

RESEARCH ARTICLE

Open Access



Transcriptomics analysis revealing candidate genes and networks for sex differentiation of yesso scallop (*Patinopecten yessoensis*)

Liqing Zhou^{1,2†} , Zhihong Liu^{1,2†}, Yinghui Dong^{3*}, Xiujun Sun^{1,2}, Biao Wu^{1,2}, Tao Yu⁴, Yanxin Zheng⁴, Aiguo Yang^{1,2*}, Qing Zhao^{1,5} and Dan Zhao^{1,5}

Abstract

Background: The Yesso scallop, *Patinopecten (Mizuhopecten) yessoensis*, is a commercially important bivalve in the coastal countries of Northeast Asia. It has complex modes of sex differentiation, but knowledge of the mechanisms underlying this sex determination and differentiation is limited.

Results: In this study, the gonad tissues from females and males at three developmental stages were used to investigate candidate genes and networks for sex differentiation via RNA-Seq. A total of 901,980,606 high quality clean reads were obtained from 18 libraries, of which 417 expressed male-specific genes and 754 expressed female-specific genes. Totally, 10,074 genes differentially expressed in females and males were identified. Weighted gene co-expression network analysis (WGCNA) revealed that turquoise and green gene modules were significantly positively correlated with male gonads, while coral1 and black modules were significantly associated with female gonads. The most important gene for sex determination and differentiation was *Pydmrt 1*, which was the only gene discovered that determined the male sex phenotype during early gonadal differentiation. Enrichment analyses of GO terms and KEGG pathways revealed that genes involved in metabolism, genetic and environmental information processes or pathways are sex-biased. Forty-nine genes in the five modules involved in sex differentiation or determination were identified and selected to construct a gene co-expression network and a hypothesized sex differentiation pathway.

Conclusions: The current study focused on screening genes of sex differentiation in Yesso scallop, highlighting the potential regulatory mechanisms of gonadal development in *P. yessoensis*. Our data suggested that WGCNA can facilitate identification of key genes for sex differentiation and determination. Using this method, a hypothesized *P. yessoensis* sex determination and differentiation pathway was constructed. In this pathway, *Pydmrt 1* may have a leading function.

Keywords: Yesso scallop (*Patinopecten yessoensis*), Gonad, Sex determination and differentiation, Transcriptome, Weighted gene co-expression network analysis (WGCNA)

* Correspondence: dongyinghui118@126.com; yangag@ysfri.ac.cn

[†]Liqing Zhou and Zhihong Liu contributed equally to this work.

³Zhejiang Wanli University, Ningbo, China

¹Key Laboratory for Sustainable Development of Marine Fisheries, Ministry of Agriculture, Yellow Sea Fisheries Research Institute, Chinese Academy of Fishery Science, Qingdao, China

Full list of author information is available at the end of the article



Background

Patinopecten (Mizuhopecten) yessoensis is a cold-water species distributed along the coasts of China, Korea, Japan, and Russia. Particularly in the northern provinces of China, Liaoning, and the northern edge of the Shandong peninsula, it is large in size, fetches a high market price, and has become an important scallop species since it was introduced to China from Japan in 1980 [1]. In recent years, it has been hypothesized that hermaphrodites of the species could be used to improve a variety of traits through the construction of inbred lines, however, knowledge of the mechanisms underlying sex determination and differentiation in this species is very limited, which has hindered these improvements. It is known that the existence of hermaphrodites is commonly associated with the deterioration of aquaculture environments [2], nevertheless, we do not yet understand the molecular mechanisms underlying this association. Activation of the testis or ovarian pathway, or the repression of the alternative pathway, with many genes being expressed in a sexually dimorphic manner, determines the resulting sex, while all the pathway and mechanism of sex differentiation in mollusks were not clear. Amplified fragment length polymorphism (AFLP) markers have been used by several Chinese research groups to construct genetic linkage maps for *P. yessoensis* since 2009 [3, 4]. Based on these genetic linkage maps, in a previous study we screened some sex-related AFLP molecular markers under the assumptions that hermaphroditism was an independent group and that there were differences in genetic structure between male and female individuals [5]. In recent years, some genes related to sex determination or differentiation have been identified and characterized through transcriptome analysis of male and female mature gonads. These included *Dmrt1*, *Sox9*, *fem1*, and *Vasa* [6].

Data from the transcriptome sequencing and de novo analysis of Yesso Scallop helped to pinpoint candidate genes potentially involved in growth, reproduction, stress/immunity-responses, and so on [7]. Therefore, transcriptomics tools are widely used for screening sex determination and differentiation genes in many kinds of animals including bivalves. Teaniniuraitemoana et al. carried out transcriptome analysis on several *Pinctada margaritifera* gonadic samples from males and females at different development stages to identify potential sex differentiation and sex determining genes, such as *dmrt* and *fem-1* for males, and *foxl2* and vitellogenin for females [8]. The transcriptomes of three ovaries and three testes of Yesso scallop were sequenced and analyzed by Li et al., and the results showed that *PyFOXL2* was ovary-biased, and that *PyDMRT* and *PySOXH* were testis-biased, the three genes were presumed to be key candidates for scallop sex determination/differentiation [9]. Sex-related genes, such as *forkhead box L2 (foxl2)*, sex determining region Y-box (*Sox*), beta-catenin (β -*catenin*), chromobox homolog (*CBX*), and Sex-lethal

(*Sxl*) were also identified from the transcriptome data of gonads obtained from blood clams (*Tegillarca granosa*) [10]. As a correlation-based method that describes and visualizes networks of data points using the R software package [11], and an analysis method based on a large sample size of transcription group data, weighted gene correlation network analysis (WGCNA) can quickly identify core sex-related pathways and genes that play an important role in transcription regulation, and easily predict unknown gene regulatory relationships through known genes in the network. Yue et al. found that *CgDsx* and *CgFoxl* participate in the pathway of Pacific oyster sex determination, with *CgDsx* expressed more in males and *CgFoxl* expressed more in females, particularly during early gonad development [12]. To gain insight into the molecular mechanisms underlying female-superior sexual dimorphism with respect to growth rate, WGCNA analysis of female banana shrimp (*Fenneropenaeus merguensis*) digestive gland tissue samples was completed by Powell et al. and revealed a gene module containing a large number of transcripts with homology to transcription factors and genes associated with growth regulation [13]. Wang et al. found two gene modules (yellowgreen and salmon4) that were significantly positively correlated with female-biased sexual size dimorphism using WGCNA, and identified a series of hub genes by drawing an illustrated network map of these two modules in Chinese tongue sole (*Cynoglossus semilaevis*) [14].

Although the genomes between the sexes are largely similar, differences in gene expression in gonads can also be due to long noncoding RNA (lincRNA) [15] and other agents such as genetic sex determination (GSD) and environmental sex determination (ESD) factors. It is difficult to identify genes correlated with sex determination or differentiation through conventional RNA-Seq analysis. In the present study, we aimed to identify genes that may be important for sex differentiation by investigating transcriptional changes in Yesso scallop female and male individuals at three gonad development stages (mature, immature, and spawning) using WGCNA, based on the *Mizuhopecten yessoensis* genome data released on the NCBI (National Center for Biotechnology Information).

Results

Overall transcriptome and sequencing data

To better understand *P. yessoensis* sex determination and differentiation mechanisms, we conducted a comparative transcriptomic analysis. A total of 18 libraries were constructed and named as follows: PyUmf-1 to 3, PyUmm-1 to 3, PyMf-1 to 3, PyMm-1 to 3, PyOf-1 to 3, and PySm-1 to 3. Gonad development stage was identified via histological methods, and hermaphrodites were excluded. Unfortunately, one of the four individual gonad development stages of PySm-1 was not accurately

defined, which affect the trends of overall data, therefore, we excluded the data of PySm-1. As a result, a mean of 50,354,447 filtered clean reads with Q20 above 97.16% was obtained from each library and 1.55–2.75% of low quality reads mapped with rRNA were removed.

Identification of DEGs

Principal components analysis revealed strong clustering associated with sex, excluding sample PySm-1 (Fig. 1a). PC1 accounted for 44% of the differential expression and was correlated with sex. However, different development stages did not cluster obviously, as PC2 accounted for only 24.4% of the differential expression. In the sample relationship analysis, a heatmap plot revealed that sex identification of samples was exact except for the PySm-1 group (Fig. 1b). Therefore, we did not include the RNA-Seq data of sample PySm-1 in later analyses.

In the differential expression analysis, 3412 genes in PyUmf-vs-PyUmm, 2909 genes in PyOf-vs-PySm, and 2778 genes in PyMf-vs-PyMm were found to differentiate between females and males. Fewer differential genes were found between PyUmf and PyMf, PyMf and PyOf, PyUmf and PyOf, PyUmm and PySm, PyUmm and PyMm, and between PyMm and PySm, with 1504, 920, 381, 249, 42, and 14 differential genes, respectively (fold change ≥ 2 , $P < 0.05$). DEGs were very abundant between females and males (Fig. 2). However, there were fewer DEGs between different gonad development stages.

Subsequently, a total of 23,292 known genes and 2237 novel genes were identified via transcript reconstruction of the *P. yessoensis* genome. Of the 25,529 protein coding genes analyzed, 1171 showed significant differences between female and male gonads, and there were more female-specific genes (754) than male-specific genes (417) (Additional file 1: Figure S1).

Gene co-expression network interactions differ between the sexes

WGCNA revealed that the genes of female and male scallop gonads can be clustered into 19 modules (Fig. 3), which were further classified and clustered by similarity = 0.75 and minModuleSize = 50, with module sizes ranging from 79 to 4058. Of these modules, the turquoise module with size 1541 and the green module with size 1451 were significantly positively correlated with male gonads and were negatively correlated with female gonads. In contrast, the coral1 module with size 1371 and the black module with size 860 were significantly positively correlated with female gonads and were negatively correlated with male gonads.

