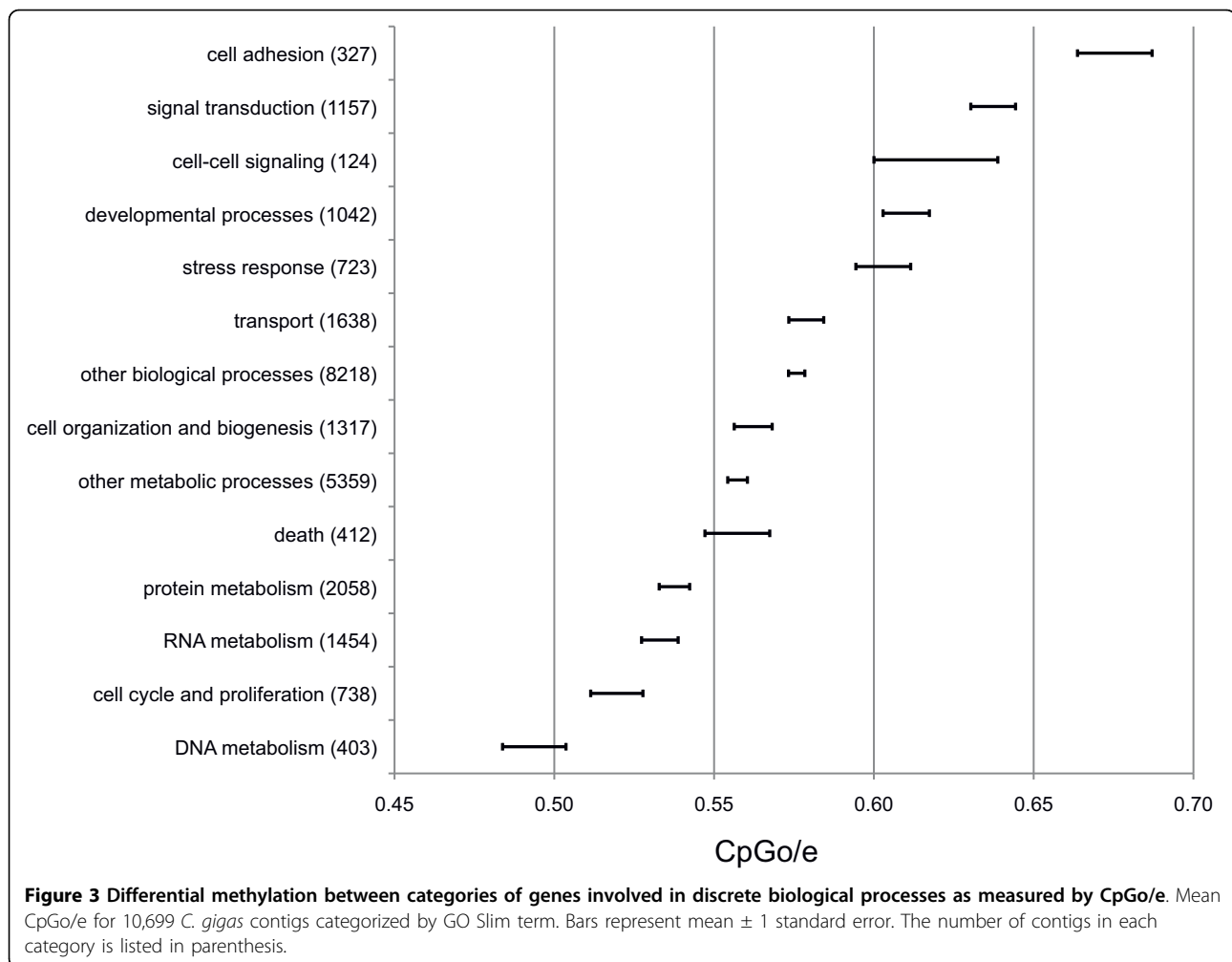




enzymatic conversion of cytosine to 5-methylcytosine. Animals that lack DNA methylation such as *C. elegans* also lack essential DNMTs, while invertebrates with an intermediate level of DNA methylation such as honey bees, sea urchins and urochordates have the full set of DNMT genes [30]. Sequences with high homology to DNMT3 (responsible for *de novo* methylation), DNMT1 (associated with maintenance methylation), and methyl-CpG-binding domain protein 2 (mediation of the effects of DNA methylation) are present in a publicly available *C. gigas* contig database, GigasDatabase version 6 [29]. These annotated sequences can be found in GigasDatabase with accession numbers CU684371.p.cg.6 (e-value $1e-61$), CU994437.p.cg.6 (e-value $2e-26$), and AM861084.p.cg.6 (e-value $1e-11$), respectively. While a DNMT2 homolog has not been identified, it may not be required for DNA

methylation in *C. gigas* as it functions primarily as a tRNA methyltransferase and shows only weak DNA methyltransferase activity *in vitro* [31]. DNMTs are an evolutionarily conserved group of proteins, but show structural diversity both within and among taxa [32]. The evolutionary diversity of DNA methylation within and among phylogenetic groups provides justification for further evaluation of the functions of this epigenetic mark.

The presence of intragenic methylation in *C. gigas* is similar to that of other invertebrates that primarily exhibit intragenic DNA methylation patterns [33,17], the roles of which have been largely unexplored. Studies of DNA methylation in mammals have generally focused on promoter regions, where hyper-methylation of promoters inhibits initiation of transcription [2]. In contrast, invertebrate genomes do not show differentially



methylated gene promoters [22]. One of the long-standing hypotheses is that intragenic DNA methylation prevents inappropriate initiation of transcription outside of promoter regions [34]; however new studies have begun to investigate a more active role for intragenic DNA methylation, namely in regulation of expression. For example, exonic DNA methylation has been shown to regulate transcription of the *phytochrome A* gene in *Arabidopsis thaliana* [35]. In humans, investigation of intragenic CpG islands (≥ 200 bp regions with G+C content of at least 50% and CpGo/e close to expected) has revealed that CpG islands in terminal exons may regulate transcription of non-coding RNAs [36]. Here, using BSP, we observed methylation variability in two CpG sites that may indicate cell-specific methylation. The function of intragenic DNA methylation in *C. gigas* cannot be conclusively determined from this study, but results of studies in other organisms suggest that it could be involved in either repression of transcription outside of transcription start sites and/or regulation of expression.

Within the transcriptome of the Pacific oyster, a significant difference in methylation pattern was observed across gene families. A majority of *C. gigas* genes analyzed were depleted in CpG dinucleotides (i.e. CpGo/e < 1.0) and show a significantly bimodal distribution, suggesting that DNA methylation is a common feature of the *C. gigas* transcriptome, and that certain groups of genes have significantly different levels of methylation. The bimodal distribution of CpGo/e is similar to the pattern observed in the honey bee *A. mellifera*, where authors reported a hyper-methylated fraction that was enriched in genes involved with general metabolic functions and a hypo-methylated fraction enriched with genes that are associated with caste-specific functions [19]. Similarly when *C. gigas* transcripts were clustered according to their functional annotations using GO Slim terms, we see that the two distributions are comprised of functionally distinct classes of genes with varying regulatory requirements. Specifically, genes predicted to be hyper-methylated are associated with housekeeping functions and those predicted to be hypo-methylated are

associated with general immune functions. Hyper-methylation of intragenic regions of housekeeping genes is consistent between *C. gigas* and *A. mellifera* [19], but stands in contrast to observations in vertebrates, where distinct hypo-methylation of housekeeping gene promoters is associated with global expression [37]. Constitutive DNA methylation in housekeeping genes in *C. gigas* could be important for repressing transcription outside of promoter regions as previously discussed. It has also been proposed that hyper-methylation of housekeeping genes in *A. mellifera* indicates epigenetic control of gene activity in housekeeping genes [23]. Further experiments will be required to determine whether hyper-methylation of housekeeping genes plays a passive role in preventing inappropriate transcription or a more active role in maintaining expression in *C. gigas*.