Functional enrichment analysis of the genes

To better understand the function of these differentially expressed genes in sex differentiation or determination, GO term enrichment (Fig. 4) and KEGG pathway enrichment were conducted. Seventy-eight GO terms (belonging to biological processes, cellular components,

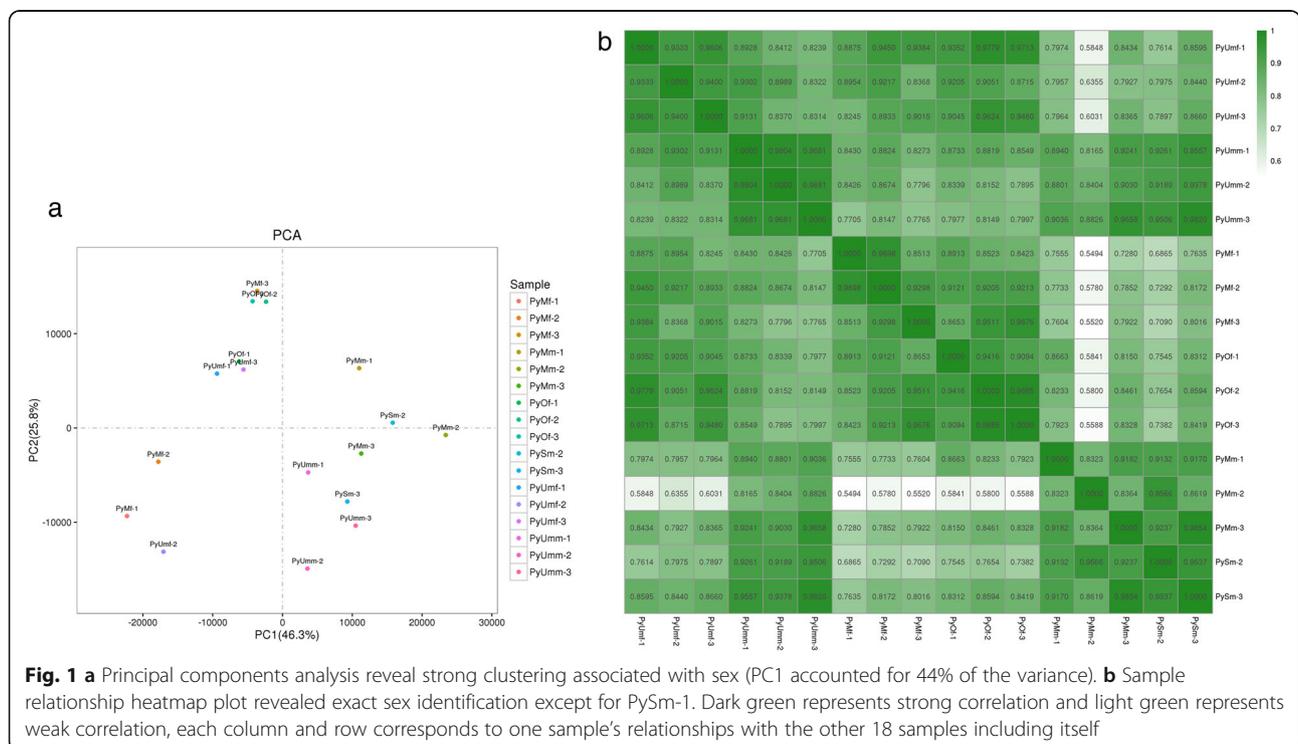
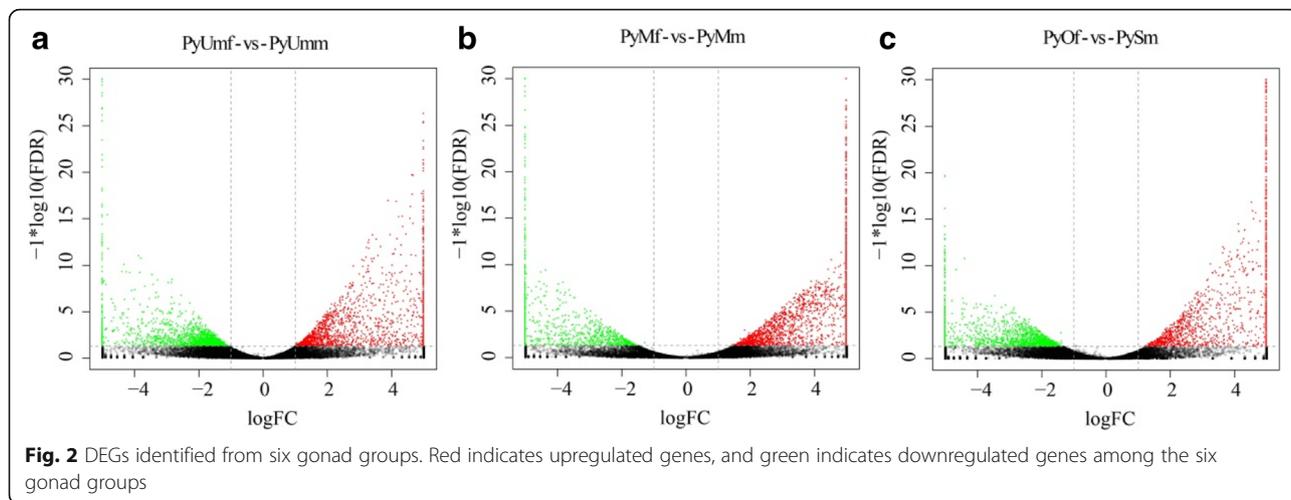
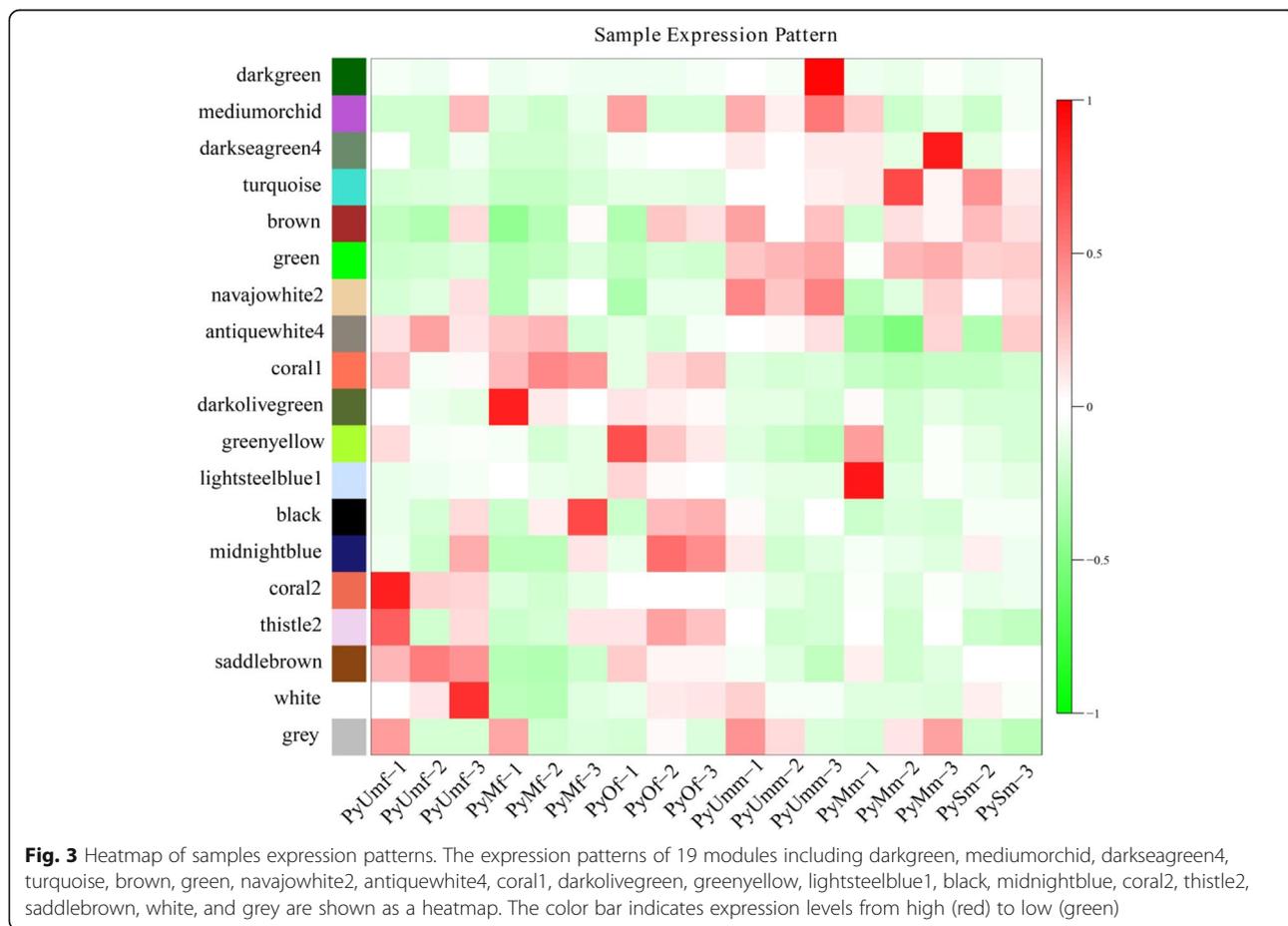


Fig. 1 a Principal components analysis reveal strong clustering associated with sex (PC1 accounted for 44% of the variance). b Sample relationship heatmap plot revealed exact sex identification except for PySm-1. Dark green represents strong correlation and light green represents weak correlation, each column and row corresponds to one sample's relationships with the other 18 samples including itself



and molecular functions) with $p < 0.05$ were significantly enriched in the coral1 module, 78 GO terms were enriched in the black module, 160 terms were significantly enriched in the turquoise module, and 237 were enriched in the green module. Most were enriched as cellular processes, metabolic processes, or single-

organism progresses, which can all be classified as biological processes. Lots were enriched as cells, cell parts, or organelles, which are classified under cellular components. Lots of terms were also enriched as binding or catalytic activities, which can be categorized as molecular functions. At least 49 genes in five modules were



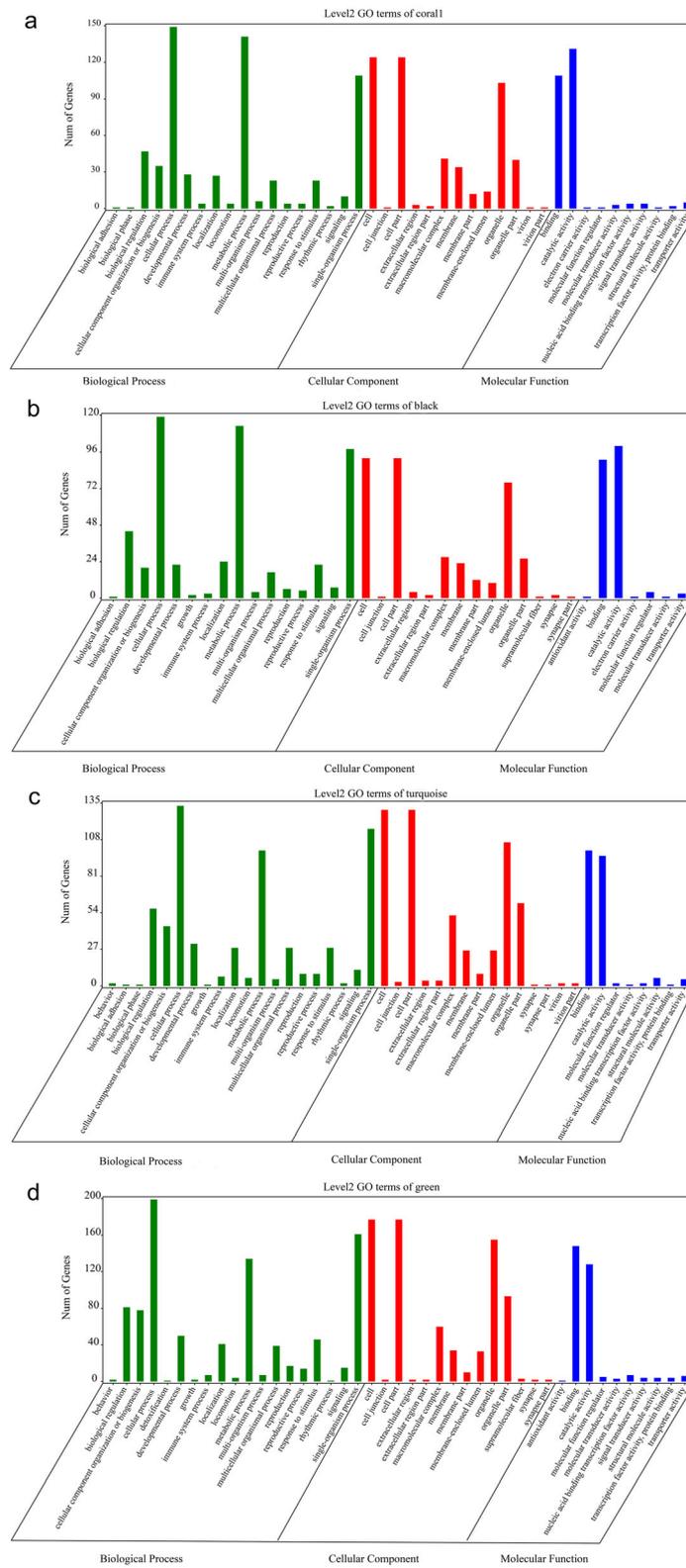


Fig. 4 Gene ontology term enrichment in four modules of RNA-Seq

identified as being involved in sex differentiation or determination (Additional file 2: Table S1), and most of these were enriched in the GO function terms mentioned above. In total, out of all of the transcripts, 5030 genes had KEGG assignments. These mainly belonged to metabolic processes, environmental information processing functions, and genetic information processing pathways. More specifically, ribosome biogenesis in eukaryotes, RNA transport, metabolism of xenobiotics by cytochrome P450, degradation of valine, leucine and isoleucine, aminoacyl-tRNA biosynthesis, and beta-Alanine metabolism were significantly enriched in females

(Fig. 5a, b), and purine metabolism, carbon metabolism, biosynthesis of amino acids, oxidative phosphorylation, the TCA cycle, and the fanconi anemia pathway were significantly enriched in males (Fig. 5c, d). All of these GO term enrichments and KEGG pathway enrichments were closely related to sex differentiation or determination. Some of them belonged to cellular process pathways, which are relevant to gamete development.

Quantitative real-time PCR

Fifteen sex-biased genes were randomly selected for qRT-PCR to validate differential expressions identified

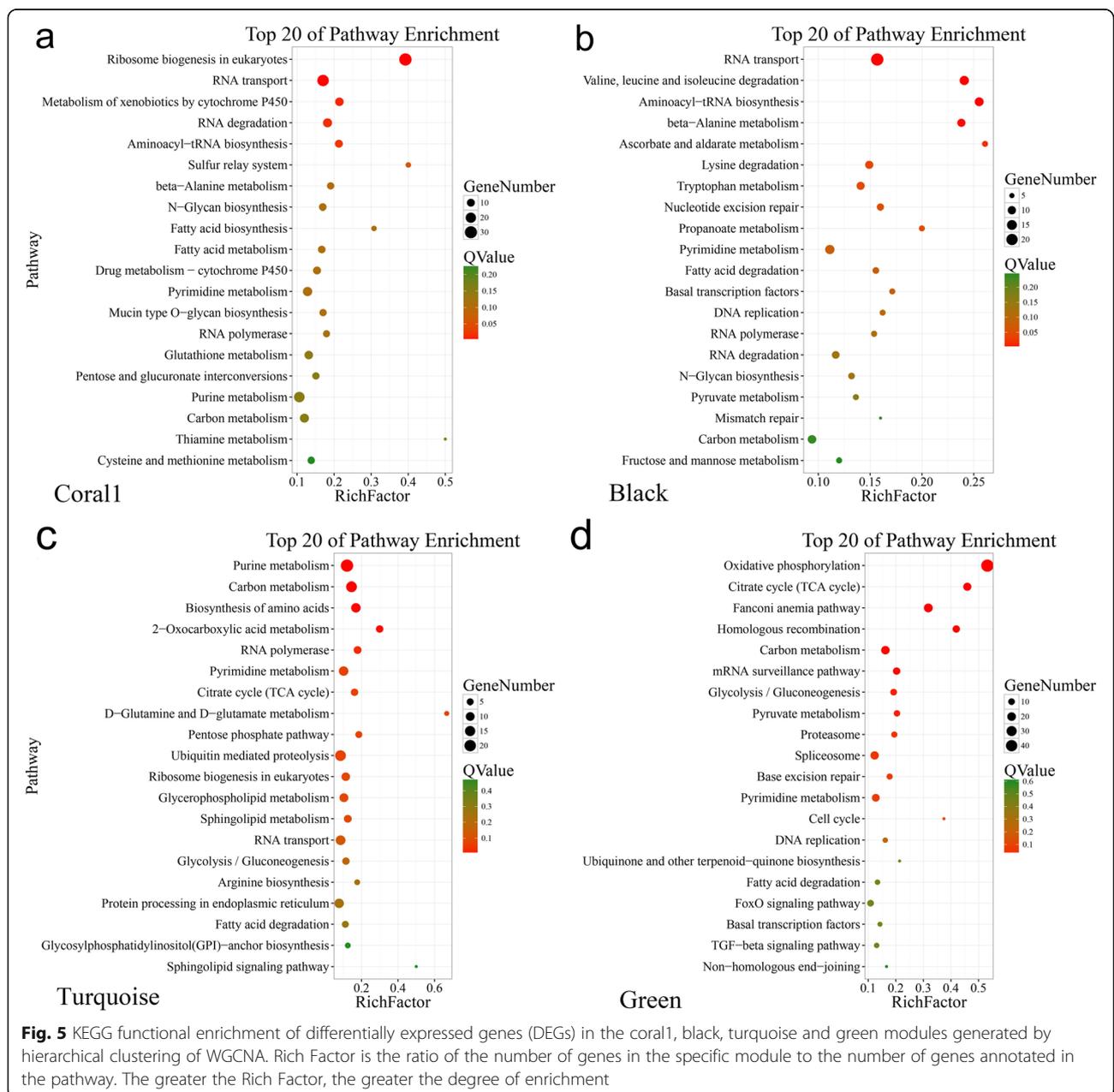
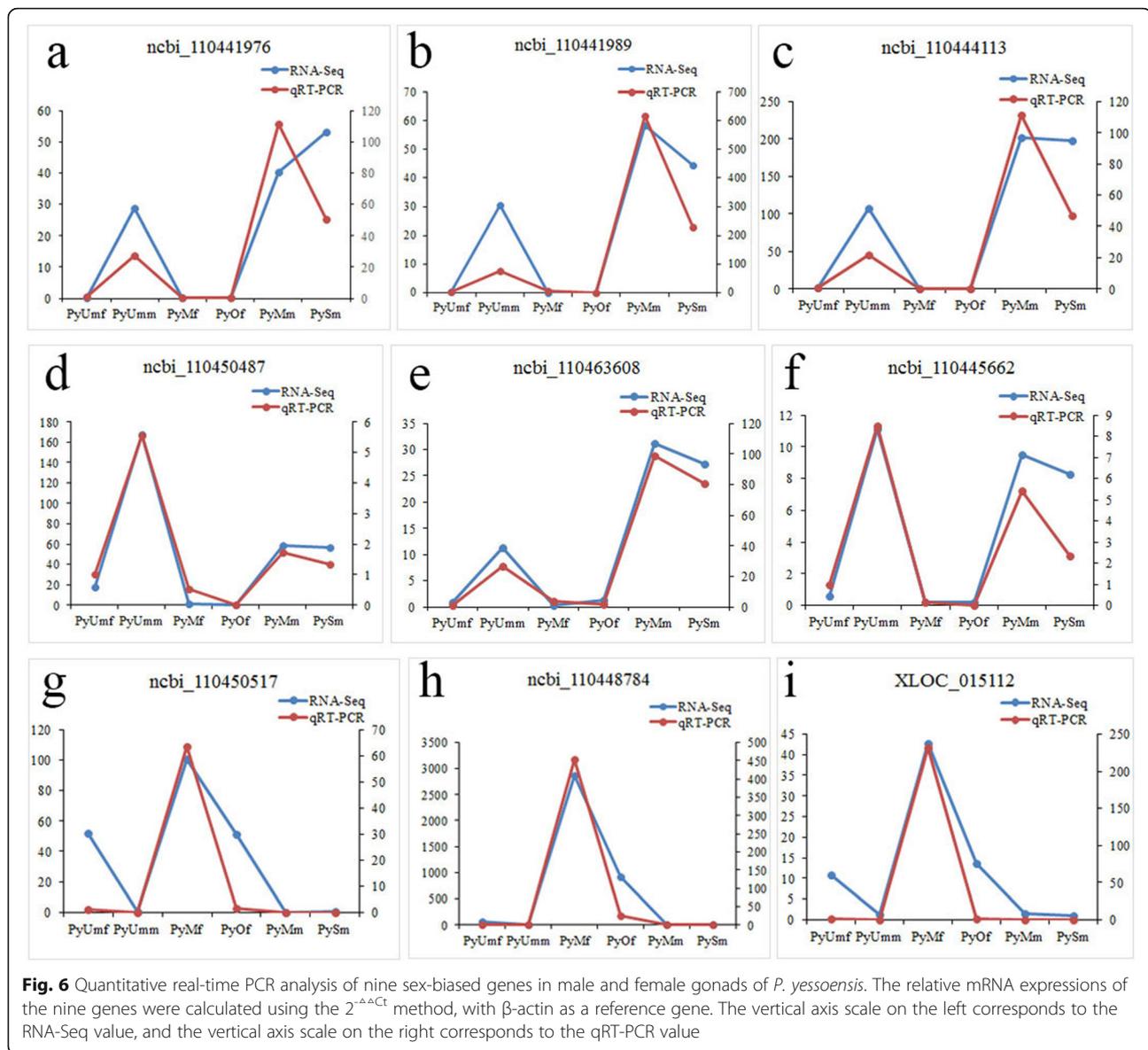