Highest CpGo/e ratios were observed in genes involved in the oyster's innate immune response, including categories of cell adhesion, cell-cell signaling, and signal transduction. Our experimental data using MSP supports the predicted hypo-methylation of this class of genes as only 1 of the 5 immune related genes were methylated. Our results do not indicate that DNA methylation is entirely absent from genes in the hypo-methylated group as CpG depletion is still observed (CpGo/e 0.7) which stands in contrast to the hypo-methylated genes in *A. mellifera* (CpGo/e >1.0). One explanation as to why it would be advantageous for this class of genes to be hypo-methylated is that it allows for greater epigenetic flexibility and higher regulatory control. Oysters have been shown to have high phenotypic plasticity in response to environmental changes and stress [38,39] and it could be postulated that an epigenetic mark, such as DNA methylation, could provide this level of control. DNA methylation has been generally considered to be a less dynamic epigenetic mark, however, it has been reported in plants that DNA methylation levels are involved in regulating gene expression in response to stress and show active methylation and demethylation in response to various stressors [40-42]. It has been hypothesized from these studies that DNA methylation is a possible mechanism to impart protection against local stresses in future generations [43]. The identification of genes involved in demethylation in *C. gigas* would be an important step toward uncovering the nature of these epigenetic marks.

DNA methylation patterns have been shown to be heritable in mammalian taxa [44], and changes in DNA methylation patterns can persist for multiple generations [45]. Little work has been done to investigate heritability of DNA methylation in invertebrates, although a recent study of the crustacean, *Daphnia magna*, has shown transgenerational heritability of DNA methylation patterns after exposures to 5-azacytidine [46]. If DNA

methylation does play a role in regulation of transcription in *C. gigas* it may provide a mechanism not only for regulating responses to stress, but also for adapting to local stressors through heritability of DNA methylation patterns. Investigating the potential of epigenetic control in mechanisms of local adaptation may prove useful in understanding impacts of anthropogenic inputs in aquatic ecosystems and populations. Likewise, it is possible that epigenetic mechanisms may provide an explanation for other phenomena associated with heritability such as inbreeding depression and hybrid vigour.

Elucidating functional significance of DNA methylation in aquatic invertebrates may change the way we study impacts of environmental change in aquatic organisms. A range of factors such as diet [47,48], xenobiotic chemicals [49], and endocrine disruptors [11] have been shown to disrupt DNA methylation patterns. These epigenetic disruptions are increasingly associated with disease susceptibility, which in some cases can be passed on for multiple generations [50]. Although these investigations have been performed almost exclusively in mammalian species, recent studies have reported a dose dependent relationship between concentration of mercury and cadmium and total DNA methylation in *D. magna* [46,51]. Understanding which environmental factors can affect DNA methylation and elucidating the functional significance of DNA methylation in these important bioindicator species will be major steps toward clarifying the complex interactions between the environment, gene expression, and organismal responses.

Conclusions

The Pacific oyster genome displays methylation. *In silico* analysis reveals intragenic regions are targeted for methylation consistent with reports of methylation in other invertebrate species. Results of this investigation suggest that DNA methylation has regulatory functions in *Crassostrea gigas*, particularly in gene families involved in stress and environmental response. Experiments are underway in our lab to investigate relationships between the environment, DNA methylation, and control of gene expression to better characterize this process. In-depth analysis of methylation patterns in *Crassostrea gigas*, will help to advance the field of evolutionary epigenetics and will serve to illuminate functions of DNA methylation in invertebrates.

Methods

Animal collection & DNA isolation

Oysters used in this study were collected from naturalized *C. gigas* populations in Puget Sound, Washington. To isolate genomic DNA, 25 mg of gill tissue was processed according to the manufacturer's protocol using the DNeasy Blood & Tissue Kit (Qiagen, CA).

Methylation Sensitive PCR

Oyster genomic DNA was enzyme digested with either HpaII or MspI. Five immune related genes containing one or more CCGG recognition sites and covering a broad range of predicted methylation status (based on CpGo/e) were selected from a set of ESTs generated from a cDNA library of plated hemocytes [52]. PCR primers were designed to flank one or more restriction sites. Primer sequences are provided in Additional file 2: Primer Sequences. Quantitative PCR was performed using digested (HpaII or MspI) and undigested samples using 1× Immomix Master Mix (Bioline USA Inc., Boston, MA), 2 uM SYTO-13 (Invitrogen, Carlsbad, CA) and 0.2 uM forward and reverse primers in an Opticon 2 System (Bio-Rad, Hercules, CA) with the following cycling conditions: 10 min at 95C, followed by 37 cycles of 15 sec at 95C, 30 sec at 55C, and 1 min at 72C and a final extension at 72C for 10 min. Results were scored qualitatively based on the presence or absence of amplification as determined by fluorescence.