Fig. 5 KEGG functional enrichment of differentially expressed genes (DEGs) in the coral1, black, turquoise and green modules generated by hierarchical clustering of WGCNA. Rich Factor is the ratio of the number of genes in the specific module to the number of genes annotated in the pathway. The greater the Rich Factor, the greater the degree of enrichment



by RNA-Seq. Comparing the transcriptome data with the qRT-PCR results, we found that the patterns of transcript abundance detected for genes using qRT-PCR were consistent with the RNA-Seq results (Fig. 6). The expression pattern of ncbi_110450487 (*Dmrt 1*) (Fig. 6d) was different from the expression patterns of the male-biased genes shown in Figs. 6a, b, c, e, and f. Its expression was also remarkably different from the expressions of the female-biased genes shown in Figs. 6g, h, and i. Therefore, *Pydmrt 1* was found to be the only one gene which determined the male sex phenotype during early gonadal differentiation in darkgreen module.

Identification of sex-biased hub genes and network construction

In the turquoise module, the genes in the core positions were ncbi_110441989 (*cAMP* and *cAMP-inhibited cGMP 3',5'-cyclic phosphodiesterase 10A, PDE*), ncbi_110442454 (*testis-specific serine/threonine-protein kinase 3-like, tssk-3*), ncbi_110466991 (*WD repeat-containing protein on Y chromosome-like, WD rcp*), and ncbi_110444113 (*leucine-rich repeat-containing protein, LRR*) (Additional file 3: Figure S2). In the green module, ncbi_110443603 (*choline transporter-like protein 1, CTL1*), ncbi_110465229 (*actin 5C*), and ncbi_110441893 (*basic helix-loop-helix and HMG box*

domain-containing protein 1, bHLH) were in the core positions (Additional file 4: Figure S3). Other genes associated with these genes all had the same expression patterns, and were much more highly expressed in male gonads than in female gonads.

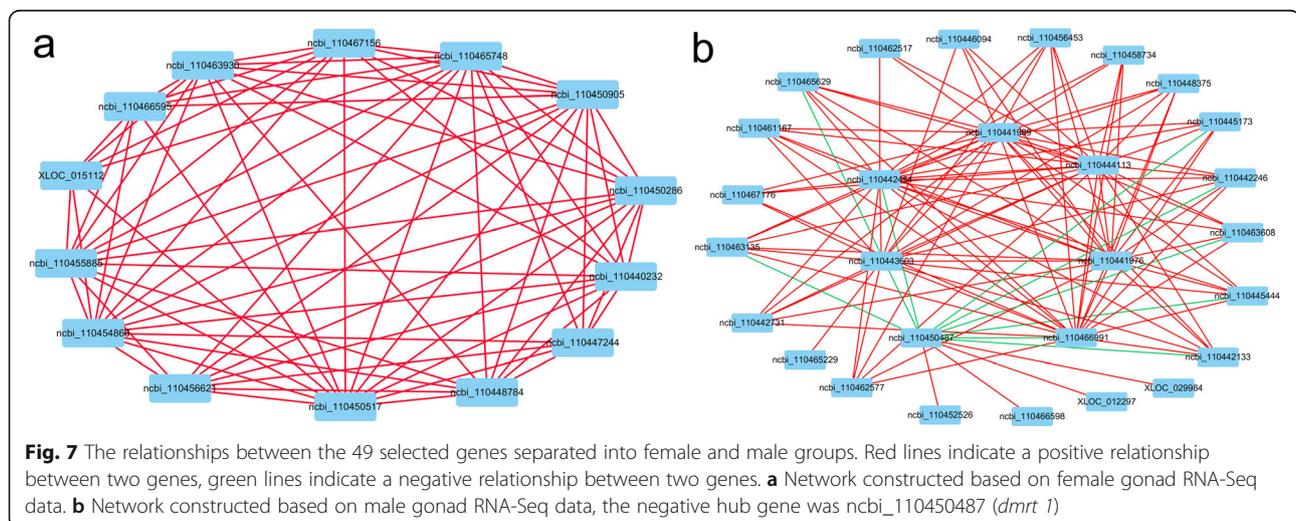
In the corall1 module, ncbi_110465748 (*polypeptide N-acetylgalactosaminyltransferase 4, GALNT4*), ncbi_110454866 (*protein ovo-like isoform X4*), ncbi_110450517 (Cytochrome P450 1A4-like, *CYP1A4-like*), ncbi_110450286 (*forkhead box protein A2-A-like, fox A2-like*), and XLOC_015112 (*transcriptional regulatory protein TBS1-like, TBS1-like*) were in the core positions (Additional file 5: Figure S4). In the black module, ncbi_110456621 (*glycoprotein 3-alpha-L-fucosyltransferase A, Fuc-T*), ncbi_110447244 (*collagen alpha-2(I) chain*), ncbi_110440232 (uncharacterized protein LOC105336037), and ncbi_110448784 (*Vitellogenin, Vg*) were in the core positions (Additional file 6: Figure S5). Other genes associated with these genes all had the same expression patterns, and were much more highly expressed in female gonads than in male gonads.

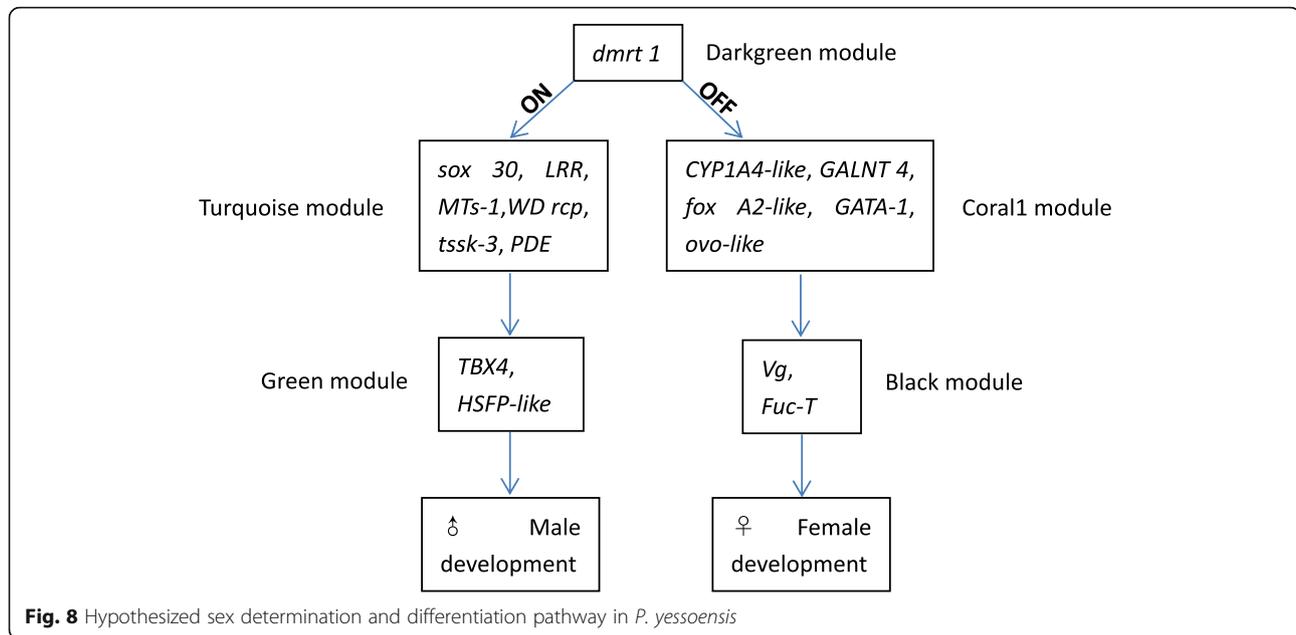
The 49 selected genes were used to construct networks based on their relationship coefficients of expression mode in female and male groups (Fig. 7). In the female group, ncbi_110450517 (*CYP1A4-like*), ncbi_110465748 (*GALNT4*) and ncbi_110454866 (*protein ovo-like*) are the hub genes, other genes, such as ncbi_110447244 (*collagen alpha-2(I) chain*), XLOC_015112 (*TBS1-like*) and ncbi_110456621 (*Fuc-T*) are positively related to these hub genes. In the male group, ncbi_11450487 (*dmrt 1*) was a negative hub gene, while ncbi_110441989 (*PDE*), ncbi_110444113 (*LRR*), ncbi_110441976 (*sox 30*), ncbi_110466991 (*WD rcp*), ncbi_110443603 (*CTL 1*) and ncbi_110442454 (*tssk*) were all positive hub genes, ncbi_110463608 (*RFX4*), ncbi_

110463135 (*IF-2-like*), ncbi_110442133 (*MARCH3*), ncbi_110442246 (*stabilizer of axonemal microtubules, MTs*), ncbi_110445444 (*Ras guanine nucleotide*) and ncbi_110445173 (*Biorientation of chromosomes in cell division protein 1*) were closely negatively related to ncbi_11450487 (*dmrt 1*), but positively related to other hub genes.

Hypothesized sex differentiation pathway in *Patinopecten yessoensis*

By analyzing the overall gene expression profiles of gonads, at least 49 genes in five modules involved in sex differentiation or determination were identified (Additional file 2: Table S1). This study found that *dmrt 1* was in the dark-green module, and that its expression was much higher in PyUmm than in other male stages, while very low or none in all female gonads. This expression trend was consistent both in q RT-PCR and de novo analysis (Fig. 6), which means that high expression of *Pydmrt 1* determines the male sex phenotype during early gonadal differentiation. Further studies should focus on validating the function of *dmrt 1*. Other selected target genes were identified as transcription factors (TFs), and they were enriched in males or females by comparing all the datasets, especially the correlation coefficients, expression levels, and gonad developmental stage. Therefore, we propose that *dmrt 1* may have a leading function in the sex differentiation pathway of *P. yessoensis*, and that significantly high expression of *dmrt 1* directly activates *sox 30* (*transcription factor SOX-30*), *LRR* (*leucine-rich repeat-containing protein*), *MTs*, *WD rcp* (*WD repeat-containing protein on Y chromosome*), *tssk-3* (*testis-specific serine/threonine-protein kinase 3*), and *cAMP and cAMP-inhibited cGMP 3',5'-cyclic phosphodiesterase (PDE)*, which were all male-specific genes in the turquoise module. *TBX4* (*T-box transcription factor 4*), *CTL 1* (*choline transporter-like protein 1*), *Protein B4*, and *HSFP-like* (*heat*





shock factor protein-like) were in the green module and were highly expressed genes. On the other hand, *CYP1 A4-like* (*Cytochrome P450 1A4-like*), *GALNT 4* (*polypeptide N-acetylgalactosaminyltransferase 4*), *fox A2-like* (*forkhead box protein A2-like*), *GATA-1*, and *ovo-like* were in the coral1 module and were female-specific genes. *Vg* (*Vitellogenin*) and *Fuc-T* (*glycoprotein 3-alpha-L-fucosyltransferase A*) were high expression genes in the black module. Therefore, we propose the following sex differentiation pathway in *P. yessoensis* (Fig. 8):

Discussion

By analyzing the overall gene expression profiles of scallop gonads, at least 49 genes in five modules involved in sex differentiation or determination were identified. These included *dmrt 1*, *sox 30*, *REFX 4*, *TBS1*, *Vg*, *fox E4* or *fox A2*, and other potential candidates, which have previously been reported in vertebrates and were assumed to be present in mollusks. Some of these genes were validated by qRT-PCR and can be considered highly relevant to sex differentiation or determination. Among these candidate genes, *dmrt 1* is documented as a well-known sex determination and differentiation gene in vertebrates [16, 17]. Naimi et al. found that a *dmrt 1* gene named *Cg-DM1* was expressed in female and male oyster gonads, but that its expression in the testis was significantly higher than in the ovaries during late gametogenesis [18]. Therefore, the study speculated that *Cg-DM1* played a role in sex determination and differentiation, which is consistent with our results and with the results of Yang et al. [6]. Testis-specific expression of *Ha-DMRT 1* and ovary-specific expression of *Ha-*

VTG1 have also been validated through semiquantitative RT-PCR as important genes in the gonad development of *Haliothis asinine* [19].

Eighteen male-specific genes from the turquoise module, 12 genes highly expressed in males from the green module, nine female-specific genes from the coral1 module, and five genes highly expressed in females from the black module were selected in our study. These genes were all previously reported to be related to sex or gonad development. The *sox* genes are SRY (sex-determining region on Y chromosome) boxes, and they encode a group of proteins with a conserved HMG (high mobility group) domain which can bind and bend DNA to open the testis determination pathway directly and close the ovary pathway indirectly in mammalian species [20]. Some members of the *forkhead box* family are transcription factors essential for the early regulation of ovarian development, and Liu et al. discovered that *Cf-foxl 2* is mainly expressed in the ovaries in a sexually dimorphic pattern at the RNA level using semiquantitative RT-PCR [21]. *Pyfoxl 2* has also been found to be ovary-biased, and *Pydmrt* and *Pysox H* have been classified as testis-biased [9]. This is consistent with our results that *fox E4* and *fox A2* were ovary-biased, and that *dmrt 1* and *Sox 30* were testis-biased. However (Table 1), *sox 9* and *foxl 2* were not found to be sex biased in our RNA-Seq data, and this is worthy of further research. Furthermore, Ghiselli et al. determined that *Sox 30* was a good candidate gene for the differentiation of developing male germ cells in the manila clam (*Ruditapes philippinarum*), and that its function was differentiation of developing male cells [22]. *Cg-sox E* mRNA expression is not restricted to one sex, one type of germ cell and two early stages, Santerre et al. concluded that *Cg-sox E* was a new potential gene in the

Table 1 Primers for quantitative real-time PCR and gene annotations

Gene ID	Annotation	Sequence 5' → 3'
ncbi_110441976	Transcription factor SOX-30	F GAACTTTGATGACGAGGCG R GCTGACATTGAAGAATCATCGGT
ncbi_110441989	cAMP and cAMP-inhibited cGMP 3',5'-cyclic phosphodiesterase 10A isoform X2	F GACTCCCAGCGGCATCTAA R CAAGGAACAGCGGTGAGCAT
ncbi_110444113	Leucine-rich repeat-containing protein 74B-like isoform X2	F GGTGTGTAAGAAGTTGGGTGTT R CGAACTTCCCAGACCGTG
ncbi_110450487	Doublesex- and mab-3-related transcription factor 1 isoform 1	F CCCCTAAATGTTGCTGCTGTC R TGAGGATAACGGTGTATGAGCA
ncbi_110463608	Transcription factor RFX4 isoform X3	F GTGATGACGGACCCCTGG R GGTTTACAATAGGGGCAGGC
ncbi_110445662	Protein B4	F GAGTCGCATCGTCTGTAAGGA R GCCGCTTTTCTACCCCTGA
ncbi_110450517	Cytochrome P450 1A4-like	F GCTTTACTTCTCGCCCAAT R AACAGTAGGTCTCCGATGATGG
ncbi_110448784	Vitellogenin	F GCCCTTCTATGCGACTCAAT R TATCCATTCTCTCAGGTAGTAG
XLOC_015112	Uncharacterized transcriptional regulatory protein TBS1-like	F TCACAGGGCAATGACGCAC R GCCCATCTACTGACCCTG
β-actin		F CCAAAGCCAACAGGGAAAAAG R TAGATGGGGACGGTGTGAGTG

sex-determining pathway of the Pacific oyster (*Crassostrea gigas*), a hermaphroditic Lophotrochozoan [23].

Shi et al. (2018) put forward a sex reversal/differentiation pathway for *Chlamys nobilis*. They deduced that the significantly high expressions of *foxl 2*, *β-catenin*, and *sry* in intersex individuals provided preliminary evidence for possible interactions among these male- and female-promoting genes [24]. As upstream genes in sex determination/differentiation, *Wnt 6-like* and *Wnt 11B* were highly expressed in the female immature stage gonads, and *β-catenin* was highly expressed in the male immature stage gonads. These genes may have threshold values in the early gonad development stage which serve as a valve to control downstream gene expression.

In terms of male-related genes, leucine-rich repeat (LRR) proteins often mediate protein–protein interactions, and act as a scaffold to link signaling molecules, including PP1, to the manchette near potential substrate proteins important for spermatogenesis [25]. Recent studies have shown that the ovaries constantly repress male-specific genes from embryonic stages of development to adulthood [26]. The degree of polyglutamylation or hyperglutamylation serves as stabilizer and must be tightly controlled for the doublet stability of *MTs* [27]. WD repeat-containing proteins on the Y chromosome were first reported in the white campion (*Silene latifolia*), which is one of the few plants with separate sexes and with X and Y sex chromosomes, as for human XY-linked genes, the sex-linked plant loci encoding WD-repeat proteins are likely to be involved in cell proliferation, they cease recombining at different