Bisulfite Conversion and Sequencing

Genomic DNA was bisulfite converted using the Epitect Bisulfite conversion kit (Qiagen, Carlsbad, CA). Briefly, 1.75 ug of DNA was subjected to treatment with sodium bisulfite at increased temperature to deaminate unmethylated cytosine residues to uracil. Following treatment, the solution was desulfonated on a column, washed and eluted.

To identify methylated cytosines in expressed regions of the oyster genome, Meth Primer [53] was used to design primers that flank multiple CpG sites, but do not contain CpGs. Primer sequences are provided in Additional file 2: Primer Sequences. The mean expected amplicon length for bisulfite primers was ~180 bp. PCR of bisulfite treated samples (54 ng/PCR reaction) was carried out using Taq DNA Polymerase Master Mix (Apex BioResearch Products, Research Triangle Park, NC) for 10 min at 95 C, followed by 40 cycles of 15 sec at 95 C, 30 sec at 55 C, and 1 min at 72C and a final extension at 72C for 10 min.

PCR products were separated using gel electrophoresis. Single bands were excised from the gel, purified using Ultra-DA purification columns (Ambion, Foster City, CA) and cloned using TOPO TA Cloning kit (Invitrogen). Four clones were sequenced for each primer pair. Methylation status was determined by comparing the sequence of bisulfite treated DNA to sequence of untreated DNA using Geneious 4.5.4 software (Biomatters Ltd., Auckland, NZ) and annotated using BLAST [54].

In Silico Analysis: Predicted DNA Methylation Status

For *in silico* analysis, the non-redundant *C. gigas* expressed sequence tag (EST) contig database,

'GigasDatabase' version 6 (http://public-contigbrowser.sigenae.org:9090/Crassostrea_gigas/index.html, [29]), was utilized. Analysis was limited to annotated sequences (n = 12,210) in order to be confident that sequences were reported in the 5' to 3' direction. It should be noted that this transcriptomic dataset is appropriate for predicting methylation status of the *C. gigas* genome as investigation into other invertebrate species shows that DNA methylation is specifically targeted to transcribed regions of the genome [17,12].

CpG observed/expected ratio (CpGo/e) was calculated using the following equation where *l* is the number of nucleotides in the contig:

$$\text{CpGo/e} = \frac{\text{number of CpG}}{\text{number of C} \times \text{number of G}} \times \frac{l^2}{l-1}$$

To evaluate the distribution of Pacific oyster contigs, a mixture model was fit to the CpGo/e ratios using the mixtools package [55] in R [56] yielding a two component mixture where $p_1 + p_2 = 1$. Hence the data C_i are distributed as:

$$C_i \sim p_1 N(\mu_1, \sigma_1) + p_2 N(\mu_2, \sigma_2).$$

The log likelihood statistic of the bimodal mixture model was compared to the normal null model to test for a significant improvement in fit.

In order to evaluate the variation of CpGo/e within and among functional classes of genes, contigs from the GigasDatabase annotated with a biological process GO term (n = 10,699 contigs) were assigned a functional group based on the MGI GO Slim database <http://www.informatics.jax.org>[57]. Since each contig may have multiple GO terms, contigs were allowed to fall into multiple GO Slim bins. However, to avoid weighting within a single category, an individual contig was not allowed to be included more than once in a single GO Slim bin. The mean CpGo/e and standard errors were calculated for each GO Slim term. A one-way ANOVA followed by Tukey's test for multiple comparisons was performed using SPSS software (SPSS Inc., Chicago, IL). A significance level of $p < 0.05$ was accepted.

Additional material

Additional file 1: Matrix of p-values for comparisons between GO Slim categories based on CpGo/e. CpGo/e for GO Slim categories were compared with Tukey's multiple comparison test. This file contains the p-values of each comparison. Significant differences ($p < 0.05$) are highlighted.