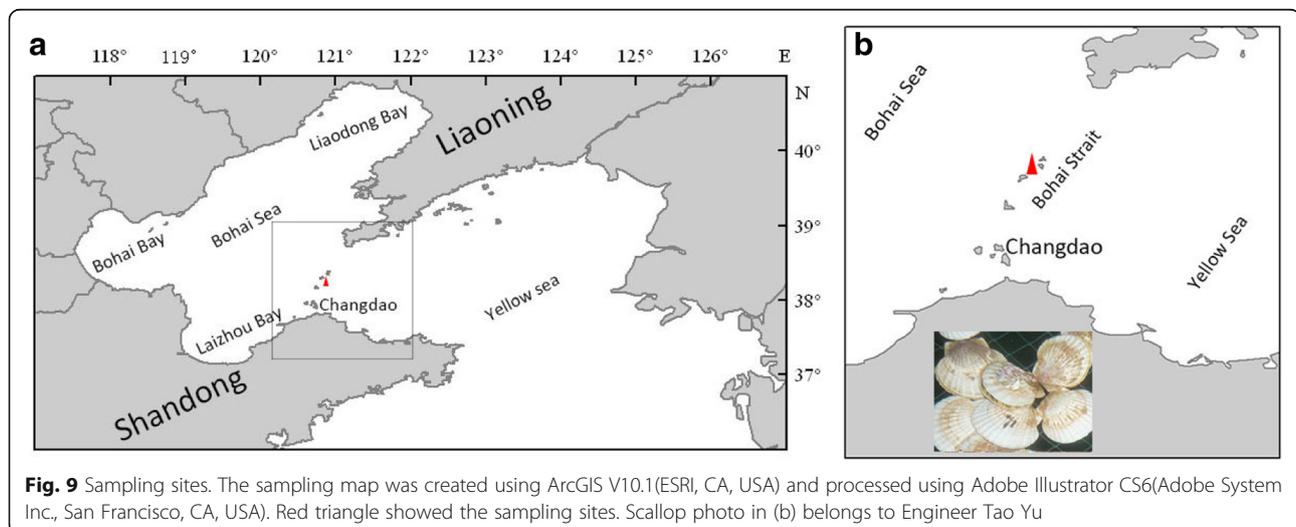
times and reveal distinct events in the evolutionary history of the sex chromosomes [28]. Expression of *tssk-3* is induced at puberty, persists during adulthood, and is restricted to the interstitial Leydig cells of mature male mice [29]. Giorgia et al. reported that cyclic nucleotides were involved in germ cell development and function, and discovered the presence of a calcium-calmodulin dependent PDE with high affinity for both cAMP and cGMP in male mouse sperm cells in addition to a calcium-calmodulin-independent PDA with high affinity for cAMP which controlled the sperm capacitation and motility [30]. *TBX4* and other clubfoot susceptibility genes increase predisposition to clubfoot in human males; variation in penetrance can be caused by their modifier(s) with sex-biased expression(s), resulting in different expression patterns in males versus females [31]. Yuan et al. reported that *mCTLI* mRNA was expressed in several mouse tissues such as the muscles, brain, heart, and testis [32]. Numerous heat shock proteins (HSP) and heat shock factors (HSF) are involved in a wide variety of physiological regulation processes and signaling pathways, exhibit a cell-type-specific expression pattern during spermatogenesis, and play crucial roles in germ cell development. The altered expression of HSP/HSFs may be responsible for abnormal germ cell apoptosis and subsequent impaired spermatogenesis [33]. All the above mentioned genes have been identified as male-specific genes.

Some female-specific genes were identified in our research, and were also observed and verified in other studies. CYP2AU1 signals were observed in follicular cells from

female gonads of all different gonadic stages, while in males only the spermatid follicle cells of the wall in the pre-spawning stage showed this signal. This result indicates that the cytochrome P450 gene product was involved in reproduction [34]. Meanwhile, many female-related genes were also reported in a large number of studies. The polypeptide N-acetylgalactosaminyltransferase-like protein 5 (GALNTL5) is involved in male fertility, it might play an important role in a range of reproductive processes as well as in Hu sheep sperm motility and capacitation [35]. However, the peptide derived from mouse testis associated spermatid protein (AWN) was not glycosylated by either isoform of ppGaNTase, these differences suggest that multiple forms of ppGaNTase are required to optimally O-glycosylate multi-site substrates [36]. We observed that *GALNT4* was a female-specific gene in the coral1 module in our investigation. Transthyretin (TTR) is a major thyroid hormone-binding protein in amphibian tadpoles whose plasma mRNA and protein levels are altered during metamorphosis. TTR transcripts are more abundant in males than females, and *TTR* is partially regulated by transcription factor FoxA2 in *Xenopus laevis* [37]. Interestingly, *fox A2* was a female-specific gene in *P. yessoensis* according to our observations. Female-predominant expression of the mouse gene *Cyp2b9* may be responsible for sexually dimorphic expression, and forkhead box A2 (FoxA2) (hepatic nuclear factor 3 β) had a large contribution to the promoter activity of *Cyp2b9* gene expression in both sexes, the differences in the expression of CYP2B9 mRNA between males and females might be regulated by FoxA2 protein, then sexually dimorphic secretion of growth hormone is involved in female predominant expression of these genes [38]. The *Drosophila ovo* gene encodes a putative transcription factor (Ovo) with TFIIIA-like zinc fingers. It is required for *Drosophila* female germ line maintenance and gametogenesis, and does not have a

function in males or in somatic tissues [39]. Both in mice and in humans, the gonadotropin hormones play fundamental roles in the male reproductive system. The sole targets for follicle-stimulating hormone (FSH) in males are the Sertoli cells, while *E2F* and *GATA-1* are required for the Sertoli cell-specific promoter activity of the follicle-stimulating hormone receptor gene [40]. According to our RNA-Seq data, *GATA-1* (*GATA-type zinc finger protein 1*), *Vg* (*Vitellogenin*), *Fuc-T* (*N-acetylglucosaminide 3- α -L-fucosyltransferase*), and *TBS1-like* were female-specific genes. As a nonpolar molecular carrier and a storage protein, *Vg* can combine and transfer lipids, proteins, vitamin, and carotenoids to oocytes during oogenesis, it also participates in the host immune defense in the noble scallop *Chlamys nobilis* [41]. GDP-fucose: N-acetylglucosaminide 3- α -L-fucosyltransferase (Fuc-T) activities were found to be expressed in the Chinese hamster ovary cells; it is involved in the biosynthetic pathway for the fucosylation of poly-lactosamine sequences in glycolipids and glycol-proteins [42]. However, the function of *TBS1* has not been discovered in any organism.

Despite the identification of some candidate genes, sex determination pathways in mollusks remain elusive. In our study, we proposed that *dmrt 1* may have a leading function in the sex differentiation pathway of *P. yessoensis*, because *dmrt 1* is an upstream regulator in vertebrates and was highly expressed in immature male scallop gonads. Its expression was different from genes in turquoise, green, coral1, and black modules. The results of a chromatin immunoprecipitation (ChIP) analysis and an electrophoretic mobility shift assay (EMSA) have demonstrated that the male sex-differentiation factor *dmrt 1* positively regulates the transcription of the Nile tilapia *sox 9b* gene [43], this is a good evidence for the validity of our results. *Dmrt 1* may directly or indirectly activate male- and female-related genes to determine the direction of sex differentiation.



Conclusion

WGCNA can facilitate identification of key genes for sex differentiation and determination. Using this method, 49 sex-related genes belonging to five modules were identified, and a hypothesized *P. yessoensis* sex determination and differentiation pathway was constructed. In this pathway, *Pydmrt1* may have a leading function.

Methods

Animal and tissue sampling

One- to 2-year-old Yesso scallops were collected monthly between January 2017 and July 2017 from Changdao, Shandong, China (Fig. 9). Gonad tissues were sampled from each scallop and were placed into liquid nitrogen for RNA extraction; a portion of each gonad was fixed for histological analysis. Other tissues were frozen and preserved separately in a -80°C freezer.

Biological traits such as shell length, shell height, shell width, shell color, body weight, shell weight, adductor muscle weight, and sex of each individual were recorded. Gonad development stage of the samples was identified and hermaphrodites were excluded using histological methods. In the immature stage (named PyUm), numerous oogonia or spermatogonia in the gonad follicles can be distinguished under a microscope through cellular morphology and size. In the mature stage (named PyM), gonads are completely filled with mature eggs or sperm cells. In the spawning stage (named PyO for females and PyS for males), numerous primary and mature gametes remain in the gonad follicles which become empty for spawning. Fifty-three animals were selected, and every two (or three or four) tissues from males or females were mixed into one sample. For convenience, every stage samples were assigned with the suffix “P” or “m” to distinguish samples from each sex. Thus, 18 samples were created and labelled as follows: PyUmf-1 to 3, PyUmm-1 to 3, PyMf-1 to 3, PyMm-1 to 3, PyOf-1 to 3, and PySm-1 to 3. Three biological repeats of male and female gonad tissues were used for RNA sequencing.

RNA-Seq sequencing, gene expression analysis, novel gene identification

Total RNA was extracted from 18 samples using Trizol reagent (Invitrogen, USA), and RNA integrity was assessed using the Agilent 2100 Bioanalyzer (Agilent Technologies, Palo Alto, CA, USA). Eighteen cDNA libraries were constructed using $3\ \mu\text{g}$ RNA (RIN >7) from each sample, following the conventional protocol. The prepared cDNA libraries were sequenced on Illumina HiSeqTM 4000 by Gene Denovo Biotechnology Co., Ltd. (Guangzhou, China).

Clean data were obtained after the adapters were removed from raw reads, and after reads with more than 10% unknown nucleotides and more than 50% low

quality bases (Q value <20) were removed. The clean reads of each sample were then mapped to reference genome using TopHat2 (version 2.0.3.12) [44]. The read numbers mapped to each gene were counted via RMSE software [45]. The gene expression level was normalized using the FPKM method, and the expected number of fragments per kilobase of transcript sequence per million base pairs sequenced was calculated based on the length of the gene and the read count mapped to this gene. Then, reconstruction of transcript assemblies was conducted using the reference annotation-based transcripts (RABT) assembly program within the Cufflinks software package to obtain a comprehensive set of transcripts for further downstream differential expression analysis.

To identify sex-related novel transcripts, the reconstruction transcripts were aligned to the *P. yessoensis* genome and were divided into 12 categories using Cuffcompare. Genes with classcode “u,” lengths ≥ 200 bp, and exon numbers ≥ 2 were defined as novel genes, and were further annotated through alignment with the Nr and KEGG databases.

Identification of differentially expressed genes (DEGs)

Differential expression analysis of groups among the six different gonadic categories was performed using the ‘edgeR’ package (<http://www.r-project.org/>). The resulting *P* values were adjusted to less than 0.05 to control the false discovery rate. Genes with $|\log_2(\text{FoldChange})| > 1$ and a false discovery rate-adjusted *P* < 0.05 were classified as differentially expressed genes. Differentially expressed gene (DEG) union was performed to assess the transcriptional pattern variations among the different gonadic categories, and a heat map of these DGEs was created using R scripts.

Relationship analysis of samples

The correlation coefficient between two replicas was calculated to evaluate repeatability between samples. Principal component analysis (PCA) was performed using the ‘gmodels’ package in R (<http://www.r-project.org/>), and sample relationship analysis was presented via a heatmap.