Additional file 2: Primer Sequences. This file contains primer sequences used for methylation sensitive PCR and bisulfite sequencing PCR analysis.

Acknowledgements

We thank Dustin Lennon for statistical support. This research was partially supported by the School of Aquatic and Fishery Sciences at the University of Washington.

Authors' contributions

MG and SR conceived of the study and prepared the manuscript. MG carried out the laboratory procedures. All authors read and approved the final manuscript.

Received: 26 February 2010 Accepted: 27 August 2010

Published: 27 August 2010

References

- Jablonka E, Lamb M: **The Changing Concept of Epigenetics.** *Ann N Y Acad Sci* 2002, **981**:82-96.
- Boyes J, Bird A: **Repression of genes by DNA methylation depends on CpG density and promoter strength: Evidence for involvement of a methyl-CpG binding protein.** *EMBO J* 1992, **11**:327-333.
- Kass SU, Pruss D, Wolffe AP: **How does DNA methylation repress transcription?** *Trends Genet* 1997, **13**:444-449.
- Hsieh CL: **Dependence of transcriptional repression on CpG methylation density.** *Mol Cell Biol* 1994, **14**:5487-5494.
- Maloisel L, Rossignol JL: **Suppression of crossing-over by DNA methylation in *Ascolobolus*.** *Genes Dev* 1998, **12**:1381-1389.
- Bell AC, Felsenfeld G: **Methylation of a CTCF-dependent boundary controls imprinted expression of the *Igf2* gene.** *Nature* 2000, **405**:482-5.
- Csankovszki G, Nagy A, Jaenisch R: **Synergism of Xist Rna, DNA Methylation, and Histone Hypoacetylation in Maintaining X Chromosome Inactivation.** *J Cell Biol* 2001, **153**:773-784.
- Okano M, Bell DW, Haber DA, Li E: **DNA methyltransferases Dnmt3a and Dnmt3b are essential for de novo methylation and mammalian development.** *Cell* 1999, **99**:247-257.
- Li E, Bestor TH, Jaenisch R: **Targeted mutation of the DNA methyltransferase gene results in embryonic lethality.** *Cell* 1992, **69**:915-926.
- Jones PA, Baylin SB: **The epigenomics of cancer.** *Cell* 2007, **128**:683-92.
- Anway MD, Skinner MK: **Epigenetic Transgenerational Actions of Endocrine Disruptors.** *Endocrinology* 2006, **147**:43-49.
- Bird AP, Taggart MH: **Variable patterns of total DNA and rDNA methylation in animals.** *Nucleic Acid Res* 1980, **8**:1485-1497.
- Gowher H, Leisemann O, Jeltsch A: **DNA of *Drosophila melanogaster* contains 5-methylcytosine.** *EMBO J* 2000, **19**:6918-6923.
- Simpson VJ, Johnson TE, Hammen RF: ***Caenorhabditis elegans* DNA does not contain 5-methylcytosine at any time during development or aging.** *Nucleic Acids Res* 1986, **14**:6711-6719.
- Bird AP, Taggart MH, Smith BA: **Methylated and unmethylated DNA compartments in the sea urchin genome.** *Cell* 1979, **17**:889-901.
- Simmen MW, Bird AP: **Sequence analysis of transposable elements in the sea squirt, *Ciona intestinalis*.** *Mol Biol Evol* 2000, **17**:1685-1693.
- Suzuki MM, Kerr ARW, De Sousa D, Bird A: **CpG methylation is targeted to transcription units in an invertebrate genome.** *Genome Res* 2007, **17**:625-631.
- Wang Y, Jorda M, Jones PL, Maleszka R, Ling X, Robertson HM, Mizzen CA, Peinado MA, Robinson GE: **Functional CpG methylation system in a social insect.** *Science* 2006, **314**:645-647.
- Elango N, Hunt BG, Goodisman MAD, Yi S: **DNA methylation is widespread and associated with differential gene expression in castes of the honeybee, *Apis mellifera*.** *Proc Natl Acad Sci USA* 2009, **106**:11206-11211.
- Hung MS, Karthikeyan N, Huang B, Koo HC, Kiger J, Shen CJ: ***Drosophila* proteins related to vertebrate DNA (5-cytosine) methyltransferases.** *Proc Natl Acad Sci USA* 1999, **96**:11940-11945.
- Lyko F, Ramsahoye BH, Jaenisch R: **DNA methylation in *Drosophila melanogaster*.** *Nature* 2000, **408**:538-540.
- Elango N, Yi SV: **DNA Methylation and Structural and Functional Bimodality of Vertebrate Promoters.** *Mol Biol and Evol* 2008, **25**:1602-1608.
- Foret S, Kucharski R, Pittelkow Y, Lockett GA, Maleszka R: **Epigenetic regulation of the honey bee transcriptome: unravelling the nature of methylated genes.** *BMC Genomics* 2009, **10**:472.
- Petrović V, Pérez-García C, Pasantes JJ, Šatović E, Prats E, Plohl M: **A GC-rich satellite DNA and karyology of the bivalve mollusk *Donax trunculus*: a dominance of GC-rich heterochromatin.** *Cytogenet Genome Res* 2009, **124**:63-71.
- Markert BA, Breure AM, Zechmeister HG: **Molluscs as Bioindicators.** In *Bioindicators and Biomonitoring*. Edited by: Markert BA, Breure AM, Zechmeister HG. Amsterdam: Elsevier Science; 2003:577-634.
- Coulondre C, Miller JH, Farabaugh PJ, Gilbert W: **Molecular basis of base substitution hotspots in *Escherichia coli*.** *Nature* 1978, **274**:775-780.
- Schorderet DF, Gartler SM: **Analysis of CpG suppression in methylated and nonmethylated species.** *Proc Natl Acad Sci USA* 1992, **89**:957-961.
- Shimizu TS, Takahashi K, Tomita M: **CpG distribution patterns in methylated and non-methylated species.** *Gene* 1997, **205**:103-107.
- Fleury E, Huvet A, Lelong C, de Lorgeril J, Boulo V, Gueguen Y, Bachère E, Tanguy A, Moraga D, Fabioux C, Lindeque P, Shaw J, Reinhardt R, Prunet R, Davey G, Lapègue S, Sauvage C, Corporeau C, Moal J, Gavery F, Wincker P, Moreeuv F, Klopp C, Mathieu M, Boudry P, Favrel B: **Generation and analysis of a 29,745 unique Expressed Sequence Tags from the Pacific oyster (*Crassostrea gigas*) assembled into a publicly accessible database: the GigasDatabase.** *BMC Genomics* 2009, **10**:341.
- Cañestro C, Yokoi H, Postlethwait JH: **Evolutionary developmental biology and genomics.** *Nat Rev Genet* 2007, **8**:932-942.
- Schaefer M, Lyko F: **Solving the Dnmt2 enigma.** *Chromosoma* 2010, **119**:35-40.
- Colot V, Rossignol J: **Eukaryotic DNA methylation as an evolutionary device.** *BioEssays* 1999, **21**:402-411.
- Tweedie S, Charlton J, Clark V, Bird A: **Methylation of genomes and genes at the invertebrate-vertebrate boundary.** *Mol Cell Biol* 1997, **17**:1469-1475.
- Bird AP: **Gene number, noise reduction and biological complexity.** *Trends Genet* 1995, **11**:94-100.
- Chawla R, Nicholson SJ, Folta KM, Srivastav V: **Transgene-induced silencing of *Arabidopsis* phytochrome A gene via exonic methylation.** *Plant J* 2007, **52**:1105-1118.
- Medvedeva Y, Fridman MV, Oparina NJ, Malko DB, Ermakova EO, Kulakovskiy IV, Heinzel A, Makeev V: **Intergenic, gene terminal, and intragenic CpG islands in the human genome.** *BMC Genomics* 2010, **11**:48.
- Weber M, Schubeler D: **Genomic patterns of DNA methylation: targets and function of an epigenetic mark.** *Current Opin Cell Biol* 2007, **19**:273-280.
- Hamdoun AM, Cheney DP, Cherr GN: **Phenotypic Plasticity of HSP70 and HSP70 Gene Expression in the Pacific Oyster (*Crassostrea gigas*): Implications for Thermal Limits and Induction of Thermal Tolerance.** *Biol Bull* 2003, **205**:160-169.
- Honkoop PJC, Bayne BL, Drent J: **Flexibility of size of gills and palps in the Sydney rock oyster *Saccostrea glomerata* (Gould, 1850) and the Pacific oyster *Crassostrea gigas* (Thunberg, 1793).** *J Exp Mar Biol Ecol* 2003, **282**:113-133.
- Kovarik A, Koukalova B, Bezdek M, Opatrn Z: **Hypermethylation of tobacco heterochromatic loci in response to osmotic stress.** *Theor Appl Genet* 1997, **95**:301-306.
- Steward N, Ito M, Yamaguchi Y, Koizumi N, Sano H: **Periodic DNA Methylation in Maize Nucleosomes and Demethylation by Environmental Stress.** *J Biol Chem* 2002, **277**:37741-37746.
- Choi CS, Sano H: **Abiotic-stress induces demethylation and transcriptional activation of a gene encoding a glycerophosphodiesterase-like protein in tobacco plants.** *Mol Genet Genome* 2007, **277**:589-600.
- Chinnusamy V, Zhu J: **Epigenetic regulation of stress responses in plants.** *Current Opin Plant Biol* 2009, **12**:133-139.
- Anway MD, Cupp AS, Uzumcu M, Skinner MK: **Epigenetic transgenerational actions of endocrine disruptors and male fertility.** *Science* 2005, **308**:1466-1469.
- Cropley JE, Suter CM, Beckman KB, Martin DIK: **Germ-line epigenetic modification of the murine A(vy) allele by nutritional supplementation.** *Proc Natl Acad Sci USA* 2006, **103**:17308-17311.
- Vandegheuchte MB, Lemièrre F, Vanhaecke L, Berghe WW, Janssen CR: **Direct and transgenerational impact on *Daphnia magna* of chemicals with a known effect on DNA methylation.** *Comp Biochem Physiol C Toxicol Pharmacol* .
- Wilson MJ, Shivapurkar N, Poirier LA: **Hypomethylation of hepatic nuclear DNA in rats fed with a carcinogenic methyl-deficient diet.** *Biochem J* 1984, **218**:987-990.