Weighted gene correlation network analysis (WGCNA)

To identify candidate genes and networks from sex differentiation DEGs, weighted gene correlation network analysis (WGCNA) was conducted to identify specific modules of co-expressed genes associated with each gonad development stage. We investigated the specific gene modules associated with the female and male gonad development stage separately. Prior to WGCNA, low-quality genes or samples (more than 50% were not expressed) were filtered out to improve the accuracy of the resulting network. To discover biologically or clinically significant modules, module eigengenes were used

to calculate correlation coefficients with samples or sample traits. Intramodular connectivity of each gene was calculated and genes with high connectivity tended to be hub genes which might have important functions. The networks were visualized using Cytoscape_3.3.0. For genes in each module, gene ontology (GO) and KEGG pathway enrichment analyses were conducted to analyze the biological functions of modules. Significantly enriched GO terms and pathways in genes in a module compared to background genes and pathways were defined by a hypergeometric test and a threshold of false discovery rate (FDR) of less than 0.05.

Forty-nine hub genes related to gonad development were selected to serve as key regulators connected to a large number of nodes to construct co-expression networks based on modules or sex group data. Based on sex group samples, the following selection conditions were set: $\text{cor} > 0.95$, $\text{cor} < -0.6$, and $p < 0.05$. Potential hub genes with sex differences in the transcript abundance were selected for real-time PCR.

Validation of RNA-Seq data through quantitative real-time PCR assays

To further validate the confidence of the high-throughput transcriptome sequencing and the genes that play important roles in sex differentiation or determination, 15 differentially expressed genes were selected and analyzed via qPCR. The qRT-PCR analysis was performed with three biological and three technical replicates. The RNA samples conformed to the required purity criteria (A260/A230 > 2.0, and A260/A280 of 1.8–2.0), and the integrity of the RNA samples was assessed by agarose gel electrophoresis. Then, 1 µg total RNA samples were reverse transcribed into cDNA using the PrimeScript™ RT reagent Kit with gDNA Eraser (Takara, Japan). Quantitative real-time PCR was performed using SYBR® Premix Ex Taq (Takara) according to the manufacturer's instructions on a Mx3000P (Agilent Stratagene, Agilent Technologies, Palo Alto, CA, USA) in 20-µl reactions. The PCR amplification procedure was carried out at 95 °C for 90 s, followed by 40 cycles at 95 °C for 5 s, 60 °C for 15 s, and 72 °C for 20 s; this was followed by dissociation curve analysis in an ABI 7500 fast real-time PCR system (Applied Biosystems, USA).

The *P. yessoensis* β-actin gene was used as an internal reference [6, 46]. The comparative Ct method ($2^{-\Delta\Delta C_t}$ method) was used to calculate the relative gene expressions of the samples, which were normalized using the β-actin mRNA level. The expression data were subsequently subjected to independent t-tests in SPSS 18.0 to determine whether there were any significant differences at the $P < 0.05$ level. Fifteen sex-biased genes were randomly selected to design primers based on the NCBI database, nine gene primer pairs created using these genes are listed in Table 1. The fold changes of these genes in female versus male gonads

were calculated via FPKM. The genes' log₂ fold change values of qRT-PCR and RNA-Seq were used for graphical presentation.

Additional files

Additional file 1: Figure S1. Venn diagram of sex-biased genes. (PNG, 28 kb). (PNG 27 kb)

Additional file 2: Table S1. Forty-nine representative genes related to sex determination and differentiation in the transcriptome of *P. yessoensis*. (WORD, 36 kb) (DOCX 35 kb)

Additional file 3: Figure S2. Male-related gene co-expression networks for turquoise module. Red squares represent the hub genes ncbi_110441989 (*PDE*), ncbi_110442454 (*tssk-3*), ncbi_110466991 (*WD rcp*), and ncbi_110444113 (*LRR*) in the turquoise module. (PDF, 11 kb) (PDF 10 kb)

Additional file 4: Figure S3. Male-related gene co-expression networks for green module. Red squares represent the hub genes ncbi_110443603 (*CTL1*), ncbi_110465229 (*actin 5C*), and ncbi_110441893 (*bHLH*) in the green module. (PDF, 14 kb) (PDF 13 kb)

Additional file 5: Figure S4. Female-related gene co-expression networks for coral1 module. Red squares represent the hub genes ncbi_110465748 (*GALNT4*), ncbi_110454866 (protein ovo-like isoform X4), ncbi_110450517 (*CYP1A4-like*), ncbi_110450286 (*fox A2-like*), and XLOC_015112 (*TBS1-like*) in the coral1 module. (PDF, 10 kb) (PDF 9 kb)

Additional file 6: Figure S5. Female-related gene co-expression networks for black module. Red squares represent the hub genes ncbi_110456621 (*Fuc-T*), ncbi_110447244 (*collagen alpha-2(I) chain*), ncbi_110440232 (uncharacterized protein LOC105336037), and ncbi_110448784 (*Vg*) in the black module. (PDF, 6 kb) (PDF 5 kb)

Abbreviations

bHLH: Basic helix-loop-helix and HMG box domain-containing protein; *Cg*: *Crassostrea gigas*; *CTL*: Choline transporter-like protein; *DEG*: Differentially expressed gene; *dmt 1*: *Dsx* and *mab-3* related transcription factor 1; *fox*: Forkhead box; *FPKM*: Fragments per kilobase of exon per million reads mapped; *Fuc-T*: Glycoprotein 3- α -L-fucosyltransferase; *GALNT*: Polypeptide *N*-acetylgalactosaminyltransferase; *GO*: Gene ontology; *KEGG*: Kyoto Encyclopedia of Genes and Genomes; *LRR*: Leucine-rich repeat; *Mf*: Mature females; *Mm*: Mature males; *MTs*: Stabilizer of axonemal microtubules; *Of*: Ovulation of females; *PCA*: Principal component analysis; *PDE*: *cAMP* and *cAMP*-inhibited *cGMP* 3',5'-cyclic phosphodiesterase; *Py*: *Patinopecten yessoensis*; *Sm*: Sperm emission of males; *sox*: *Sry*-type high mobility group box; *SRY*: Sex-determining region on Y chromosome; *TBS 1*: Transcriptional regulatory protein *TBS1*; *TBX*: *T-box* transcription factor; *Tfs*: Transcription factors; *tssk*: Testis-specific serine/threonine-protein kinase; *Umf*: Immature females; *Umm*: Immature males; *Vg*: Vitellogenin; *WD rcp*: *WD* repeat-containing protein on Y chromosome-like; *WGCNA*: Weighted gene co-expression network analysis

Acknowledgements

We are grateful to Professor Qingyin Wang for his advice on experimental design, and to Guangzhou Genedenovo Biotechnology Co., Ltd. for their assistance in sequencing and bioinformatic analyses.

Authors' contributions

LQZ, ZHL, and YHD conceived of and designed the experiments. LQZ, TY, YXZ, QZ, DZ, and AGY performed scallop tissue sampling and sex identification. XJS and BW conducted qPCR validation experiments. LQZ and DZ analyzed the data and wrote this paper. All authors read and approved the final manuscript.

Funding

In this study, the design of the study, the analysis, the interpretation of data and in writing the manuscript were supported in main by the National Natural Science Foundation of China (Grant no. 31672637), the sample collection and analysis were supported in part by the Zhejiang Provincial

Top Discipline of Bioengineering (Level A) of China (Grant no. KF2018008), the National Key R & D Program of China (2018YFD0900800), and the Qingdao People's livelihood Science and Technology Program of Shandong Province in China. (Grant no. 18-6-1-110-nsh).

Availability of data and materials

The raw reads produced in this study were deposited in the NCBI SRA with the accession number SRP 199818 under Bioproject PRJNA544564.

Ethics approval and consent to participate

In the present study, the collection and handling of all animals was approved by the Animal Care and Use Committee of the Chinese Academy of Fishery Sciences. In addition, we performed all experimental procedures in accordance with the guidelines outlined in the Care and Use of Laboratory Animals of the Chinese Academy of Fishery Sciences.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Key Laboratory for Sustainable Development of Marine Fisheries, Ministry of Agriculture, Yellow Sea Fisheries Research Institute, Chinese Academy of Fishery Science, Qingdao, China. ²Laboratory for Fisheries Science and Food Production Processes, Qingdao National Laboratory for Marine Science and Technology, Qingdao, China. ³Zhejiang Wanli University, Ningbo, China. ⁴Changdao Enhancement and Experiment Station, Chinese Academy of Fishery Science, Changdao, China. ⁵College of Fisheries and Life Science, Shanghai Ocean University, Shanghai, China.