48. Dolinoy DC, Weidman JR, Waterland RA, Jirtle RL: **Maternal Genistein Alters Coat Color and Protects A^Y Mouse Offspring from Obesity by Modifying the Fetal Epigenome.** *Environ Health Perspect* 2006, **114**:567-572.
49. Sutherland JE, Costa M: **Epigenetics and the environment.** *Ann NY Acad Sci* 2003, **983**:151-160.
50. Jirtle RL, Skinner MK: **Environmental epigenomics and disease susceptibility.** *Nat Rev Genet* 2007, **8**:253-262.
51. Vandeghechuchte MB, Vanholme B, Haegeman A, Gheysen G, Janssen CR: **Occurrence of DNA methylation in *Daphnia magna* and influence of multigeneration Cd exposure.** *Environ Int* 2009, **35**:700-706.
52. Roberts S, Goetz G, White S, Goetz F: **Analysis of genes isolated from plated hemocytes of the Pacific oyster, *Crassostrea gigas*.** *Mar Biotechnol* 2009, **11**:24-44.
53. Li LC, Dahiya R: **MethPrimer: designing primers for methylation PCRs.** *Bioinformatics* 2002, **18**:1427-31.
54. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ: **Basic local alignment search tool.** *J Mol Biol* 1990, **215**:403-410.
55. Young DS, Hunter DR, Elmore RT, Xuan F, Hettmansperger TP, Thomas H: **The mixtools Package Version 0.2.0: Tools for Mixture Models.** *R Package Version 0.2.0* 2007 [<http://www.r-project.org>].
56. R Development Core Team: **R: A language and environment for statistical computing.** [<http://www.R-project.org>].
57. **Mouse Genome Database (MGD) at the Mouse Genome Informatics.** [<http://www.informatics.jax.org>].

doi:10.1186/1471-2164-11-483

Cite this article as: Gavery and Roberts: DNA methylation patterns provide insight into epigenetic regulation in the Pacific oyster (*Crassostrea gigas*). *BMC Genomics* 2010 **11**:483.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