Received: 21 May 2019 Accepted: 9 August 2019

Published online: 23 August 2019

References

- Guo XM, Luo YS. Chapter - scallops and scallop aquaculture in. *Dev Aquac Fish Sci*. 2016;40:937–52.
- Zhou LQ, Yang AG, Wang QY, Zheng YX, Liu ZH, Wu B, Sun XJ, Zheng LB. Histological research on the gonad of hermaphrodite *Patinopecten yessoensis* in breeding period. *Chinese High Technology Letters*. 2014;24(8):874–80.
- Xu KF, Li Q, Kong LF, Yu RH. A first-generation genetic map of the Japanese scallop *Patinopecten yessoensis*-based AFLP and microsatellite markers. *Aquac Res*. 2008;40:35–43.
- Liu WD, Bao XB, Song WT, Zhou ZC, He CB, Yu XJ. The construction of a preliminary genetic linkage map in the Japanese scallop *Mizuhopecten yessoensis*. *Hereditas* (Beijing). 2009;31(6):629–37.
- Zhou LQ, Yang AG, Liu ZH, Wang QY, Wu B, Sun XJ, Zheng LB, Zheng YX. Screening the sex-related AFLP molecular markers in *Patinopecten yessoensis* based on the genetic linkage map. *Progress in fishery sciences*. 2014;35(6):109–3.
- Yang D, Yin C, Chang YQ, Dou Y, Hao ZL, Ding J. Transcriptome analysis of male and female mature gonads of Japanese scallop *Patinopecten yessoensis*. *Gen Genomics*. 2016;38:1041–52.
- Hou R, Bao ZM, Wang S, Su HL, Li Y, Du HX, Hu JJ, Wang S, Hu XL. Transcriptome sequencing and *De Novo* analysis for yesso scallop (*Patinopecten yessoensis*) using 454 GS FLX. *PLoS One*. 2011;6(6):e21560.
- Teaniniuraitemoana V, Huvet A, Levy P, Klopp C, Lhuillier E, Gaertner-Mazouni N, Gueguen Y, Moullac GL. Gonad transcriptome analysis of pearl oyster *Pinctada margaritifera*: identification of potential sex differentiation and sex determining genes. *BMC Genomics*. 2014;15:491.
- Li YP, Zhang LL, Sun Y MX, Wang J, Li RJ, Zhang MW, Wang S, Hu XL, Bao ZM. Transcriptome sequencing and comparative analysis of ovary and testis identifies potential key sex-related genes and pathways in scallop *Patinopecten yessoensis*. *Mar Biotechnol*. 2016;18(4):453–65.
- Chen H, Xiao GQ, Chai XL, Lin XG, Fang J, Teng SS. Transcriptome analysis of sex-related genes in the blood clam *Tegillarca granosa*. *PLoS One*. 2017;12(9):e0184584.
- Langfelder P, Horvath S. WGCNA: an R package for weighted correlation network analysis. *BMC Bioinformatics*. 2008;9:559.
- Yue CY, Li Q, Yu H. Gonad transcriptome analysis of the Pacific Oyster *Crassostrea gigas* identifies potential genes regulating the sex determination and differentiation process. *Marine biotechnol*. 2018;20:206–19.
- Powell D, Elizur A, Knibb W. Sex-specific transcript expression in the hepatopancreas of the banana shrimp (*Fenneropenaeus merguensis*). *Hydrobiologia*. 2018;825(1):81–90.
- Wang N, Wang RK, Wang RQ, Chen SL. Transcriptomics analysis revealing candidate networks and genes for the body size sexual dimorphism of Chinese tongue sole (*Cynoglossus semilaevis*). *Funct Integr Genomics*. 2018;18(3):327–39.
- Yu H, Zhao XL, Li Q. Genome-wide identification and characterization of long intergenic noncoding RNAs and their potential association with larval development in the Pacific oyster. *Sci Rep*. 2016;6:20796.
- Nanda I, Kondo M, Hornung U, Asakawa S, Winkler C, Shimizu A, Shan Z, Haaf T, Shima A. A duplicated copy of DMRT1 in the sex-determining region of the Y chromosome of the medaka, *Oryzias latipes*. *Proc Natl Acad Sci*. 2002;99:11778–83.
- Guo Y, Cheng H, Huang X, Gao S, Yu H, Zhou R. Gene structure, multiple alternative splicing, and expression in gonads of zebrafish *dmrt1*. *Biochem Biophys Res Commun*. 2005;330:950–7.
- Naimi A, Martinez AS, Specq ML, Mrac A, Diss B, Mathieu M, Sourdain P. Identification and expression of a factor of the DM family in the oyster *Crassostrea gigas*. *Comp Biochem Physiol A Mol Integr Physiol*. 2009;152:189–96.
- Amparyup P, Klinbunga S, Jarayabhand P. Identification and expression analysis of sex-specific expression markers of Thai abalone *Haliotis asinina*, Linnaeus, 1758. *J Shellfish Res*. 2010;29(3):765–73.
- Jiang T, Hou CC, She ZY, Yang WX. The SOX gene family: function and regulation in testis determination and male fertility maintenance. *Mol Biol Rep*. 2013;40:2187–94.
- Liu XL, Zhang ZF, Shao MY, Liu JG, Muhammad F. Sexually dimorphic expression of foxl2 during gametogenesis in scallop *Chlamys farreri*, conserved with vertebrates. *Dev Genes Evol*. 2012;222:279–86.
- Ghiselli F, Milani L, Chang PL, Hedgecock D, Davis JP, Nuzhdan SV, Passamonti M. De novo assembly of the manila clam *Ruditapes philippinarum* transcriptome provides new insight into expression bias, mitochondrial doubly uniparental inheritance and sex determination. *Mol Biol Evol*. 2012;29(2):771–86.
- Santerre C, Sourdain P, Adeline B, Martinez AS. Cg-SoxE and cg-β-catenin, two new potential actors of the sex-determining pathway in a hermaphrodite lophotrochozoan, the Pacific oyster *Crassostrea gigas*. *Comp Biochem Physiol A Mol Integr Physiol*. 2014;167:68–76.
- Shi Y, Liu WG, He MX. Proteome and transcriptome analysis of ovary, intersex gonads, and testis reveals potential key sex reversal/differentiation genes and mechanism in scallop *Chlamys nobilis*. *Mar Biotechnol*. 2018;20:220–45.
- Wang R, Kaul A, Sperry AO. TLRR (Irrc67) interacts with PP1 and is associated with a cytoskeletal complex in the testis. *Biol Cell*. 2010;102:173–89.
- Veitia RA. FOXL2 versus SOX9: a lifelong "battle of the sexes". *Bioessays*. 2010;32:375–80.
- O'Hagan R, Piasecki BP, Silva M, Pirke P, Nguyen KCQ, Hall DH, Swoboda P, Barr MM. The tubulin deglutamylase CCPP-1 regulates the function and stability of sensory cilia in *C. elegans*. *Curr Biol*. 2011;21:1685–94.
- Atanassov I, Delichère C, Filatov DA, Charlesworth D, Negrutiu I, Monéger F. Analysis and evolution of two functional Y-linked loci in a plant sex chromosome system. *Mol Biol Evol*. 2001;18(12):2162–8.
- Zuercher G, Rohrbach V, Andres AC, Ziemiecki A. A novel member of the testis specific serine kinase family, tskk-3, expressed in the Leydig cells of sexually mature mice. *Mech Dev*. 2000;93:175–7.
- Giorgi M, Piscitelli D, Rossi P, Geremia R. Purification and characterization of a low - gm 3' :5'-cyclic adenosine phosphodiesterase from post-meiotic male mouse germ cells. *Biochim Biophys Acta*. 1992;1121:178–82.
- Peterson JF, Ghaloul-Gonzalez L, Madan-Khetarpal S, Hartman J, Surti U, Rajkovic A, Yatsenko SA. Familial microduplication of 17q23.1–q23.2 involving TBX4 is associated with congenital clubfoot and reduced penetrance in females. *Am J Med Genet Part A*. 2014;164A:364–9.
- Yuan ZF, Wagner L, Poloumienko A, Bakovic M. Identification and expression of a mouse muscle-specific CTL1 gene. *Gene*. 2004;341:305–12.
- Ji ZL, Duan YG, Mou LS, Allam JP, Haidl G, Cai ZM. Association of heat shock proteins, heat shock factors and male infertility. *Asian Pac J Reprod*. 2012;1(1):76–84.

34. Reis IMM, Mattos JJ, Garcez RC, Zacchi FL, Miguelão T, Flores-Nunes F, Toledo-Silva G, Sasaki ST, Taniguchi S, Bicego MC, Cargnin-Ferreira E, Bairy ACD. Histological responses and localization of the cytochrome P450 (CYP2AU1) in *Crassostrea brasiliiana* exposed to phenanthrene. *Aquat Toxicol*. 2015;169:79–89.
35. Yao XL, MA El-S, Feng X, Zhang TT, Li FZ, Zhang GM, Pang J, Nie HT, Wang F. Expression and localization of polypeptide N-acetylgalactosaminyltransferase-like protein 5 in the reproductive organs and sperm of Hu sheep. *Anim Reprod Sci*. 2017;187:159–66.
36. Zara J, Hagen FK, Hagen KGT, Wuyckhuysen BC, Tabak LA. Cloning and expression of mouse UDP-GalNAc: polypeptide N-Acetylgalactosaminyltransferase-T3. *Biochem Biophys Res Commun*. 1996;228:38–44.
37. Ishihara A, Nishiyama N, Yu M, Yamauchi K. The genomic structure and the expression profile of the *Xenopus laevis* transthyretin gene. *Gene*. 2012;510:126–32.
38. Hashita T, Sakuma T, Akada M, Nakajima A, Yamahara H, Ito S, Takesako H, Nemoto N. Forkhead box A2-mediated regulation of female-predominant expression of the mouse Cyp2b9 gene. *Drug Metab Dispos*. 2008;36(6):1080–7.
39. Mével-Ninio M, Terracol R, Kafatos FC. The ovo gene of *Drosophila* encodes a zinc finger protein required for female germ line development. *EMBO J*. 1991;10(8):2259–66.
40. Kim JS, Griswold MD. E2F and GATA-1 are required for the sertoli cell-specific promoter activity of the follicle-stimulating hormone receptor gene. *J Androl*. 2001;22(4):629–39.
41. Zheng HP, Zhang Q, Liu HL, Liu WH, Sun ZW, Li SK, Zhang T. Cloning and expression of vitellogenin (vg) gene and its correlations with total carotenoids content and total antioxidant capacity in noble scallop *Chlamys nobilis* (bivalve: Pectinidae). *Aquaculture*. 2012;366-367:46–53.
42. Howard DR, Fukuda M, Fukuda MN, Stanley P. The GDP-fucose: N-Acetylglucosaminide 3- α -L-Fucosyltransferases of LEC11 and LEC12 Chinese hamster ovary mutants exhibit novel specificities for glycolipid substrates. *J Biol Chem*. 1987;262(35):16830–7.
43. Wei L, Li XY, Li MH, Tang YH, Wei J, Wang DS. Dmrt1 directly regulates the transcription of the testis-biased Sox9b gene in Nile tilapia (*Oreochromis niloticus*). *Gene*. 2019;687:109–15.
44. Kim D, Pertege G, Trapnell C, Pimentel H, Kelley R, Salzberg SL. TopHat2: accurate alignment of transcriptomes in the presence of insertions, deletions and gene fusions. *Genome Biol*. 2013;14(4):R36.
45. Li B, Dewey CN. RSEM: accurate transcript quantification from RNA-Seq data with or without a reference genome. *BMC bioinformatics*. 2011;12(1):323.
46. Sun XJ, Liu ZH, Wu B, Zhou LQ, Wang Q, Wu W, Yang AG. Differences between fast and slow muscles in scallops revealed through proteomics and transcriptomics. *BMC Genomics*. 2018;19:377.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

